

University of Groningen

Interleukin-18 Inhibition in Inflammatory Bowel Disease

Geertsema, Sem; Fagundes, Raphael R.; Otten, Antonius T.; Dijkstra, Gerard; Faber, Klaas Nico; Bourgonje, Arno R.

Published in:
Inflammatory Bowel Diseases

DOI:
[10.1093/ibd/izae042](https://doi.org/10.1093/ibd/izae042)

IMPORTANT NOTE: You are advised to consult the publisher's version (publisher's PDF) if you wish to cite from it. Please check the document version below.

Document Version
Publisher's PDF, also known as Version of record

Publication date:
2024

[Link to publication in University of Groningen/UMCG research database](#)

Citation for published version (APA):
Geertsema, S., Fagundes, R. R., Otten, A. T., Dijkstra, G., Faber, K. N., & Bourgonje, A. R. (2024). Interleukin-18 Inhibition in Inflammatory Bowel Disease: A Delicate Balance. *Inflammatory Bowel Diseases*, 30(4), 693–694. <https://doi.org/10.1093/ibd/izae042>

Copyright

Other than for strictly personal use, it is not permitted to download or to forward/distribute the text or part of it without the consent of the author(s) and/or copyright holder(s), unless the work is under an open content license (like Creative Commons).

The publication may also be distributed here under the terms of Article 25fa of the Dutch Copyright Act, indicated by the "Taverne" license. More information can be found on the University of Groningen website: <https://www.rug.nl/library/open-access/self-archiving-pure/taverne-amendment>.


Take-down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

Downloaded from the University of Groningen/UMCG research database (Pure): <http://www.rug.nl/research/portal>. For technical reasons the number of authors shown on this cover page is limited to 10 maximum.

Letter to the Editor

Interleukin-18 Inhibition in Inflammatory Bowel Disease: A Delicate Balance

Sem Geertsema, MD,* Raphael R. Fagundes, PhD,[†] Antonius T. Otten, MD,*
Gerard Dijkstra, MD, PhD,* Klaas Nico Faber, PhD,* and Arno R. Bourgonje, MD, PhD*[‡], 

*Department of Gastroenterology and Hepatology, University Medical Center Groningen, University of Groningen, Groningen, the Netherlands

[†]Department of Cell and Chemical Biology, Leiden University Medical Center, Leiden, the Netherlands

[‡]Henry D. Janowitz Division of Gastroenterology, Department of Medicine, Icahn School of Medicine at Mount Sinai, New York, NY, United States

Address correspondence to: Arno R. Bourgonje, MD, PhD, Department of Gastroenterology and Hepatology, University of Groningen, University Medical Center Groningen, Hanzeplein 1, 9713 GZ, Groningen, the Netherlands (a.r.bourgonje@umcg.nl).

To the Editors:

We read with great interest the study by Ikegami et al,¹ which addresses the inhibition of mature interleukin (IL)-18 in dextran sodium sulfate–induced mice models using a monoclonal antibody targeting a neoepitope of caspase-cleaved mature IL-18. The study analyzed several outcome parameters including inflammatory cytokines, intestinal epithelial permeability assays, goblet cell function, and gut microbiota alterations. The authors provide compelling data of beneficial effects of IL-18 inhibition using monoclonal antibodies to target mature IL-18, which might become novel therapeutic agents for patients with inflammatory bowel disease. However, the authors also aptly conclude that the precise mechanisms by which IL-18 regulates tight junction integrity and mucus production remain ambiguous. Here, we would like to emphasize and expand on the delicate balance between pro- and anti-inflammatory functions that IL-18 seems to fulfill within the intestinal mucosa.

