

University of Groningen

Intestinal bile acid reabsorption in health and disease

van de Peppel, Ivo Pieter

IMPORTANT NOTE: You are advised to consult the publisher's version (publisher's PDF) if you wish to cite from it. Please check the document version below.

Document Version

Publisher's PDF, also known as Version of record

Publication date:

2019

[Link to publication in University of Groningen/UMCG research database](#)

Citation for published version (APA):

van de Peppel, I. P. (2019). *Intestinal bile acid reabsorption in health and disease*. [Thesis fully internal (DIV), University of Groningen]. Rijksuniversiteit Groningen.

Copyright

Other than for strictly personal use, it is not permitted to download or to forward/distribute the text or part of it without the consent of the author(s) and/or copyright holder(s), unless the work is under an open content license (like Creative Commons).

The publication may also be distributed here under the terms of Article 25fa of the Dutch Copyright Act, indicated by the "Taverne" license. More information can be found on the University of Groningen website: <https://www.rug.nl/library/open-access/self-archiving-pure/taverne-amendment>.

Take-down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

Downloaded from the University of Groningen/UMCG research database (Pure): <http://www.rug.nl/research/portal>. For technical reasons the number of authors shown on this cover page is limited to 10 maximum.

CHAPTER

3

Bile acid homeostasis in gastrointestinal and metabolic complications of cystic fibrosis

Ivo P. van de Peppel
Frank A. J. A. Bodewes
Henkjan J. Verkade
Johan W. Jonker

Journal of Cystic Fibrosis 2018, Aug 21

Abstract

With the improved treatment of the pulmonary complications of cystic fibrosis (CF), gastrointestinal problems have become more important in the morbidity in CF. A hallmark of the gastrointestinal phenotype of CF, apart from pancreatic insufficiency, is a disruption of bile acid homeostasis. Bile acid homeostasis is important for many gastrointestinal processes including fat absorption, inflammation, microbial composition, as well as regulation of whole body energy metabolism. This review describes the impairment of bile acid homeostasis in CF, its possible consequences for gastrointestinal and metabolic complications and its potential as a target for therapy.

Introduction

Cystic fibrosis (CF), caused by a mutation in the gene encoding the CF transmembrane conductance regulator (CFTR), results in production of abnormally thick, viscous mucus in various organ systems (1). Aside from pulmonary problems, CF patients often suffer from GI disorders, hepatobiliary problems and cystic fibrosis related diabetes (CFRD). A specific GI feature common among CF patients is impaired bile acid (BA) homeostasis manifesting via BA malabsorption and subsequent increased fecal excretion (2). Recent studies show the interrelation of BA homeostasis with various other intestinal, hepatic and metabolic parameters. In this review we discuss the role of impaired BA homeostasis in CF, explaining its potential role in other GI and metabolic complications and as a therapeutic target.

Gastrointestinal complications of cystic fibrosis

A functional GI system is essential for maintaining adequate nutritional status and whole body homeostasis. Similar to pulmonary complications, viscous mucus, as a consequence of deficient surface fluid and bicarbonate flux is an important underlying factor in the GI phenotype of CF (3). Exocrine pancreatic insufficiency (EPI) is used as a marker for severity of the CF phenotype. However, other manifestations of the GI phenotype of CF are often highly variable and do not strongly correlated to allelic CFTR variation (4). Patients experience various symptoms including malabsorption, fatty stools (steatorrhea), abdominal pain, nausea, anorexia, bloating, gastro-esophageal reflux, constipation, distal intestinal obstruction syndrome (DIOS) and flatulence. Although most of the GI complications are interrelated, they can be subdivided in pancreatic, hepatobiliary and intestinal-luminal categories.

A severe *CFTR* gene mutation in both alleles results in little or no CFTR Cl⁻ channel activity and destruction of the exocrine pancreas (5). EPI is an early sign of CF and can present at birth or develop in the first months of life (6). Ultimately, around 85% of CF patients develop EPI and these patients are prone to nutritional deficiencies, severe malnutrition and growth retardation (7). Fortunately, EPI can be successfully treated with pancreatic enzyme replacement therapy (PERT). However, even with optimal PERT, fat malabsorption and GI complaints are often not fully corrected (8–10). Mice with targeted mutations in the *Cftr* gene do not display EPI but nevertheless have a lower bodyweight upon *ad libitum* feeding, possibly due to bacterial overgrowth or from impaired epithelial absorption of

nutrients (11). This suggests that, in addition to EPI, there are other changes in the intestinal tract in CF that have important effects on nutrient absorption and growth.

CF patients can suffer from a multitude of hepatobiliary problems including gall stones, hepatitis, steatosis and cirrhosis. Hepatobiliary problems are common in pediatric CF patients with reported prevalence rates up to 25% (12,13). Cystic fibrosis related liver disease (CFLD) was thought to develop mainly in early childhood. However, a recent follow-up study of a cohort of CF patients into adulthood incorporated novel markers into the CFLD diagnostic algorithm and suggests an additional wave of adult-onset CFLD with a median age of 37 years (14). Another recent study that assessed a large retrospective cohort of French CF patients found that CFLD incidence increased by approximately 1% every year reaching 32.2% by the age of 25 (13).

In the liver, CFTR is exclusively expressed at the apical membrane of cholangiocytes lining the bile ducts (15). CFLD is characterized by focal biliary cirrhosis which can lead to multilobular cirrhosis and portal hypertension in 1-10% of patients (16). The pathophysiology of biliary cirrhosis has been hypothesized to be secondary to occlusion of small bile ducts and/or to increased bile toxicity. In CF mouse models, however, evidence to support the hypothesis that increased bile toxicity contributes to CFLD has not been reported (17).

Luminal GI complications are highly prevalent in CF. Approximately 15-20% of CF infants present with meconium ileus, an obstruction of the distal small intestine by dehydrated mucofeculent material (18). After the neonatal phase, acute fecal obstruction of the ileocecum known as DIOS can occur and incidence increases with age (19,20). Nearly half of pediatric CF patients suffer from constipation and this is even more prevalent in adulthood (21). Another common luminal GI feature of CF is a change in intestinal microbiota characterized by small intestinal bacterial overgrowth (SIBO) and colonic dysbiosis (22). Important contributing factors include delayed intestinal transit time, luminal hyperacidity due to decreased bicarbonate secretion by pancreas and intestinal epithelium, frequent antibiotic use and inspissated mucus. Intestinal microbial composition is important for immune function and various metabolic processes in the body (23). Its disruption in CF is therefore likely to contribute to various aspects of the phenotype (24).

Along with the increased life expectancy the CF population has been shown to become exposed to an increased risk of malignant tumors especially of the small intestine and colon (22,25,26), possibly due to an increased proliferation rate of epithelial cells and disruption of anti-apoptotic pathways (27). Additionally, a recent study has shown a direct role of CFTR as a tumor suppressor gene in intestinal

cancer (28). After lung transplantation the risk for malignancies in CF patients is even further increased due to the use of immunosuppressant drugs (25,29).

Impaired bile acid homeostasis and farnesoid X receptor signaling in cystic fibrosis

One of the hallmarks of the GI complications in CF patients as well as in murine CF models is an up to 3-fold increase in fecal bile acid (BA) excretion (2,30–32). This increase is independent of exocrine pancreatic insufficiency and fat malabsorption. In the physiological situation the enterohepatic circulation of BAs is a tightly regulated system in which ~95% of total BAs are reabsorbed and the remaining ~5% is excreted via the feces (**Fig. 1**).

