Molecular mechanism of temperature sensing by the circadian clock of *Neurospora crassa*

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Expression levels and ratios of the long (l) and short (s) isoforms of the *Neurospora* circadian clock protein FREQUENCY (FRQ) are crucial for temperature compensation of circadian rhythms. We show that the ratio of l-FRQ versus s-FRQ is regulated by thermosensitive splicing of intron 6 of *frq*, a process removing the translation initiation site of l-FRQ. Thermosensitivity is due to inefficient recognition of nonconsensus splice sites at elevated temperature. The temperature-dependent accumulation of FRQ relative to bulk protein is controlled at the level of translation. The 5′-UTR of *frq* RNA contains six upstream open reading frames (uORFs) that are in nonconsensus context for translation initiation. Thermosensitive trapping of scanning ribosomes at the uORFs leads to reduced translation of the main ORF and allows adjustment of FRQ levels according to ambient temperature.

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Circadian clocks are cellular oscillators that organize temporal expression of large numbers of genes in many organisms. They are synchronized to the 24-h cycle of earth rotation, and changes in light and ambient temperature are major natural cues (zeitgebers) for entrainment. Without zeitgebers, circadian clocks oscillate with their endogenous free-running periods, which often deviate from 24 h. In fungi, plants, and animals, circadian rhythmicity depends on phosphorylation (Stanewsky 2003; Dunlap and Loros 2004; Gachon et al. 2004). The role of temperature in regulation of circadian rhythmicity is complex and not fully understood. In general, temperature shifts result in the resetting of circadian clocks, i.e., affect the phase of the circadian oscillation. However, while biochemical processes are generally temperature dependent, the free-running periods of circadian rhythms are precise and efficiently temperature compensated with a Q10 close to unity (Izumo et al. 2003; Tsuchiya et al. 2003; Dunlap and Loros 2004). Yet, temperature affects the circadian clock, allowing its adaptation to seasonal variation of day length.

In *Drosophila*, thermostressive splicing in the 3′-UTR of the clock gene *period (per)* leads to phase advanced accumulation of *per* RNA on cold days, which is crucial for seasonal adaptation of circadian locomotor activity (Majercak et al. 1999). Thermosensitive splicing is affected by light, the clock, and phospholipase C (Collins et al. 2004; Majercak et al. 2004).

In *Neurospora*, FREQUENCY (FRQ) is a central component of the circadian clock (Dunlap and Loros 2004). *frq* RNA is rhythmically expressed and synthesized with a 5′-UTR containing six upstream open reading frames (uORFs) (Liu et al. 1997). A large [l] and a small [s] isoform of FRQ are translated from a common primary transcript. It has been shown that the abundance and ratio of l-FRQ versus s-FRQ are crucial for robust free-running rhythmicity, as well as for temperature compensation of the circadian clock (Garceau et al. 1997; Liu et al. 1997). l-FRQ versus s-FRQ is expressed in a temperature-dependent fashion. Expression of l-FRQ relative to bulk protein increases with increasing temperature, while expression of s-FRQ is essentially temperature independent. The underlying molecular mechanisms by which *Neurospora* senses temperature to adjust levels and ratio of l-FRQ versus s-FRQ are not known. It has been proposed that ribosomes scan more efficiently through the AUG codon of l-FRQ at low temperature and thus favor initiation at the downstream in-frame AUG of s-FRQ (Liu et al. 1997). No mechanism has been proposed for the steep temperature dependence of l-FRQ abundance relative to bulk protein.

Here we have analyzed the molecular mechanisms underlying temperature-dependent levels and ratios of l-FRQ versus s-FRQ. We show that the ratio of l-FRQ versus s-FRQ is regulated by thermosensitive splicing of intron 6 of *frq* RNA. Thermosensitivity is a direct consequence of inefficient recognition of nonconsensus splice sites by the splicing machinery at elevated temperature. Preferential splicing of I-6 at low temperature results in excision of the translation initiation site of l-FRQ, such that the I-6 spliced messages encode only s-FRQ. Interconnected with thermosensitive splicing is a temperature-dependent translational control mechanism that is crucial for the increase in FRQ relative to bulk protein with increasing temperature. The Kozak sequences of the uORFs in the 5′-UTR of *frq* are in nonconsensus context for translation initiation. As a consequence, the uORFs are more efficiently translated at low temperature, leading to a reduction of translation initiation at the downstream main ORF encoding l- and s-FRQ.
FRQ. The thermosensitive trapping of scanning ribosome at of the uORFs allows adjustment of FRQ levels according to ambient temperature.

