Neurobehavioral teratogenicity of perfluorinated alkyls in an avian model☆

Adi Pinkas a, Theodore A. Slotkin b, Yael Brick-Turin a, Eddy A. Van der Zee c, Joseph Yanai a,b,∗

a The Ross Laboratory for Studies in Neural Birth Defects, Department of Medical Neurobiology, Institute for Medical Research—Israel–Canada, The Hebrew University, Hadassah Medical School, Box 12272, 91120 Jerusalem, Israel
b Department of Pharmacology and Cancer Biology, Duke University Medical Center, Durham, NC 27710, USA
c Department of Molecular Neurobiology, University of Groningen, PO Box 14 9750 AA Haren, The Netherlands

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Perfluorinated alkyls are widely-used agents that accumulate in ecosystems and organisms because of their slow rate of degradation. There is increasing concern that these agents may be developmental neurotoxicants and the present study was designed to develop an avian model for the neurobehavioral teratogenicity of perfluorooctanoic acid (PFOA) and perfluorooctane sulfonate (PFOS). Fertilized chicken eggs were injected with 5 or 10 mg/kg of either compound on incubation day 0. On the day of hatching, imprinting behavior was impaired by both compounds. We then explored underlying mechanisms involving the targeting of protein kinase C (PKC) isoforms (α, β, γ) in the intermedial part of the hyperstriatum ventrale, the region most closely associated with imprinting. With PFOA exposure, cytosolic PKC concentrations were significantly elevated for all three isoforms; despite the overall increase in PKC expression, membrane-associated PKC was unaffected, indicating a defect in PKC translocation. In contrast, PFOS exposure evoked a significant decrease in cytosolic PKC, primarily for the β and γ isoforms, but again without a corresponding change in membrane-associated enzyme; this likely partial, compensatory increases in translocation to offset the net PKC deficiency. Our studies indicate that perfluorinated alkyls are indeed developmental neurotoxicants that affect posthatch cognitive performance but that the underlying synaptic mechanisms may differ substantially among the various members of this class of compounds, setting the stage for disparate outcomes later in life.

1. Introduction

Perfluorooctanoic acid (PFOA) and perfluorooctane sulfonate (PFOS) are prominent members of the family of perfluorinated alkyls (PFA). These compounds are widely used for manufacturing non-stick coatings and stain repellents in fabrics, as well as in food-packaging and lubricants. PFAs are very poorly biodegradable and are accumulating in humans and ecosystems worldwide [17], extending even to animals in remote locations [10]. Thus, despite recent efforts to reduce or phase out PFA production [9], these agents will persist in the environment [20,26].

Parallel to PFA bioaccumulation, toxicological studies in adult rats given high doses have shown damage to several internal organs, including the liver, kidneys and heart [43], preceded by substantial pathological changes in gene expression patterns [12]; parallel findings have been reported in zebra fish [29] and chicks [42]. In developing rodents, prenatal exposure to perfluorooctane sulfonate (PFOS) compromises survival rates and delays general growth and development [35]. In chicks, prenatal exposure to PFOS or perfluorooctanoic acid (PFOA) elicits reductions in hatchability, interference with pigmentation and pathological changes in gene expression patterns [12]; parallel findings have been reported in zebra fish [29] and chicks [42]. In our earlier work, we pointed out that the biochemical characteristics that underlie plumage pigmentation in the chick are also critical for nervous system development [38], suggesting that the PFAs might be developmental neurotoxicants, a prediction we confirmed with in vitro models of neuronal development [30].