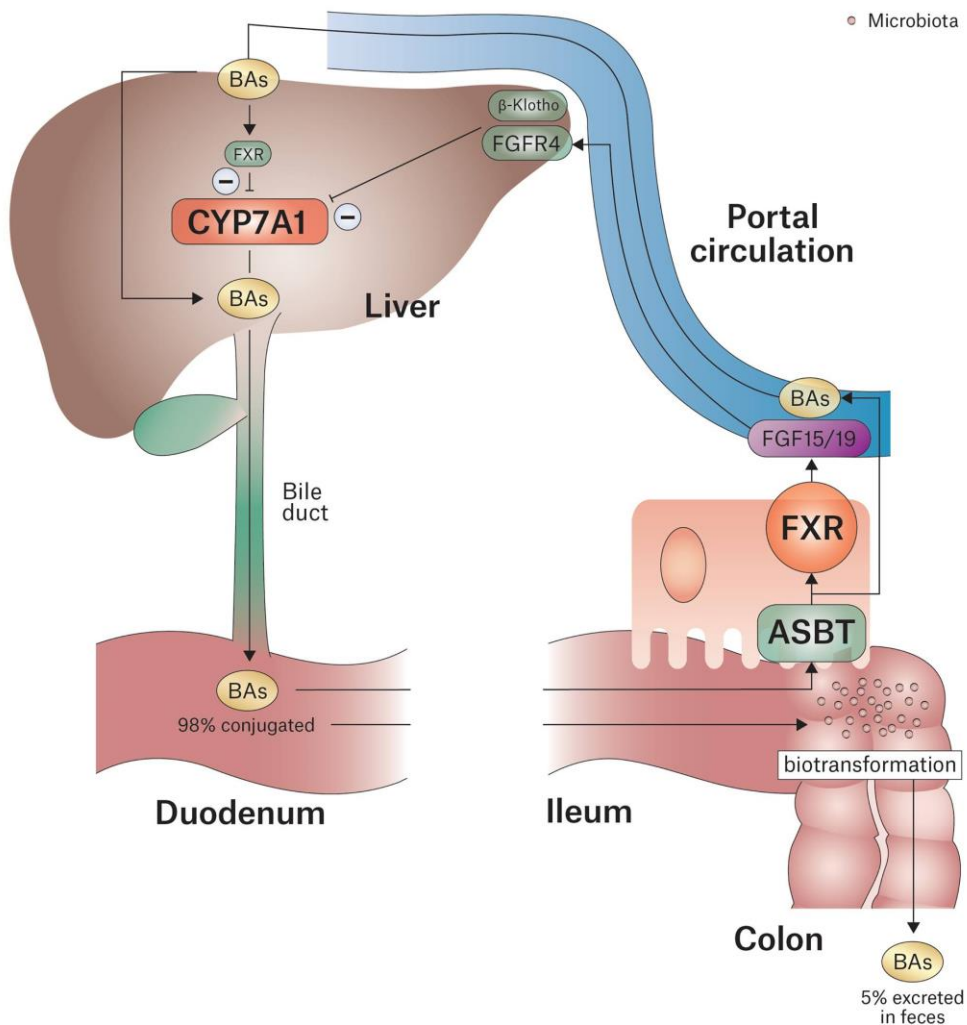


Figure 1. Schematic representation of the enterohepatic circulation of bile acids. Bile acids (BAs) are synthesized and conjugated in the liver after which they are actively secreted via the bile duct into the duodenum. In the ileum BAs are reabsorbed by the enterocytes through the apical sodium-dependent bile acid transporter (ASBT). In the enterocyte BAs activate the nuclear farnesoid X receptor (FXR) which leads to subsequent release of fibroblast growth factor 19 (FGF19 or 15 in mice). FGF15/19 travels to the liver where it binds to fibroblast growth factor receptor 4 (FGFR4) β-Klotho complex and inhibits cholesterol 7 α -hydroxylase (CYP7A1), the rate controlling enzyme of BA synthesis. Reabsorbed BAs can also travel directly to the liver and inhibit CYP7A1 by activating hepatic FXR. Microbiota in the intestine can further biotransform BAs to deconjugated and secondary BA species. After reabsorption, the remaining 5% of BAs (in the physiological situation) is excreted in feces.

Reabsorption mainly takes place by active transport of conjugated BAs into the ileal enterocyte by the apical sodium-dependent bile acid transporter (ASBT, SLC10A2) (33). In the ileal enterocyte BAs activate the farnesoid X receptor (FXR), a ligand-activated transcription factor of the family of nuclear receptors, which leads to increased expression and subsequent release of fibroblast growth factor 19 (FGF19, Fgf15 in mice) in the circulation (34). In the liver FGF19 can bind to and activate the FGF receptor 4 (FGFR4)/ β -Klotho complex which in turn exerts negative feedback on the rate controlling enzyme of BA synthesis, cholesterol 7 α -hydroxylase (CYP7A1). Reabsorbed BAs can also cause negative feedback by directly activating hepatic FXR. However, organ specific Fxr knockout studies in mice indicated a much more prominent role for the FXR-FGF15/19 axis in CYP7A1 repression (35).

The importance of FXR-FGF15/19 signaling in BA homeostasis is illustrated by various BA malabsorption syndromes. Clinically, BA malabsorption causes diarrhea due to high BA concentrations in the colon that lead to secretion of water and electrolytes and stimulation of propulsive contractions (36). Patients with primary BA diarrhea, a condition in which BA malabsorption occurs in the absence of ileal or other obvious GI disease, display lower levels of FGF19 and higher levels of 7 α -hydroxy-4-cholesten-3-one (C4), a surrogate marker for BA synthesis (37,38). Additionally, in a murine model of BA malabsorption it has been shown that Fxr activation or Fgf15 administration reduces fecal BA excretion (39).

Even though CF patients display BA malabsorption, (BA) diarrhea is rare and constipation due to inspissated mucus and delayed intestinal transit is more common (further explained in section 6). However, BA malabsorption in CF patients does result in impaired FXR-FGF19 signaling. Our group recently demonstrated that plasma FGF19 levels are lower and C4 levels higher in CF patients with a G551D gating-mutation as compared to healthy controls (40). Conversely, improving CFTR function by treatment with the CFTR potentiator ivacaftor, altered the levels of these parameters towards normality, supporting a role for CFTR involvement in BA homeostasis. These results also indicate the possibility of using plasma C4 and FGF19 as surrogate biomarkers for CFTR function in the GI tract in CF. This is important as currently GI markers in CF are limited and/or difficult to obtain (41).

BA homeostasis has been more extensively studied in CF mouse models than in CF patients. Debray *et al.* showed that *Cftr*^{-/-} mice have lower ileal expression levels of Fgf15 compared to wild type controls, while having similar fecal BA excretion rates (42). Interestingly, most other studies in CF mouse models showed

an about 3-fold increase in fecal BA excretion, similar to what has been observed in CF patients, but did not directly assess FXR-FGF15/19 signaling (31,32). This discrepancy in fecal BA excretion might be explained by genotypic and dietary differences between studies and requires further investigation (43). Indeed, unpublished data from our group confirms lower ileal expression levels of *Fgf15* in combination with increased hepatic *Cyp7a1* levels in *Cftr*^{-/-} mice, supporting the disruption of FXR-FGF15/19 signaling. The exact mechanism underlying the impaired FXR-FGF15/19 signaling in CF, however, remains unknown.