**Results and Discussion**

Transcription initiation sites of the frq promoter were determined by cap-specific 5’-RNA ligase-mediated rapid amplification of cDNA ends (5’RLM-RACE). A number of different sites were identified ∼1.5 kb upstream of the frq ORF, the major one at −1519 [Fig. 1A]. This site is 156 bp downstream of the proximal light-responsive element of the frq promoter [Froehlich et al. 2002]. A putative CCAAT box [Smail and Kadonga 2003] is located at −201. No obvious TATA box was identified.

Since the 5’-UTR of the primary frq transcript contains a number of putative splice sites, cDNA was analyzed by polymerase chain reaction (PCR). Six different introns, giving rise to seven splice-isoforms of frq RNA, were identified [Fig. 1B; Supplementary Table 1]. Interestingly, splicing of I-6 removes the translation initiation site of l-FRQ. The resulting frq RNA species encode s-FRQ specifically. The Lariat sequence of I-6 deviates considerably from consensus (Supplementary Table 1), and only a fraction of frq RNA spliced at I-6 (spliced/total frq RNA) was determined. [B] Protein extracts prepared from Neurospora that was grown at the indicated temperatures in LL were treated with alkaline phosphatase. Samples were analyzed by immunoblotting with α-FRQ, recognizing l-FRQ and s-FRQ [top] and quantified by densitometry [bottom]. l-FRQ [black bars] and s-FRQ [gray bars] are shown. [C] Cultures were shifted from 15°C to 35°C [dark bars] or from 35°C to 15°C [white bars]. The fraction of spliced I-6 was determined. [D] Thermosensitive splicing of I-6 is independent of functional FRQ. frqRNA was grown in LL at the indicated temperatures. RNA was prepared, and splicing of I-6 was determined.

Figure 1. The 5’-UTR of frq RNA is spliced in a complex fashion. (A) The frq gene is schematically outlined. Distal light-responsive elements (dLRE) and proximal light-responsive elements (pLRE) are indicated by hatched boxes [Froehlich et al. 2002]. The major transcription initiation site is indicated by an arrow. The black and the gray + black areas correspond to s-FRQ and l-FRQ, respectively. (Bottom) Major and minor transcription initiation sites. Eleven independent clones of cap-specific RLM-RACE products were sequenced. Numbers above the sequence indicate the 5’-ends of these clones. The major initiation site [seven clones] corresponds to position −1519. Initiator consensus sequences [Inr] for transcription [Smail and Kadonga 2003] are indicated below the sequence. [Y indicates C or T; N indicates any nucleotide]. (B) Schematic outline of the 5’-UTR of frq. Splice donor (D) and acceptor (A) sites and translation initiation sites for l-FRQ [AUG1] and s-FRQ [AUG3] are shown. AUG2 is not used for initiation [Liu et al. 1997]. Dark-gray arrows indicate uORF1 to uORF6 in the 5’-UTR and ORF7 overlaps the AUG of s-FRQ. The arrows below the scheme indicate oligonucleotide primers for amplification of cDNA. The outlined unspliced [I] and spliced [II-VIII] frq RNA species [left panel] were amplified by reverse transcription and PCR [right panel] and identified by sequencing [black arrowheads] or by diagnostic PCR [white arrowheads]. RNA was prepared from cells grown at 25°C in LL. (Std) DNA size standard, (gDNA) genomic DNA.