Studies in rodents similarly point to behavioral deficits in adulthood after neonatal exposure to PFOA or PFOS [16], reflecting an adverse impact on functioning of critical neurochemical events required for synaptic development and function [15]. Nevertheless, rodent models have inherent methodological shortcomings from confounding maternal

⁎ Corresponding author. The Ross Laboratory for Studies in Neural Birth Defects, Department of Medical Neurobiology, Institute for Medical Research—Israel–Canada, The Hebrew University, Hadassah Medical School, Box 12272, 91120 Jerusalem, Israel.
Tel.: +972 2 675 8439; fax: +972 2 675 8443.
E-mail address: yanai@md.huji.ac.il (J. Yanai).
factors, notably maternal care and mother–offspring interaction [27,32], and the so-called “litter effect,” [33], where animals within a given litter are not independent of each other. Obviously, in the chick model, these limitations do not apply, since there is no maternal interaction and every individual represents an independent sample; further, because the chick is more mature at hatching than are newborn rats or mice, cognitive behavior, in the form of imprinting performance, can be evaluated immediately, prior to any potential impact of prenatal treatment on feeding or other indirect contributors to behavioral abnormalities. A newborn chick tends to follow the first object it sees after hatching [21] and can thus be imprint upon an artificial object, which then becomes a suitable subject for studying the effect of prenatal treatments on imprinting behavior [5,8,31]. We have already demonstrated how this parallels hippocampus-related visuospatial cognitive performance in rodents [39], involving in the chick the left intermedial part of the hyperstriatum ventrale (IMHV) [5,13]; specifically, cholinergic innervation in the left IMHV controls the chicks’ perception of the imprinting stimulus [22,36]. Given the postulated involvement of cholinergic deficits in the long-term neurobehavioral effects of neonatal PFOA and PFOS exposure in rodents [16], in the present study we evaluated whether these agents have an adverse effect on imprinting performance in the chick model.

In addition to behavioral assessments, we also evaluated the impact of PFOA and PFOS exposure on the concentration of protein kinase C (PKC) isoforms, which play an integral mechanistic role in transducing cholinergic input involved in learning and memory. Specifically, the translocation of PKC from the cytoplasm to the membrane is required for imprinting [3,4,25,37]. In our studies, we obtained the brain samples immediately after the imprinting test so as to examine the ability of the imprinting stimulus to evoke PKC translocation. Indeed, in earlier work with other neurotoxicants known to target cholinergic function, we confirmed the direct relationship between adverse effects on PKC translocation and imprinting performance [14]. Here, we identified different patterns of effects of PFOA and PFOS on PKC-related mechanisms in association with impaired imprinting behavior.

2. Methods

2.1. Teratogen treatments

Fertile heterogeneous stock eggs (60 ± 3 g) of the Cobb I chicken broiler strain (Gallus gallus domesticus) were obtained from a commercial source and placed in an incubator. To introduce substances, a hole was drilled in the chorioallantois end (pointed end) of the shell and then conducted at doses of 5–0 mg/kg of egg for PFOA and PFOS (Sigma, Israel) were then administered before incubation, i.e. on incubation day 0 (the PFOA in its free acid form and the PFOS in its salt form, due to dilution difficulties). The hole was then covered with an adhesive silicone glue cap (Medical Type Savfahan, GA) at 37.5 °C and 50% humidity, and were candled on incubation days 5, 12 and 19. At hatching, morphological and functional individuality represents an independent sample; further, because the chick is more mature at hatching than are newborn rats or mice, cognitive behavior, in the form of imprinting performance, can be evaluated immediately, prior to any potential impact of prenatal treatment on feeding or other indirect contributors to behavioral abnormalities. A newborn chick tends to follow the first object it sees after hatching [21] and can thus be imprint upon an artificial object, which then becomes a suitable subject for studying the effect of prenatal treatments on imprinting behavior [5,8,31]. We have already demonstrated how this parallels hippocampus-related visuospatial cognitive performance in rodents [39], involving in the chick the left intermedial part of the hyperstriatum ventrale (IMHV) [5,13]; specifically, cholinergic innervation in the left IMHV controls the chicks’ perception of the imprinting stimulus [22,36]. Given the postulated involvement of cholinergic deficits in the long-term neurobehavioral effects of neonatal PFOA and PFOS exposure in rodents [16], in the present study we evaluated whether these agents have an adverse effect on imprinting performance in the chick model.