In wild type mice it has been shown that uptake of BAs by Asbt activates *Cftr* (31). Altered functionality or expression of ASBT might also be involved in BA malabsorption in CF. However, results on ileal *Asbt* expression in murine CF models have been conflicting, (31,42,44), likely due to differences in experimental setups. Additionally, gene expression might not adequately reflect protein expression or activity. When looking at protein abundance, Debray *et al.* (42) found decreased expression of *Asbt* by western blot in *Cftr*^{-/-} mice, while Bijvelds *et al.* (31) showed a robust *Asbt* immunohistochemistry staining pattern and intensity in both *Cftr*^{-/-} and F508del-*Cftr* mice indistinguishable from WT mice. The regulation of ASBT expression is complex and can be influenced by many factors including intestinal BA concentrations and microbial composition (33). ASBT expression is under a negative feedback regulation by intestinal BA concentrations in mice and humans (45,46). The proposed mechanism of this feedback regulation is that BAs activate FXR in the enterocyte which leads to subsequent small heterodimer partner (SHP) and liver receptor homologue-1 (LRH-1) activation which downregulate ASBT expression. This hypothesis is supported by the fact that BA depletion by feeding mice a BA binding resin increased *Asbt* expression (47). However, Debray *et al.* showed, besides decreased *Asbt* expression in *Cftr*^{-/-} mice, a decrease in expression of ileal *Fxr* target genes, *Fgf15* and *Shp*, arguing against suppression of ASBT by FXR (42). Conversely, while decreased intestinal BA concentrations seemed to induce ASBT expression, Stravitz *et al.* showed an induction of *Asbt* gene and protein expression in rats by feeding cholic acid or perfusing the intestine with taurocholic acid (48). Intestinal dysbiosis or SIBO could contribute to altered ASBT expression in CF. Germfree or antibiotic treated mice generally show higher expression levels of *Asbt* and a decrease in fecal BA excretion (49–51). Higher expression levels of *Asbt* were also observed more proximal in the intestine when mice were treated with antibiotics, regulated through reduced expression of the transcription factor *Gata4* (50).

Next to their indirect effect on ASBT expression, microbiota are also directly involved in BA homeostasis and FXR-FGF15/19 signaling. Different species of intestinal microbiota have the ability to biotransform BAs mainly by deconjugation and subsequent de-hydroxylation, the latter resulting in more hydrophobic secondary BAs. The various BA species have different properties regarding absorption and receptor activation. Many species of microbiota express bile salt hydrolase (BSH) activity which results in deconjugation of part of the luminal BAs, thereby making them unable to be reabsorbed by ASBT but, due to decreased polarity (more hydrophobic), easier to be passively reabsorbed. Germ-free mice lack the microbial ability to deconjugate BAs, resulting in higher concentrations of tauro- β -muricholic acid, a conjugated BA that acts as an Fxr antagonist that can lower Fgf15 levels (49).

CF patients and murine CF models display intestinal dysbiosis and sometimes even bacterial overgrowth, potentially affecting BA handling. *Cftr*^{-/-} mice generally show a decreased intestinal microbiota biodiversity with an increase in species associated with inflammation (52). Decreases within the *Bacteroides* and *Firmicutes* phyla were observed in *Cftr*^{-/-} mice as well as CF patients (24). The *Firmicutes* phylum contains species known to be involved in de-hydroxylation of BAs (53). A decrease in one of the most important secondary BAs, deoxycholic acid (DCA), has been observed in feces of *Cftr*^{-/-} mice (17,32). These observations suggest an important relation between intestinal dysbiosis in CF and BA handling. The exact role, however, of microbiota in BA homeostasis in CF remains to be elucidated.

Bile acid homeostasis and its relation to metabolic function in cystic fibrosis

BAs are primarily known as detergents with their main function to aid in digestion and absorption of fat and fat soluble vitamins. However, recent studies show that BAs also act as ligands for receptors resulting in the release of hormones that affect other metabolic processes in the body such as glucose metabolism (54). CF patients often display metabolic abnormalities, such as hyperglycemia, hypertriglyceridemia or steatosis, and these are more common with increasing age (55). Malnutrition is generally regarded as an important problem in CF management. However, due to better nutritional and supportive therapy, the prevalence of malnutrition in CF has declined and some CF patients even develop overweight or obesity (56). A recent study showed that some CF patients display “normal weight obesity”, defined by a normal body mass index (BMI) but increased fat percentage that was associated with decreased pulmonary function (57).

	Effects of BA homeostasis	Result	Findings in CF patients
Lipid metabolism	Catabolism of cholesterol for BA synthesis	Lower plasma cholesterol	Often low plasma cholesterol (55,58) BA synthesis is elevated (40)
Glucose metabolism	Intestinal bile acids activate TGR5 signaling and GLP-1 release	GLP-1 improves post-prandial insulin secretion	Reduced GLP-1 release in CF patients (59–61)
	Intestinal bile acids activate FXR to release FGF19	FGF19 inhibits bile acid synthesis and improves insulin sensitivity	Lower FGF19 due to reduced BA absorption (40)

Table 1. Summary of the metabolic effects of bile acid homeostasis and the phenotype in CF as discussed in section 4. BA: bile acid; CF: cystic fibrosis; TGR5: G-protein coupled bile acid receptor 1; FGF19: fibroblast growth factor 19; GLP-1: glucagon-like peptide-1; CFRD: cystic fibrosis related diabetes.

BA homeostasis is also highly involved in lipid and cholesterol metabolism (54). Conversion of cholesterol to BAs and subsequent fecal excretion is one of the main routes of cholesterol disposal. Most CF patients display serum low-density lipoprotein cholesterol (LDL-C) levels that are lower compared to control subjects of similar age (55,58). However, some CF patients display elevated LDL-C and triglyceride levels which are associated with older age and other metabolic abnormalities such as a high BMI and lower insulin sensitivity (62–64). With increasing age the incidence of CF related diabetes (CFRD) is also rising. The pathophysiology of CFRD is not completely understood but its incidence is correlated with EPI and fibrosis (65). A combination of partial insulin deficiency and episodes of insulin resistance are often present. Interestingly, CF patients display lower levels of glucagon like peptide 1 (GLP-1), an incretin hormone regulating postprandial insulin secretion, that is improved by pancreatic enzyme replacement therapy (59–61). Intestinal activation of the G-protein coupled bile acid receptor 1 (GPBAR1, GPCR19 also known as TGR5) by BAs enhances the secretion of GLP-1 (66). Secondary BAs lithocholic acid (LCA) and DCA are the most potent naturally occurring TGR5 agonists. As these BA species are lower in murine CF models, it is tempting to speculate that TGR5 activation is reduced in CF due to impaired BA homeostasis. Unfortunately, however, no studies directly addressing this relationship have been performed.

In addition to the role of TGR5, the FXR-FGF15/19 axis has been implicated in other aspects of glucose metabolism and metabolic disorders. In humans, both obesity and type 2 diabetes mellitus (T2DM) are associated with lower plasma FGF19 levels (67,68). A derivative of the naturally occurring FXR agonist CDCA, 6 α -ethyl-CDCA (obeticholic acid, OCA), was recently approved by the Food and Drug Administration (FDA) for the treatment of primary biliary cholangitis (PBC), and is in clinical trials for treatment of non-alcoholic steatohepatitis. Administration of OCA to patients with non-alcoholic fatty liver disease and T2DM increased plasma FGF19 levels, decreased liver enzymes (alanine aminotransferase and γ -glutamyltransferase) and improved insulin sensitivity (69). In a phase 3 clinical trial, OCA ameliorated histology scores of non-alcoholic steatohepatitis patients (70). Animal studies, however, have not generated unambiguous results regarding the effects of the FXR-FGF15/19 axis on metabolism. Transgenic mice with hepatic overexpression of FGF19 display an increased metabolic rate and decreased adiposity (71). Kir *et al.* found that administration of FGF19 to WT mice improved glucose metabolism by inducing hepatic glycogen and protein synthesis (72). Conversely, *Fgf15*^{-/-} mice showed glucose intolerance and a reduced hepatic glycogen content. Administration of FGF19 was even found to reverse diabetes mellitus in *ob/ob* mice (73). A direct involvement of *Fxr-Fgf15* signaling was demonstrated by intestinal inactivation of *Fxr* or reducing intestinal *Fxr* signaling through remodeling the intestinal microbial profile by the anti-oxidant tempol, both of which reduced diet induced obesity and hepatic triglyceride accumulation in mice (74,75). The metabolic improvements by tempol were explained by reduced species of *Lactobacillus* and their BSH activity resulting in increased levels of tauro- β -muricholic acid, a naturally occurring FXR antagonist. This claim was further supported by a study in which administration of glyco- β -muricholic acid (Gly-MCA), a selective FXR inhibitor, reduced obesity and improved related metabolic abnormalities in mice (76). On the other hand, intestine-specific FXR activation using the intestinally restricted FXR agonist fexaramine, reduced diet-induced obesity, insulin resistance and steatosis in mice fed a HFD (77). However, a recent study explained the beneficial effects of fexaramine by an indirect effect on the microbiota and activation of TGR5 via pronounced alterations in BA composition (78).