Figure 2. I-6 is spliced in a temperature-dependent fashion. (A) Neurospora was grown at the indicated temperatures in LL, and RNA was prepared and quantified by RT–PCR [see Materials and Methods]. [Left] Total frq RNA was determined. frq RNA/actin RNA at 35°C was set equal to 1. [Right] The fraction [%] of frq RNA spliced at I-6 (spliced/total frq RNA) was determined. [B] Protein extracts prepared from Neurospora that was grown at the indicated temperatures in LL were treated with alkaline phosphatase. Samples were analyzed by immunoblotting with α-FRQ, recognizing l-FRQ and s-FRQ [top] and quantified by densitometry [bottom]. l-FRQ [black bars] and s-FRQ [gray bars] are shown. [C] Cultures were shifted from 15°C to 35°C [dark bars] or from 35°C to 15°C [white bars]. The fraction of spliced I-6 was determined. [D] Thermosensitive splicing of I-6 is independent of functional FRQ. frqRNA was grown in LL at the indicated temperatures. RNA was prepared, and splicing of I-6 was determined.
from 28% to 7% and reached a value <3% after 4 h (Fig. 2C). Correspondingly, upon shift from 35°C to 15°C, the fraction of spliced I-6 increased rapidly from ~3% to 18% after 30 min and to 30% after 4 h (Fig. 2C). The fast adaptation of frq RNA splicing to ambient temperature suggests that the process may not require de novo protein synthesis.

The mutant frq<sup>o</sup> allele expresses a full-size RNA that encodes (due to a premature stop) a nonfunctional, C-terminally truncated protein (Aronson et al. 1994). I-6 of frq<sup>o</sup> RNA was spliced in a temperature-dependent fashion (Fig. 2D), demonstrating that FRQ is not required for this function.

Next, splice sites of I-6 were optimized toward consensus [I-6<sup>opt</sup>] and mutagenized [I-6<sup>mut</sup>] to nonsplice sites (Supplementary Table 1) by site-directed mutagenesis, and frq RNA and FRQ were analyzed. I-6<sup>opt</sup> was efficiently spliced in a temperature-independent fashion (Fig. 3A). s-FRQ was synthesized at all temperatures, and only trace amounts of l-FRQ were detected (Fig. 3B). In contrast, I-6<sup>mut</sup> was not spliced (Fig. 3A). l-FRQ was synthesized, and little s-FRQ was detected (Fig. 3B). Thus, in frq<sup>o</sup>, s-FRQ is primarily synthesized from frq RNA species spliced at I-6.

How does splicing of I-6 affect temperature compensation of the circadian clock? The free-running period of Neurospora can be determined by analyzing the conidiation rhythm (banding) on race tubes. Banding can be observed most clearly between 22°C and 28°C. The free-running period of wild type decreases only slightly between 22°C and 28°C (Fig. 3D; Supplementary Fig. 1), demonstrating that the clock is well temperature compensated. The period of the I-6<sup>mut</sup> strain decreased steeper compared with wild type, while the period of the I-6<sup>opt</sup> strain increased. The data suggest that l-FRQ tends to shorten the period while s-FRQ causes period lengthening.

The data suggest that the molecular basis for the temperature-dependent splicing of I-6 is provided by the interaction of the splicing machinery with nonconsensus splice sites, which are more efficiently recognized at low temperature.

To test this hypothesis, we inserted into the frq ORF I-8 of the period gene [dmpi8], which is spliced in a thermosensitive fashion in Drosophila (Majercak et al. 1999). The chimeric frq-dmpi8 allele was expressed at different temperatures, and splicing was analyzed by qualitative RT-PCR (Fig. 3E). Like I-6, dmpi8 was also spliced in a thermosensitive fashion.

It should be pointed out that the ratio of l-FRQ versus s-FRQ correlates with the fraction of spliced I-6, while the increase of overall FRQ levels with increasing temperature is independent of I-6. Temperature-dependent expression of FRQ does not correlate with levels of frq RNA, which are temperature independent (Fig. 2, cf. A and B). Apparently, FRQ levels are regulated in a post-transcriptional fashion.

What is the molecular basis for the temperature dependence of FRQ accumulation? We constructed a strain where the long 5′-UTR of the frq gene was deleted (Fig. 4A). In this strain, FRQ levels increased only by a factor of 1.4 between 15°C and 35°C, suggesting that the 5′-UTR affects the temperature-dependent expression of FRQ. frq RNA levels increased in a similar fashion (Supplementary Fig. 2A). FRQ levels were significantly lower in frqΔUTR than in frq<sup>+</sup>, while RNA levels were similar. This may be due to a reduced efficiency of translation of frqΔUTR, which is often observed when the translation initiation site is close to the 5′-end of the RNA (Hinnebusch and Natarajan 2002; Vilela and McCarthy 2003).