Previous studies using murine models revealed anti-inflammatory and immune-protective properties of IL-18,^{2,3} while others presented evidence supporting proinflammatory IL-18 signaling^{4,5} in chemically induced intestinal inflammation. These diverging observations may arise from a disruption in the communication between gut microbiota, their metabolites, and inflammasome-mediated IL-18 production. For instance, bacteria-produced short-chain fatty acids can activate the inflammasome, thereby promoting IL-18 maturation and eliciting barrier-protective and immunoregulatory effects.^{6,7} Our recent study proposed that IL-18 regulates intestinal homeostasis through activation of the HIF-1 α (hypoxia-inducible factor 1 α) pathway, facilitated by commensal short-chain fatty acid–producing bacteria (eg, *Faecalibacterium prausnitzii*) in a large inflammatory bowel disease patient cohort.⁸ Moreover, taurine—a bile acid

conjugate—has also been suggested to positively modulate inflammasome activation.^{3,9}

Collectively, IL-18 seems to have a dual role in colitis: while it contributes to the maintenance of barrier integrity in homeostasis, during inflammation it may become implicated in goblet cell dysfunction and disrupted barrier integrity. The study by Ikegami et al offers compelling data of beneficial effects of IL-18 inhibition that are mainly targeted at the site of the intestinal epithelium. However, the nature of the effects of IL-18 inhibition may depend on the gut microbiota composition (eg, differing between homeostatic and dysbiotic microbiomes). Therefore, it remains challenging to discern the potential benefits and risks of therapeutically modulating this equilibrium. The specific cellular effects and interrelationships between IL-18, gut microbiota–derived metabolites, and mucosal immunity remain incompletely defined.¹⁰ Finally, a temporal divergence may also exist, with anti-inflammatory effects of IL-18 observed in early stages of colitis-induced inflammation, while proinflammatory effects may manifest at later disease stages.⁵

Funding

None received.

Conflicts of Interest

None declared.

References

1. Ikegami S, Maeda K, Urano T, et al. Monoclonal antibody against mature interleukin-18 ameliorates colitis in mice and improves epithelial barrier function. *Inflamm Bowel Dis*. 2023; doi:[10.1093/ibd/izad292](https://doi.org/10.1093/ibd/izad292). [Online ahead of print].

2. Harrison OJ, Srinivasan N, Pott J, et al. Epithelial-derived IL-18 regulates Th17 cell differentiation and Foxp3⁺ Treg cell function in the intestine. *Mucosal Immunol.* 2015;8(6):1226-1236.
3. Levy M, Thaïss CA, Zeevi D, et al. Microbiota-modulated metabolites shape the intestinal microenvironment by regulating NLRP6 inflammasome signaling. *Cell.* 2015;163(6):1428-1443.
4. Nowarski R, Jackson R, Gagliani N, et al. Epithelial IL-18 equilibrium controls barrier function in colitis. *Cell.* 2015;163(6):1444-1456.
5. Pu Z, Che Y, Zhang W, et al. Dual roles of IL-18 in colitis through regulation of the function and quantity of goblet cells. *Int J Mol Med.* 2019;43(6):2291-2302.
6. Macia L, Tan J, Vieira AT, et al. Metabolite-sensing receptors GPR43 and GPR109A facilitate dietary fibre-induced gut homeostasis through regulation of the inflammasome. *Nat Commun.* 2015;6:6734.
7. Arpaia N, Rudensky AY. Microbial metabolites control gut inflammatory responses. *Proc Natl Acad Sci U S A.* 2014;111(6):2058-2059.
8. Fagundes RR, Bravo-Ruiseco G, Hu S, et al. Faecalibacterium prausnitzii promotes intestinal epithelial IL-18 production through activation of the HIF1 α pathway. *Front Microbiol.* 2023;14:1298304.
9. van Eijk LE, Offringa AK, Bernal ME, Bourgonje AR, van Goor H, Hillebrands J-L. The disease-modifying role of taurine and its therapeutic potential in coronavirus disease 2019 (COVID-19). *Adv Exp Med Biol.* 2022;1370:3-21.
10. Hand TW. Interleukin-18: the bouncer at the mucosal bar. *Cell.* 2015;163(6):1310-1312.