As the pathology of CFRD is thought to be mainly due to (relative) insulin deficiency rather than insulin resistance as in T2DM, the effects of modulating the FXR-FGF15/19 axis in CFRD could be less pronounced. On the other hand, CF is associated with direct impairment of BA homeostasis and FXR-FGF15/19

signaling. It will therefore be interesting to evaluate strategies to improve FXR-FGF15/19 signaling in CF, especially in the prevention of metabolic complications such as CFRD and hepatic steatosis.

Bile acid homeostasis and its role in cystic fibrosis liver disease

Cystic fibrosis liver disease (CFLD) is a severe complication of CF and an independent risk factor for mortality (79,80). According to the 2016 CF patient registry report, CFLD accounted for 2.7% of mortality in CF patients (81). CFLD is an umbrella term used to describe various types of liver dysfunction in CF of which the pathophysiology is often not completely understood (82). In the liver, CFTR is expressed exclusively at the apical membrane of cholangiocytes lining the bile ducts (15,83). One proposed mechanism for the development of CFLD is that loss of CFTR function leads to obstruction of the bile ducts by thickening of the mucus, eventually resulting in obstructive biliary cirrhosis. Murine CF models generally do not display CFLD except upon ageing (84,85). The absence of CFLD in murine CF models might be partially explained by lower BA toxicity due to differences in biliary BA composition between mice and humans. Mice generally have a higher concentration of hydrophilic BAs which is regarded as less cytotoxic since hydrophilic BAs have a lower detergent capacity than hydrophobic BAs (86). High concentrations of hydrophobic BAs, mainly DCA, have been associated with increased risk of cholesterol gallstone disease, colon cancer and liver cancer (53,87).

FXR-FGF15/19 signaling and hepatic FXR activation have been consistently shown to affect liver regeneration and proliferation (88–91). This regenerative/proliferative response, measured by hepatic staining of the proliferation marker Ki67, is absent in *Cftr*^{-/-} mice upon feeding cholic acid (CA), a strong FXR agonist (17). This suggests that a CF liver might have reduced regenerative ability due to BA malabsorption. Interestingly, in humans the presence of CFLD is associated with normalization of fecal BA excretion which could be due to impaired production as declining liver function occurs but this has not been further investigated (2).

Currently, ursodeoxycholic acid (UDCA) is the only recommended and widely used drug in the treatment of CFLD. However, the clinical efficacy of UDCA is controversial. The most recent Cochrane review only identified a small number of trials assessing the effectiveness of UDCA (92). The authors concluded that there is 'currently insufficient evidence to justify its routine use in cystic fibrosis'. UDCA treatment is often started early in life to prevent severe CFLD and related

complications. This was also challenged by the results of a recent study, showing that treatment with UDCA started earlier in life had no effect on development of severe CFLD (13).

The effects of UDCA on BA homeostasis and FXR signaling have not been fully elucidated. UDCA increases hepatocellular and cholangiocellular secretion thereby increasing bile flow and reducing biliary toxicity (93). UDCA is also suggested to decrease hepatic steatosis in mice (94–96). In obese subjects UDCA was reported to lower FGF19 and subsequently increase hepatic bile acid synthesis (97). The authors explained this as UDCA having FXR antagonistic effects which was supported by showing decreased FXR activation in an avidin biotin complex DNA-assay. However, *in vitro* assays suggest UDCA has neither FXR agonistic nor antagonistic properties (96,98). UDCA is readily absorbed and constitutes a predominant part of the BA pool in UDCA treated patients (from 40% in PBC treated patients (99) to almost 90% in obese patients (97)), which is likely to contribute to the effects on the FXR-FGF15/19 axis. Fujita *et al.* showed a clear decrease in relative and absolute levels of muricholic acid levels in mouse livers after UDCA treatment, arguing the beneficial effects on hepatic steatosis (at least in mice) might be due to a reduction of the FXR antagonistic muricholic acid species (96).

FXR agonism has been shown to protect against hepatotoxicity in a rat model of intrahepatic cholestasis (100). In that study a systemic FXR agonist (i.e. GW4064) was used and effects could therefore be at least partly due to hepatic FXR activation. Direct evidence of the benefits of intestinal FXR-FGF15/19 signaling has been shown by Modica *et al.* (101) who demonstrated that transgenic overexpression of intestinal FXR or administration of FGF19 in mice protects against liver damage in three different models of cholestasis. The beneficial effects were attributed to a reduced BA pool size and more hydrophilic (i.e. less cytotoxic) biliary BA composition. However, FGF19 is a growth factor and is also associated with the induction of liver proliferation and growth of cancer cells (102). To overcome the potential tumorigenic effects of FGF19, a modified variant of FGF19 (M70) has been produced, with reduced tumorigenicity but retained benefits in cholestatic liver disease in mice (103,104).

These results make intestinal FXR an interesting target in developing treatment and prevention strategies for CFLD. Unfortunately, CF mice have not been an ideal model for CFLD. Recently, however, other potentially more useful CF animal models have been developed, including the CF pig which already shows signs of

CFLD at birth (105,106). CF pigs could therefore be interesting to study CFLD (107,108).

Modulating bile acid homeostasis to improve gastrointestinal outcomes in cystic fibrosis

Considering the close relation between BA homeostasis and GI and metabolic function, it is interesting to speculate about the effect of modulating the factors involved. In the previous section the effects of altering FXR-FGF15/19 and TGR5 signaling to improve metabolism were considered. Modulating the FXR-FGF15/19 axis might also ameliorate GI outcomes. In turn, modulating certain GI factors could improve BA homeostasis and metabolic outcomes.

CF patients often display SIBO or colonic dysbiosis. Not only do these conditions generate direct symptoms including abdominal discomfort, diarrhea and flatulence, they also increase the risk of developing metabolic abnormalities and liver disease (109). The relationship between BAs and intestinal microbiota is complex, with mutual interactions (87,110). In the CF intestine, inspissated mucus accumulates, making it easier for harmful bacteria to thrive (27). Other factors contributing to an altered microbial profile include a low intestinal pH due to reduced bicarbonate secretion, a longer intestinal transit time and exposure to antibiotics that CF patients frequently receive for (suspected) pulmonary infections. Interestingly, bacterial overgrowth itself has also been suggested to contribute to mucus secretion. Antibiotic treatment aimed at eradication of bacterial overgrowth in *Cftr*^{-/-} mice reduced mucus accumulation without a major effect on mucin gene expression, suggesting a more direct role for bacteria on mucus secretion by intestinal epithelium (111).