We noticed that the uORFs in the 5′-UTR of frq are in nonconsensus context for translation initiation (Supplementary Table 2; Bruchez et al. 1993), suggesting that they are inefficiently translated. If translation initiation at the uORFs were more efficient at low temperature, trapping of ribosomes (Gaba et al. 2001; Hinnebusch and Natarajan 2002; Vilela and McCarthy 2003; Gebauer and Hentze 2004) could provide a mechanism for the temperature-dependent expression of the main ORF.

A major species of frq RNA is spliced at I-2 and contains uORF1 and uORF6 (Fig. 1). We changed the AUG codons of uORF1 and uORF6 to GUG [frq1/6<sup>mut</sup>] to prevent translation. Expression of FRQ was less tempera-
corresponding to I-2 was deleted (Fig. 4D). In one, the AUG codons of uORF1 and uORF6 were left unchanged (ΔI-2 frq1/6mut), and in the other, they were mutated (ΔI-2 frq1/6mut). In addition, the splice donor of I-6, which is located in uORF6, was deleted (Materials and Methods) resulting in an altered C terminus of uORF6. Expression of FRQ encoded by ΔI-2 frq1/6mut exhibited similar temperature dependence as frq* (Fig. 4D), whereas FRQ levels were significantly higher in ΔI-2 frq1/6mut and essentially independent of temperature.

Finally, we altered the translation initiation site of uORF6 toward consensus by site-directed mutagenesis. FRQ levels in the frq6mut strain were substantially lower and less temperature dependent than in a corresponding control strain (Fig. 4E). The codonization pattern of frq6mut was obscured at all three temperatures (Fig. 4F), presumably due to the low amplitude of FRQ oscillation.

Together, the data demonstrate that uORFs in the 5′-UTR restrict translation of the downstream ORF, in particular at low temperature. The temperature-dependent expression of FRQ is regulated by translation of uORFs, in particular uORF1 and uORF6, rather than by putative secondary structure elements in the 5′UTR. This suggests a model in which temperature dependence of expression levels and ratios of l-FRQ versus s-FRQ is regulated by two distinct thermosensitive processes. In both cases, temperature assessment appears to be a direct consequence of low-affinity interactions of general machinery for splicing and translation with nonconsensus lariat and Kozak sequences, respectively.

When the splice sites of I-6 were mutated toward consensus, I-6 was efficiently spliced in a temperature-independent fashion. Thus, the splice sites are cis-acting elements conferring a temperature-sensitive phenotype in regards to removal of I-6. However, while reduced splicing of I-6 at high temperature can be overcome by mutation [I-6mut], there may be another sequence element or factor that contributes to the inhibition of I-6 splicing at high temperatures.

Temperature-dependent splicing of I-6 is independent of FRQ and thus probably independent of the clock. Thermosensitive splicing was reported for I-8 (dmpi8) in the 3′-UTR of the circadian clock gene period [per] of Drosophila [Majercak et al. 1999]. The temperature-dependent mean splicing efficiency of dmpi8 appears to be also clock independent [Majercak et al. 2004]. Splice sites of I-6 [lariat] and dmpi8 [acceptor] deviate from consensus, and both introns are spliced in a temperature-dependent fashion in Neurospora. This suggests that the molecular basis of thermosensitive splicing may be similar in frq and per. Thermosensitive splicing of per is modulated by the circadian clock and by phospholipase C via unknown pathways [Collins et al. 2004, Majercak et al. 2004]. Preliminary results suggest that splicing of frq is also modulated by the clock (data not shown).

In contrast to splicing of I-6, splicing of dmpi8 does not lead to expression of different PER isoforms. Rather, the splicing event per se leads to more efficient maturation and accumulation of per RNA compared with splice-deficient and intron-less mutants [Majercak et al. 1999].