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PKCγ [37], was specifically produced for this study (E.A. Van der Zee). IgG HRP-conjugated secondary antibody (Bio-Rad, Hercules, CA) was used as described earlier [28,41]. As a calibration standard, every gel contained aliquots of a pooled homogenate combined from all the samples. This approach was preferable to using a "housekeeping" protein such as α-tubulin or β-actin, since neuroteratogens are known to influence neuromorphological development and thus alter expression of these cytoskeleton proteins, unrelated to the neurobehavioral effects linked to specific cell signaling pathways [1].

2.4. Data analysis

Data are presented as means and standard errors, with differences between treatments established by multivariate ANOVA, followed by Fisher’s Least Significant Difference test for post hoc comparisons between groups. Significance for all tests was assumed at \( p < 0.05 \). For convenience, some results are presented as the percentage change from control values or as the percentage stimulation over basal PKC activity; however, statistical tests were always performed on the original data. Given the two different vehicles used to dissolve PFOA and PFOS, there were two sets of corresponding controls; further, the studies involved separate clutches of eggs for each of the PFAs, and accordingly, each treatment was compared only to its matched control group.

3. Results

On incubation day 19, fewer of the PFA-exposed eggs contained developing embryos as compared to the control groups, with reductions ranging from 30% to 50% (Fig. 1a). In addition, there were even further reductions in hatchability. However, among the chicks that hatched, there was no difference in weight (Fig. 1b) nor in the scoring for general morphology and function (Fig. 1c).

Control chicks showed typically-high imprinting scores [14] of about 0.8 (Fig. 2a). Chicks exposed to either of the PFAs lost about half their imprinting ability, although they still showed significant imprinting above the baseline score of 0.5. In general, PFOA had a greater effect than did PFOS. The effects did not show any sex preference (no interaction of treatment × sex, not shown), so the effects of males and females are shown combined. The effects on imprinting were not secondary to loss of motor function, since overall activity levels in the imprinting apparatus were not significantly affected (Fig. 2b).

The concentrations of the PKC isoforms in subcellular fractions of the left IMHV are presented as the percent change from the corresponding control levels (Fig. 3). Exposure to PFOA produced an overall increase in cytosolic PKC involving all three isoforms but there was no corresponding change in membrane-associated PKC (Fig. 3a). In contrast, PFOS elicited significant overall reductions in cytosolic PKC with a distinct isoform preference of γ-β-α; there were no significant effects on membrane-associated PKC.

4. Discussion

In the present study, embryonic exposure to PFOA and PFOS induced posthatch deficits in imprinting behavior in association with alterations in the concentrations of PKC isoforms within the left IMHV, the brain region in which imprinting is consolidated. Whereas the behavioral deficits were similar for both PFAs the changes at the molecular level differed, although as discussed below, the PKC changes for both agents are consistent with impaired expression and function of these key intermediates. Because the chick model, unlike mammalian models, does not involve maternal effects, our studies indicate that PFAs evoke neurobehavioral teratogenesis through direct effects on the developing brain, rather than through indirect compromise of maternal function or maternal–neonatal interactions; this reinforces conclusions drawn from in vitro models that similarly point to direct PFA actions on neuronal cell replication and differentiation [30].