Treatment with probiotics can also be used to alter the microbiota profile of CF patients. One study reports that administration of the probiotic *Lactobacillus Reuteri* improved digestive health and inflammation (112). The fecal microbial profile changed, showing a decrease in *Proteobacteria* and an increase of the *Firmicutes* phylum. As numerous species of the *Firmicutes* phylum are involved in BA biotransformation, one could speculate that this change affects BA homeostasis. Treatment with another probiotic, *Lactobacillus GG*, decreased fecal calprotectin, a marker for intestinal inflammation, and changed microbial composition partially towards that of healthy controls (113,114). However, in a large trial in CF children, one year of treatment with *Lactobacillus GG* versus placebo did not affect hospitalization, pulmonary outcomes or BMI (115).

Nevertheless, as probiotic use does seem to improve the GI phenotype of CF, it will be interesting to evaluate its effects on BA homeostasis and metabolic function.

Another GI feature of CF patients is a delayed intestinal transit time, possibly leading to constipation or in severe cases distal intestinal obstruction syndrome (DIOS) (116–118). Interestingly, Bijvelds *et al.* showed that active BA absorption in the ileum triggered CFTR activation and subsequent local salt and water excretion (31). It is therefore tempting to speculate that the absence of this postprandial ileal water release contributes to specific distal localization of obstruction occurring in CF patients. The inability to sufficiently hydrate intestinal content and mucus likely explains the fact that BA malabsorption does not lead to diarrhea in CF.

As CF patients often suffer from constipation, laxatives are commonly prescribed. Laxative treatment shortens intestinal transit time and is able to alter microbiota and BA homeostasis. In rats, the commonly used laxative polyethylene glycol (PEG) decreased BA dehydroxylation, increasing the amount of primary BAs in the BA pool (119). Whole body *Cftr* knockout mice display a severe intestinal phenotype and need to be kept either on a liquid diet or a solid diet in combination with laxative. A study by De Lisle *et al.* compared the effects of either a solid diet with PEG or a liquid diet with or without n-acetylcysteine (NAC), a mucolytic agent, on various aspects of the intestinal phenotype (120). Laxative treatment had pronounced effects on the intestine and improved markers of intestinal inflammation and reduced bacterial overgrowth. In CF patients, laxative treatment was also found to be associated with a decrease in occurrence of SIBO (121). Considering these beneficial effects of laxative treatment on the CF intestine, it is tempting to speculate that laxative treatment could decrease fecal BA excretion.

Summary

The presently increased life span of the CF population changes the frequency and spectrum of symptoms regarding GI and metabolic function including intestinal dysbiosis, constipation, intestinal cancer, liver disease and diabetes. One part of the CF phenotype that recently received more attention is the GI tract, including impaired BA homeostasis, characterized by increased fecal BA excretion and reduced FXR-FGF15/19 signaling. Modulating BA homeostasis directly by altering FXR-FGF15/19 or TGR5 signaling or indirectly by improving intestinal transit or modifying intestinal microbiota are potential strategies to improve other GI and metabolic CF complications. Lastly, emerging research into BA homeostasis and the GI phenotype of CF could provide novel easily measurable surrogate biomarkers (e.g. C4, FGF19). Especially in the current era of new CFTR modulator therapies (e.g. ivacaftor and lumacaftor), the need for such biomarkers has increased.

Acknowledgements

None.

Funding sources

This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

Declaration of interest

None.

References

1. Rowe SM, Miller S, Sorscher EJ. Cystic Fibrosis. *N Engl J Med*. 2005;1992-2001.
2. O'Brien S, Mulcahy H, Fenlon H, et al. Intestinal bile acid malabsorption in cystic fibrosis. *Gut*. 1993;34(December):1137-1141.
3. De Lisle RC, Borowitz D. The cystic fibrosis intestine. *Cold Spring Harb Perspect Med*. 2013;3(9).
4. Haller W, Ledder O, Lewindon PJ, Couper R, Gaskin KJ, Oliver M. Cystic fibrosis: An update for clinicians. Part 1: Nutrition and gastrointestinal complications. *J Gastroenterol Hepatol*. 2014;29(7):1344-1355.
5. Ahmed N, Corey M, Forstner G, et al. Molecular consequences of cystic fibrosis transmembrane regulator (CFTR) gene mutations in the exocrine pancreas. *Gut*. 2003;52(8):1159-1164.
6. O'Sullivan BP, Baker D, Leung KG, Reed G, Baker SS, Borowitz D. Evolution of pancreatic function during the first year in infants with cystic fibrosis. *J Pediatr*. 2013;162(4):808-812.e1.
7. Gaskin KJ. Nutritional care in children with cystic fibrosis: are our patients becoming better? *Eur J Clin Nutr*. 2013;67(5):558-564.
8. Kalivianakis M, Minich DM, Bijleveld CMA, et al. Fat malabsorption in cystic fibrosis patients receiving enzyme replacement therapy is due to impaired intestinal uptake of long-chain fatty acids. *Am J Clin Nutr*. 1999;69(1):127-134.
9. Borowitz D, Durie PRP, Clarke LL, et al. Gastrointestinal outcomes and confounders in cystic fibrosis. *J Pediatr Gastroenterol Nutr*. 2005;41(3):273-285.
10. Bodewes FAJA, Verkade HJ. Persistent fat malabsorption in cystic fibrosis - lessons from patients and mice. *J Cyst Fibros*. 2011;10(3):150-158.
11. Bijvelds MJC, Bronsveld I, Havinga R, Sinaasappel M, de Jonge HR, Verkade HJ. Fat absorption in cystic fibrosis mice is impeded by defective lipolysis and post-lipolytic events. *Am J Physiol Gastrointest Liver Physiol*. 2005;288(4):G646-53.
12. Feranchak AP, Sokol RJ. Cholangiocyte biology and cystic fibrosis liver disease. *Semin Liver Dis*. 2001;21(4):471-488.
13. Boelle P, Debray D, Guillot L, Clement A, Corvol H. Cystic Fibrosis Liver Disease: Outcomes and risk factors in a large cohort of French patients. *Hepatology*. 2018;In press.
14. Koh C, Sakiani S, Surana P, et al. Adult Onset Cystic Fibrosis Liver Disease: Diagnosis and characterization of an underappreciated entity. *Hepatology*. 2017;66(2):591-601.
15. Cohn JA, Strong T V, Picciotto MR, Nairn AC, Collins FS, Fitz JG. Localization of the cystic fibrosis transmembrane conductance regulator in human bile duct epithelial cells. *Gastroenterology*. 1993;105(6):1857-1864.
16. Moyer K, Balistreri W. Hepatobiliary disease in patients with cystic fibrosis. *Curr Opin Gastroenterol*. 2009;25(3):272-278.
17. Bodewes FAJA, Bijvelds MJ, De Vries W, et al. Cholic acid induces a Cfr