Figure 4. uORFs restrict translation of FRQ. [A] In frqΔUTR residues, 50–1477 of the 5′-UTR are deleted and an Ascl site is introduced. Western blot and densitometric quantification showing temperature-dependent expression of FRQ-encoded frqΔUTR. [B] Construction frq1/6mut. Start codons of uORF1 and uORF6 were altered by site-directed mutagenesis to GTG. Temperature-dependent expression of FRQ encoded by frq and frq1/6mut is shown. [C] The free-running period of frq1/6mut (squares) and wild-type (circles) control (frq) was determined on race tubes at 25°C and 28°C. [D] Construction of ΔI-2 frq1/6mut and ΔI-2 frq1/6mut. CDNA derived from I-2 spliced frq RNA was used as template for PCR reactions. Fragments including the translation initiation site of uORF1 and uORF6 were amplified by PCR and inserted into the Ascl site of frqΔUTR. Start codons of uORF1 and uORF6 were either left unchanged (ΔI-2 frq1/6mut) or changed to TTG and ACC (ΔI-2 frq1/6mut). In the constructs shown, the C terminus of uORF6 (...HKLLE) was changed to ...HKAGPSSSIAG. [E] frq6mut. The translation initiation site of uORF6 was altered toward consensus (CAACATGG), and expression of FRQ was measured at the indicated temperatures. The ratios of FRQ expressed by frq6mut vs. FRQ expressed by a corresponding wild-type frq allele are shown. [F] Condensation of the strain frq6mut. Race tubes were incubated in LL and incubated at 25°C in DD. After 24 h, they were transferred to the indicated temperatures.
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Preferential splicing of dmpi8 at low temperature leads to more efficient accumulation of per RNA. As a consequence, critical PER levels accumulate earlier, leading to a phase advanced locomotor activity of Drosophila at seasonally cold days. Whether and how thermosensitive splicing of frq I-6 affects seasonal adaptation of the Neurospora clock remains to be analyzed.

Recently, it has been reported that FRQ is in a complex with FRH, a homolog of an RNA helicase associated with the exosome [Cheng et al. 2005]. The exosome is a multisubunit complex involved in 3'–trimming of different RNA species, including mRNA [van Hoof et al. 2000]. However, splicing of I-6 is temperature dependent in frq-deficient strains (frq-), demonstrating that the FRQ/FRH complex is not required for thermosensitive splicing of frq RNA.

While the ratio of l-FRQ versus s-FRQ is regulated via thermosensitive splicing, the temperature-dependent accumulation of both isoforms of FRQ is regulated on translational level. uORFs in the 5’-UTR of frq affect expression of the main ORF encoding l- and s-FRQ in a temperature-dependent manner. When the AUGs of uORF1 and uORF6 are eliminated, temperature-dependent accumulation of FRQ is less pronounced; FRQ levels are elevated at low temperature but similar to wild type at high temperature. Thus, uORFs reduce FRQ expression at low temperature but have little effect at high temperature. The translation initiation sites of the uORFs deviate from consensus Kozak sequences. We suggest that the translation machinery may more efficiently initiate at the nonconsensus Kozak sequences at low temperature, while the uORFs are only inefficiently translated at elevated temperatures.

uORFs were reported to regulate translation of downstream located genes in a number of different ways. In the arg-2 gene of Neurospora and in the CPA1 gene of Saccharomyces cerevisiae, uORFs encode specific arginine-attenuator peptides that cause stalling of ribosomes [Gaba et al. 2001; Vilela and McCarthy 2003; Gebauer and Hentze 2004]. Although uORFs are also present in frq genes of filamentous fungi related to Neurospora, the encoded peptides are not conserved [data not shown]. It seems therefore unlikely that the uORF peptides cause ribosome stalling as in arg-2. The length of a uORF and the sequence context in the vicinity of the stop codon affect whether translating ribosomes terminate and fall off or resume scanning downstream of the uORF. The uORFs in frq are quite short, so ribosomes could, in principle, resume the scanning process after uORF translation. However, the sequences surrounding the stop codons of uORF1 and uORF6 are GC-rich, which is thought to favor dissociation and release of terminating ribosomes as reported for uORF4 in the GCN4 gene of S. cerevisiae [Gaba et al. 2001; Hinnebusch and Natarajan 2002; Vilela and McCarthy 2003; Gebauer and Hentze 2004]. Yet, the precise mechanism by which the uORFs regulate thermosensitive translation of the downstream frq ORF remains to be investigated.

When ratio of l-FRQ versus s-FRQ was manipulated via altered splicing, temperature compensation of the period length was affected. Although we cannot attribute distinct molecular functions to the FRQ isoforms, s-FRQ seems to cause period shortening and I-FRQ period lengthening [Fig. 3D]. When uORF1 and uORF6 were mutagenized, the levels of FRQ increased. The free-running rhythmicity of the frq1/cmut strain was lost in the low temperature range, and the period was lengthened at 28°C. This suggests that elevated FRQ levels maintain a prolonged repression of the frq locus and thus cause period lengthening. Reduction of FRQ expression by optimizing translation of uORF6 resulted in an obscured condensation pattern, suggesting that the amplitude of FRQ oscillation is crucial for a robust overt rhythmicity.