In our previous studies with avian and rodent models, prenatal exposure to teratogens from different classes of compounds, all evoked deficits in the ability of PKC isoforms to translocate from the cytosol to the membrane, in association with their ability to impair cognitive performance: phenobarbital in the mouse [6]; heroin in the mouse [18] and chick [14]; and nicotine and the organophosphates, chlorpyrifos and sarin in the chick [14,40]. We were able to establish a mechanistic connection between the molecular and behavioral effects by administering targeted treatments that reversed the deficits [7,18,28]. Similarly, in the present study with PFOA and PFOS, we found alterations in PKC expression and translocation paralleling the behavioral deficits. However, the patterns seen with the PFAs were distinct from those of the other neuroteratogens. The previously-
studied agents targeted primarily PKCγ, and to a lesser extent, PKCβ, while largely sparing PKCα, and the deficits were much larger with respect to the membrane-associated fraction as distinct from cytosolic PKC [7,14,18,41]. Here, PFOA evoked large increases in the cytosolic fraction without a corresponding effect on membrane-associated PKC, and with little or no selectivity for any of the PKC subtypes. If the effectiveness of translocation was unaffected by PFOA, then the membrane-bound PKC should have increased by exactly the same proportion as for the cytosol; the fact that this did not occur again points to a defect in the ability to translocate PKC, similar to that seen with the other neuroteratogens that produce the same behavioral outcome. Nevertheless, the point remains that PFOA affects a wider spectrum of PKC subtypes than the other agents, and has unique effects on cytosolic PKC; PFOA thus may have a wider spectrum of adverse behavioral effects, a conclusion that clearly needs to be evaluated in future work.

Although both PFOA and PFOS evoked deficits in imprinting performance, the effects of PFOS on PKC isofoms differed from those of PFOA. Instead of causing a global increase in cytosolic PKC concentrations, PFOS elicited significant deficits; as with PFOA, these were unaccompanied by a corresponding change in the membrane-associated PKC fraction. Thus, PFOS decreased the cytosolic pool of PKC available for translocation but there was compensation so that the relative proportion of membrane translocation was higher, partially offsetting the drop in total PKC. If this difference from PFOA is mechanistically connected to the behavioral outcomes, then it would be expected that PFOS would produce somewhat smaller deficits, which was in fact the outcome that we observed. By extension, we would again predict fewer long-term behavioral deficits with PFOS than with PFOA. The idea that different PFAs might produce disparate neurodevelopmental outcomes is bolstered by recent in vitro findings demonstrating divergent effects on neuronal cell differentiation, particularly involving the emergence of the acetylcholine phenotype, the neurotransmitter most closely associated with imprinting [30]. In any case, the implication is clear that PFAs cannot simply be regarded as compounds that will all act similarly on the developing nervous system but rather, they are likely to differ substantially in their net effects.

Although both PFOA and PFOS reduced hatchability, this factor does not translate well from avian to mammalian species. Avian embryos are extremely sensitive to even minor environmental perturbations, including vibration and movement, small changes in temperature and humidity, season, and clutch-to-clutch differences. In the present study, 20% of the control eggs were infertile or failed to hatch, a typical percentage for commercial hatcheries. In mammals, fetal resorption or intrauterine growth retardation occurs only at toxicant exposures that are considered hallmarks of high-dose toxicity but the same is not so for avian species, where effects on hatching often involve lower exposures than those required for neurobehavioral deficits. Thus, although our results provide interpretable cross-species results for neurodevelopmental endpoints, the nonspecific measures, such as hatching rate or malformations are of limited utility in comparing benchmarks for toxicant exposures. Another unique feature of the avian model is the fact that the egg is a closed system. Accordingly, we did not assess the posthatch levels of
PFOA and PFOS in the present study, but given that fact, combined with the resistance of these agents to chemical and biochemical breakdown, the body burden is likely to be the same as that given to the egg. Nevertheless, it would also be useful to assess the posthatch persistence of PFOA and PFOS and the relationship to more lasting neurobehavioral effects.

Production of PFOS in the U.S. was voluntarily discontinued by the manufacturer in 2002, whereas PFOA continued to be manufactured for several additional years [20,26]. Nevertheless, new members of the PFA class continue to emerge [2] and the legacy of the previously-used compounds will continue because of their long half-lives [20,26]. Our studies provide some of the first evidence that these compounds act directly as neuroteratogens and that the various members of the PFA class may differ in their ability to disrupt nervous system development.

Conflict of interest

No author has any conflict of interest to disclose.

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References