- dependent biliary secretion and liver growth response in mice. *PLoS One*. 2015;10(2):1-14.
18. Casaccia G, Trucchi A, Nahom A, et al. The impact of cystic fibrosis on neonatal intestinal obstruction: the need for prenatal/neonatal screening. *Pediatr Surg Int*. 2003;19:75-78.
 19. Van Der Doef HPJ, Kokke FTM, Van Der Ent CK, Houwen RHJ. Intestinal obstruction syndromes in cystic fibrosis: Meconium ileus, distal intestinal obstruction syndrome, and constipation. *Curr Gastroenterol Rep*. 2011;13(3):265-270.
 20. Munck A, Alberti C, Colombo C, et al. International prospective study of distal intestinal obstruction syndrome in cystic fibrosis: Associated factors and outcome. *J Cyst Fibros*. 2016;15(4):531-539.
 21. van der Doef HPJ, Kokke FTM, Beek FJA, Woestenenk JW, Froeling SP, Houwen RHJ. Constipation in pediatric Cystic Fibrosis patients: An underestimated medical condition. *J Cyst Fibros*. 2010;9(1):59-63.
 22. Garg M, Ooi CY. The Enigmatic Gut in Cystic Fibrosis: Linking Inflammation, Dysbiosis, and the Increased Risk of Malignancy. *Curr Gastroenterol Rep*. 2017;19(2):6.
 23. Nicholson JK, Holmes E, Wilson ID. Gut microorganisms, mammalian metabolism and personalized health care. *Nat Rev Microbiol*. 2005;3(5):431-438.
 24. Li L, Somerset S. The clinical significance of the gut microbiota in cystic fibrosis and the potential for dietary therapies. *Clin Nutr*. 2014;33(4):571-580.
 25. Maisonneuve P, Marshall BC, Knapp EA, Lowenfels AB. Cancer risk in cystic fibrosis: A 20-year nationwide study from the United States. *J Natl Cancer Inst*. 2013;105(2):122-129.
 26. Neglia JP, FitzSimmons SC, Maisonneuve P, et al. The Risk of Cancer among Patients with Cystic Fibrosis. *N Engl J Med*. 1995;332(8):494-499.
 27. Ooi CY, Durie PR. Cystic fibrosis from the gastroenterologist's perspective. *Nat Rev Gastroenterol Hepatol*. 2016;13(3):175-183.
 28. Than BLN, Linnekamp JF, Starr TK, et al. CFTR is a tumor suppressor gene in murine and human intestinal cancer. *Oncogene*. 2016;(April 2015):1-9.
 29. Fink AK, Yanik EL, Marshall BC, et al. Cancer risk among lung transplant recipients with cystic fibrosis. *J Cyst Fibros*. 2017;16(1):91-97.
 30. Strandvik B, Einarsson K, Lindblad A, Angelin B. Bile acid kinetics and biliary lipid composition in cystic fibrosis. *J Hepatol*. 1996;25(1):43-48.
 31. Bijvelds MJC, Jorna H, Verkade HJ, et al. Activation of CFTR by ASBT-mediated bile salt absorption. *Am J Physiol Gastrointest Liver Physiol*. 2005;289(5):G870-9.
 32. Bodewes FAJA, van der Wulp MYM, Beharry S, et al. Altered intestinal bile salt biotransformation in a cystic fibrosis (Cftr^{-/-}) mouse model with hepato-biliary pathology. *J Cyst Fibros*. 2015;14(4):440-446.
 33. Dawson P a, Lan T, Rao A. Bile acid transporters. *J Lipid Res*. 2009;50(12):2340-2357.
 34. Inagaki T, Choi M, Moschetta A, et al. Fibroblast growth factor 15 functions as an

- enterohepatic signal to regulate bile acid homeostasis. *Cell Metab.* 2005;2(4):217-225.
35. Kim I, Ahn S-H, Inagaki T, et al. Differential regulation of bile acid homeostasis by the farnesoid X receptor in liver and intestine. *J Lipid Res.* 2007;48(12):2664-2672.
 36. Camilleri M. Advances in understanding of bile acid diarrhea. *Expert Rev Gastroenterol Hepatol.* 2014;8(1):49-61.
 37. Pattni SS, Brydon WG, Dew T, Walters JRF. Fibroblast Growth Factor 19 and 7 α -Hydroxy-4-Cholesten-3-one in the Diagnosis of Patients With Possible Bile Acid Diarrhea. *Clin Transl Gastroenterol.* 2012;3(7):e18.
 38. Vijayvargiya P, Camilleri M, Carlson P, et al. Performance characteristics of serum C4 and FGF19 measurements to exclude the diagnosis of bile acid diarrhoea in IBS-diarrhoea and functional diarrhoea. *Aliment Pharmacol Ther.* 2017;46(6):581-588.
 39. Jung D, Inagaki T, Gerard RD, et al. FXR agonists and FGF15 reduce fecal bile acid excretion in a mouse model of bile acid malabsorption. *J Lipid Res.* 2007;48(12):2693-2700.
 40. Bodewes FAJA, Doktorova M, van de Peppel IP, van der Ley C, Jonker JW, Verkade HJ. Ivacaftor restores the enterohepatic feedback regulation of the bile acid homeostasis in patients with a Cfr G551D mutation. *Pediatr Pulmonol.* 2015;50:297.
 41. Bodewes FAJA, Verkade HJ, Taminiou JAJM, Borowitz D, Wilschanski M. Cystic fibrosis and the role of gastrointestinal outcome measures in the new era of therapeutic CFTR modulation. *J Cyst Fibros.* 2015;14(2):169-177.
 42. Debray D, Rainteau D, Barbu V, et al. Defects in gallbladder emptying and bile acid homeostasis in mice with cystic fibrosis transmembrane conductance regulator deficiencies. *Gastroenterology.* 2012;142(7):1581-1591.e6.
 43. Bijvelds MJC, De Jonge HR, Verkade HJ. Bile acid handling in cystic fibrosis: Marked phenotypic differences between mouse models. *Gastroenterology.* 2012;143(6):e19-e20.
 44. Stelzner M, Somasundaram S, Lee SP, Kuver R. Ileal mucosal bile acid absorption is increased in Cfr knockout mice. *BMC Gastroenterol.* 2001;1:1-10.
 45. Neimark E, Chen F, Li X, Shneider BL. Bile acid-induced negative feedback regulation of the human ileal bile acid transporter. *Hepatology.* 2004;40(1):149-156.
 46. Chen F, Ma L, Dawson PA, et al. Liver receptor homologue-1 mediates species- and cell line-specific bile acid-dependent negative feedback regulation of the apical sodium-dependent bile acid transporter. *J Biol Chem.* 2003;278(22):19909-19916.
 47. Torchia EC, Cheema SK, Agellon LB. Coordinate regulation of bile acid biosynthetic and recovery pathways. *Biochem Biophys Res Commun.* 1996;225(1):128-133.
 48. Stravitz RT, Sanyal AJ, Pandak WM, Vlahcevic ZR, Beets JW, Dawson PA. Induction of sodium-dependent bile acid transporter messenger RNA, protein, and activity in rat ileum by cholic acid. *Gastroenterology.* 1997;113(5):1599-1608.
 49. Sayin SI, Wahlström A, Felin J, et al. Gut microbiota regulates bile acid metabolism by reducing the levels of tauro-beta-muricholic acid, a naturally occurring FXR antagonist. *Cell Metab.* 2013;17(2):225-235.