Why is the expression of FRQ then regulated in such a complex fashion? In Drosophila, rhythmicity of the clock is not affected when splicing of per is challenged. Rather, thermosensitive splicing of per regulates the adaptation of locomotor activity to seasonally warm and long versus cold and short days. Accordingly, splicing and translational control of frq may contribute to fine-tuning of clock functions in Neurospora, modulating, in particular, adaptation of the circadian clock to seasonal variations of photoperiod and temperature [Tan et al. 2004a,b].

How may levels and ratios of l-FRQ versus s-FRQ contribute to temperature compensation of period length? To reliably measure time on the molecular level, it might be necessary that FRQ-dependent biochemical processes proceed at a constant net rate at all temperatures. Since protein–protein interactions are generally temperature dependent, a decrease in complex stability at elevated temperature must be compensated. The increased expression of FRQ compared with bulk protein could account for such compensation. In addition, the preferential expression of l-FRQ over s-FRQ at high temperature could reflect accumulation of a FRQ isoform that is capable of forming more stable interactions. This is supported by the observation that Neurospora mutants that express low levels of s-FRQ but no l-FRQ are arrhythmic on race tubes at elevated temperatures but become rhythmic when s-FRQ is expressed at elevated levels [Liu et al. 1997].

We suggest a molecular mechanism for temperature-sensing resulting in adjustment of expression levels and ratios of l-FRQ versus s-FRQ [Fig. 5]: In the high-temperature range, ribosomes are scanning through the non-consensus AUGs of the uORFs in the 5’-UTR of frq and initiate translation at the main ORF, leading to efficient expression of FRQ. At low temperatures, scanning ribosomes initiate translation more efficiently at the uORFs, leading to a reduced translation of the downstream frq ORF. Accordingly, overall expression of FRQ (l + s) decreases with decreasing temperature. Superimposed on this translational control is the thermosensitive splicing.
of the nonconsensus I-6, which is more efficiently spliced at low temperatures. Since I-6 spliced frq RNA encodes specifically s-FRQ, the ratio of I-FRQ versus s-FRQ is decreasing with decreasing temperature. Thus, the strategy used to measure temperature on the molecular levels appears to be the use of nonconsensus signals for splicing and translation initiation. Since these signals form weaker interactions with the splicing machinery and with ribosomes, respectively, than corresponding consensus signals, they are more efficiently recognized at low temperatures and thus provide a molecular temperature sensor.

Materials and methods

Strains

Recombinant frq alleles were generated by site-directed mutagenesis (QuickChange XL Site-Directed Mutagenesis Kit; Stratagene) of the C11 fragment carrying the frq locus [McClung et al. 1989]; inserted into the his-3 locus of frq11, his-3, bd, and strains were grown in LL as described [Gör et al. 2001].

Donor [D4], Lariat, and Acceptor [A3] sequences of I-6 were either changed toward consensus or mutagenized to nonsense sites using the oligonucleotides shown in the Supplemental Material.

Construction of frq-dm180 is described in the Supplemental Material.

RNA analysis

cDNA was synthesized from 4 µg DNase-treated total RNA using the SuperScriptII RT-system (Invitrogen) and gene-specific reverse primers. frq cDNA and actin cDNA was detected by quantitative real-time PCR (ABI-Prism 7000, Applied Biosystems) using specific primers and Taq-Man probes [Gör et al. 2001]. Primers and probes for quantification of I-6 spliced and unspliced RNA are shown in Supplemental Material. At least triplicate reactions [25 µl] containing cDNA equivalent to 0.1 µg RNA were analyzed.

Protein analysis

Protein extracts were prepared as described [Gör et al. 2001]. For dephosphorylation extracts, 5 mg/mL were incubated with alkaline phosphatase (3000 U/mL) for 1 h at 37°C. Protein was separated by SDS-PAGE [7.5 %], and Western blotting was performed [Gör et al. 2001]. Affinity-purified polyclonal antibodies directed against the C-terminal 167-amino-acid residues of FRQ were used together with peroxidase-coupled secondary antibody. Enhanced chemiluminescence signals were detected with X-ray films and quantified by densitometry.

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References