50. Out C, Patankar J V., Doktorova M, et al. Gut microbiota inhibit Asbt-dependent intestinal bile acid reabsorption via Gata4. *J Hepatol*. 2015;63(3):697-704.
51. Miyata M, Takamatsu Y, Kuribayashi H, Yamazoe Y. Administration of ampicillin elevates hepatic primary bile acid synthesis through suppression of ileal fibroblast growth factor 15 expression. *J Pharmacol Exp Ther*. 2009;331(3):1079-1085.
52. Lynch S V., Goldfarb KC, Wild YK, Kong W, De Lisle RC, Brodie EL. Cystic fibrosis transmembrane conductance regulator knockout mice exhibit aberrant gastrointestinal microbiota. *Gut Microbes*. 2013;4(1):41-47.
53. Ridlon JM, Kang D-J, Hylemon PB. Bile salt biotransformations by human intestinal bacteria. *J Lipid Res*. 2006;47(2):241-259.
54. Kuipers F, Bloks VW, Groen AK. Beyond intestinal soap--bile acids in metabolic control. *Nat Rev Endocrinol*. 2014;10(8):488-498.
55. Georgiopoulou V V., Denker A, Bishop KL, et al. Metabolic abnormalities in adults with cystic fibrosis. *Respirology*. 2010;15(5):823-829.
56. Hanna RM, Weiner DJ. Overweight and obesity in patients with cystic fibrosis: A center-based analysis. *Pediatr Pulmonol*. 2015;50(1):35-41.
57. Alvarez JA, Ziegler TR, Millson EC, Stecenko AA. Body composition and lung function in cystic fibrosis and their association with adiposity and normal-weight obesity. *Nutrition*. 2016;32(4):447-452.
58. Figueroa V, Milla C, Parks EJ, Schwarzenberg SJ, Moran A. Abnormal lipid concentrations in cystic fibrosis. *Am J Clin Nutr*. 2002;75:1005-1011.
59. Hillman M, Eriksson L, Mared L, Helgesson K, Landin-Olsson M. Reduced levels of active GLP-1 in patients with cystic fibrosis with and without diabetes mellitus. *J Cyst Fibros*. 2012;11(2):144-149.
60. Perano SJ, Couper JJ, Horowitz M, et al. Pancreatic enzyme supplementation improves the incretin hormone response and attenuates postprandial glycemia in adolescents with cystic fibrosis: A randomized crossover trial. *J Clin Endocrinol Metab*. 2014;99(7):2486-2493.
61. Kuo P, Stevens JE, Russo A, et al. Gastric emptying, incretin hormone secretion, and postprandial glycemia in cystic fibrosis - Effects of pancreatic enzyme supplementation. *J Clin Endocrinol Metab*. 2011;96(5):851-855.
62. Ishimo M-C, Belson L, Ziai S, et al. Hypertriglyceridemia is associated with insulin levels in adult cystic fibrosis patients. *J Cyst Fibros*. 2013;12(3):271-276.
63. Coderre L, Fadainia C, Belson L, et al. LDL-cholesterol and insulin are independently associated with body mass index in adult cystic fibrosis patients. *J Cyst Fibros*. 2012;11(5):393-397.
64. Rhodes B, Nash EF, Tullis E, et al. Prevalence of dyslipidemia in adults with cystic fibrosis. *J Cyst Fibros*. 2010;9(1):24-28.
65. Kelly A, Moran A. Update on cystic fibrosis-related diabetes. *J Cyst Fibros*. 2013;12(4):318-331.
66. Thomas C, Gioiello A, Noriega L, et al. TGR5-Mediated Bile Acid Sensing Controls Glucose Homeostasis. *Cell Metab*. 2009;10(3):167-177.

67. Roesch SL, Styer AM, Wood GC, et al. Perturbations of fibroblast growth factors 19 and 21 in type 2 diabetes. *PLoS One*. 2015;10(2):1-12.
68. Gallego-Escuredo JM, Gómez-Ambrosi J, Catalan V, et al. Opposite alterations in FGF21 and FGF19 levels and disturbed expression of the receptor machinery for endocrine FGFs in obese patients. *Int J Obes (Lond)*. 2015;39(April 2014):121-129.
69. Mudaliar S, Henry RR, Sanyal AJ, et al. Efficacy and safety of the farnesoid x receptor agonist Obeticholic acid in patients with type 2 diabetes and nonalcoholic fatty liver disease. *Gastroenterology*. 2013;145(3):574-582.e1.
70. Neuschwander-Tetri BA, Loomba R, Sanyal AJ, et al. Farnesoid X nuclear receptor ligand obeticholic acid for non-cirrhotic, non-alcoholic steatohepatitis (FLINT): A multicentre, randomised, placebo-controlled trial. *Lancet*. 2015;385(9972):956-965.
71. Tomlinson E, Fu L, John L, et al. Transgenic mice expressing human fibroblast growth factor-19 increased metabolic rate and decreased adiposity. *Endocrinology*. 2002;143(5):1741-1747.
72. Kir S, Beddow SA, Samuel VT, et al. FGF19 as a postprandial, insulin-independent activator of hepatic protein and glycogen synthesis. *Science (80-)*. 2011;331(6024):1621-1624.
73. Fu L, John LM, Adams SH, et al. Fibroblast growth factor 19 increases metabolic rate and reverses dietary and leptin-deficient diabetes. *Endocrinology*. 2004;145(6):2594-2603.
74. Li F, Jiang C, Krausz KW, et al. Microbiome remodelling leads to inhibition of intestinal farnesoid X receptor signalling and decreased obesity. *Nat Commun*. 2013;4(May):2384.
75. Jiang C, Xie C, Li F. Intestinal farnesoid X receptor signaling promotes nonalcoholic fatty liver disease. *J Clin Invest*. 2015;125(1):386-402.
76. Jiang C, Xie C, Lv Y, et al. Intestine-selective farnesoid X receptor inhibition improves obesity-related metabolic dysfunction. *Nat Commun*. 2015;6:10166.
77. Fang S, Suh JM, Reilly SM, et al. Intestinal FXR agonism promotes adipose tissue browning and reduces obesity and insulin resistance. *Nat Med*. 2015;21(2):159-165.
78. Pathak P, Xie C, Nichols RG, et al. Intestine farnesoid X receptor agonist and the gut microbiota activate G-protein bile acid receptor-1 signaling to improve metabolism. *Hepatology*. 2018;10.1002/hep.29857.
79. Rowland M, Gallagher CG, O'Laoide R, et al. Outcome in cystic fibrosis liver disease. *Am J Gastroenterol*. 2011;106(1):104-109.
80. Colombo C. Liver disease in cystic fibrosis. *Curr Opin Pulm Med*. 2007;13(6):529-536.
81. Cystic Fibrosis Foundation. Annual Data Report 2016 Cystic Fibrosis Foundation Patient Registry. 2017.
82. Van De Peppel IP, Bertolini A, Jonker JW, Bodewes FAJA, Verkade HJ. Diagnosis, follow-up and treatment of cystic fibrosis-related liver disease. *Curr Opin Pulm Med*. 2017;23(6):562-569.
83. Feranchak AP. CFTR: Actin(g) as a master regulator of cholangiocyte function.

- Hepatology*. 2017;1-36.
84. Durie PR, Kent G, Phillips MJ, Ackerley C a. Characteristic multiorgan pathology of cystic fibrosis in a long-living cystic fibrosis transmembrane regulator knockout murine model. *Am J Pathol*. 2004;164(4):1481-1493.
 85. Wilke M, Buijs-Offerman RM, Aarbiou J, et al. Mouse models of cystic fibrosis: Phenotypic analysis and research applications. *J Cyst Fibros*. 2011;10(SUPPL. 2):S152-S171.
 86. van Nieuwerk CMJ, Groen AK, Ottenhoff R, et al. The role of bile salt composition in liver pathology of *mdr2* (-/-) mice: differences Between Males and Females. *J Hepatol*. 1997;2(26):138-145.
 87. Ridlon JM, Kang D-J, Hylemon PB, Bajaj JS. Bile Acids and the Gut Microbiome. *Curr Opin Gastroenterol*. 2014;30(3):332-338.
 88. Naugler WE. Bile acid flux is necessary for normal liver regeneration. *PLoS One*. 2014;9(5):1-11.
 89. Zhang L, Wang YD, Chen WD, et al. Promotion of liver regeneration/repair by farnesoid X receptor in both liver and intestine in mice. *Hepatology*. 2012;56(6):2336-2343.
 90. Uriarte I, Fernandez-Barrena MG, Monte MJ, et al. Identification of fibroblast growth factor 15 as a novel mediator of liver regeneration and its application in the prevention of post-resection liver failure in mice. *Gut*. 2013;62(6):899-910.
 91. Huang W, Ma K, Zhang J, et al. Nuclear receptor-dependent bile acid signaling is required for normal liver regeneration. *Science (80-)*. 2006;312(5771):233-236.
 92. Cheng K, Ashby D, Smyth RL. Ursodeoxycholic acid for cystic fibrosis-related liver disease. *Cochrane Database Syst Rev*. 2017;12(9):CD000222.
 93. Beuers U, Trauner M, Jansen P, Poupon R. New paradigms in the treatment of hepatic cholestasis: From UDCA to FXR, PXR and beyond. *J Hepatol*. 2015;62(S1):S25-S37.
 94. Quintero P, Pizarro M, Solís N, et al. Bile acid supplementation improves established liver steatosis in obese mice independently of glucagon-like peptide-1 secretion. *J Physiol Biochem*. 2014;70(3):667-674.
 95. Tsuchida T, Shiraishi M, Ohta T, Sakai K, Ishii S. Ursodeoxycholic acid improves insulin sensitivity and hepatic steatosis by inducing the excretion of hepatic lipids in high-fat diet-fed KK-A y mice. *Metabolism*. 2012;61(7):944-953.
 96. Fujita K, Iguchi Y, Une M, Watanabe S. Ursodeoxycholic Acid Suppresses Lipogenesis in Mouse Liver: Possible Role of the Decrease in β -Muricholic Acid, a Farnesoid X Receptor Antagonist. *Lipids*. 2017;52(4):335-344.
 97. Mueller M, Thorell A, Claudel T, et al. Ursodeoxycholic acid exerts farnesoid X receptor-antagonistic effects on bile acid and lipid metabolism in morbid obesity. *J Hepatol*. 2015;62:1398-1404.
 98. Zhang Y, LaCerte C, Kansra S, Jackson JP, Brouwer KR, Edwards JE. Comparative potency of obeticholic acid and natural bile acids on FXR in hepatic and intestinal in vitro cell models. *Pharmacol Res Perspect*. 2017;5(6):1-10.

99. Dilger K, Hohenester S, Winkler-Budenhofer U, et al. Effect of ursodeoxycholic acid on bile acid profiles and intestinal detoxification machinery in primary biliary cirrhosis and health. *J Hepatol.* 2012;57(1):133-140.
100. Liu Y, Binz J, Numerick MJ, et al. Hepatoprotection by the farnesoid X receptor agonist GW4064 in rat models of intra- and extrahepatic cholestasis. *J Clin Invest.* 2003;112(11):1678-1687.
101. Modica S, Petruzzelli M, Bellafante E, et al. Selective activation of nuclear bile acid receptor FXR in the intestine protects mice against cholestasis. *Gastroenterology.* 2012;142(2):355-365.e4.
102. Lin BC, Desnoyers LR. FGF19 and cancer. *Adv Exp Med Biol.* 2012;728:183-194.
103. Luo J, Ko B, Elliott M, et al. Liver Disease: A nontumorigenic variant of FGF19 treats cholestatic liver diseases. *Sci Transl Med.* 2014;6(247):1-12.
104. Zhou M, Learned RM, Rossi SJ, Depaoli AM, Tian H, Ling L. Engineered fibroblast growth factor 19 reduces liver injury and resolves sclerosing cholangitis in Mdr2-deficient mice. *Hepatology.* 2016;63(3):914-929.
105. Olivier AK, Gibson-Corley KN, Meyerholz DK. Animal models of cystic fibrosis: gastrointestinal, pancreatic, and hepatobiliary disease and pathophysiology. *Am J Physiol Gastrointest Liver Physiol.* 2015;308(6):G459-71.
106. Lavelle GM, White MM, Browne N, McElvaney NG, Reeves EP. Animal Models of Cystic Fibrosis Pathology: Phenotypic Parallels and Divergences. *Biomed Res Int.* 2016;2016:5258727.
107. Rogers CS, Stoltz DA, Meyerholz DK, et al. Disruption of the CFTR gene produces a model of cystic fibrosis in newborn pigs. *Science (80-).* 2008;321(September):1837-1842.
108. Meyerholz DK, Stoltz DA, Pezzulo AA, Welsh MJ. Pathology of gastrointestinal organs in a porcine model of cystic fibrosis. *Am J Pathol.* 2010;176(3):1377-1389.
109. Arslan N. Obesity, fatty liver disease and intestinal microbiota. *World J Gastroenterol.* 2014;20(44):16452-16463.
110. Wahlström A, Sayin SI, Marschall H-U, Bäckhed F. Intestinal Crosstalk between Bile Acids and Microbiota and Its Impact on Host Metabolism. *Cell Metab.* 2016:1-10.
111. De Lisle RC, Roach E a, Norkina O. Eradication of small intestinal bacterial overgrowth in the cystic fibrosis mouse reduces mucus accumulation. *J Pediatr Gastroenterol Nutr.* 2006;42(January 2006):46-52.
112. del Campo R, Garriga M, Pérez-Aragón A, et al. Improvement of digestive health and reduction in proteobacterial populations in the gut microbiota of cystic fibrosis patients using a *Lactobacillus reuteri* probiotic preparation: A double blind prospective study. *J Cyst Fibros.* 2014;13(6):716-722.
113. Bruzzese E, Callegari ML, Raia V, et al. Disrupted intestinal microbiota and intestinal inflammation in children with cystic fibrosis and its restoration with *Lactobacillus* gg: A randomised clinical trial. *PLoS One.* 2014;9(2):1-12.
114. Bruzzese E, Raia V, Gaudiello G, et al. Intestinal inflammation is a frequent feature of cystic fibrosis and is reduced by probiotic administration. *Aliment Pharmacol Ther.*

- 2004;20(7):813-819.
115. Bruzzese E, Raia V, Ruberto E, et al. Lack of efficacy of Lactobacillus GG in reducing pulmonary exacerbations and hospital admissions in children with cystic fibrosis: A randomised placebo controlled trial. *J Cyst Fibros.* 2018;17(3):375-382.
 116. Rovner AJ, Schall JI, Mondick JT, Zhuang H, Mascarenhas MR. Delayed small bowel transit in children with cystic fibrosis and pancreatic insufficiency. *J Pediatr Gastroenterol Nutr.* 2013;57:81-84.
 117. Hedsund C, Gregersen T, Joensson IM, Olesen H V, Krogh K. Gastrointestinal transit times and motility in patients with cystic fibrosis. *Scand J Gastroenterol.* 2012;47(8-9):920-926.
 118. Gelfond D, Ma C, Semler J, Borowitz D. Intestinal pH and gastrointestinal transit profiles in cystic fibrosis patients measured by wireless motility capsule. *Dig Dis Sci.* 2013;58(8):2275-2281.
 119. van der Wulp MYM, Cuperus FJC, Stellaard F, et al. Laxative treatment with polyethylene glycol does not affect lipid absorption in rats. *J Pediatr Gastroenterol Nutr.* 2012;55(4):1.
 120. De Lisle RC, Roach E, Jansson K. Effects of laxative and N-acetylcysteine on mucus accumulation, bacterial load, transit, and inflammation in the cystic fibrosis mouse small intestine. *Am J Physiol Gastrointest Liver Physiol.* 2007;293(3):G577-84.
 121. Fridge JL, Conrad C, Gerson L, Castillo RO, Cox K. Risk factors for small bowel bacterial overgrowth in cystic fibrosis. *J Pediatr Gastroenterol Nutr.* 2007;44:212-218.

