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Refining Susceptibility Loci of Chronic Obstructive Pulmonary Disease with Lung eqtls

Maxime Lamontagne1, Christian Couture1, Dirkje S. Postma2, Wim Timens3, Don D. Sin4,5, Peter D. Paré4,5, James C. Hogg4,6, David Nickle7, Michel Laviolette1, Yohan Bosse1,8*

1 Centre de recherche de l’Institut universitaire de cardiologie et de pneumologie de Québec, Québec, Canada, 2 University of Groningen, University Medical Center Groningen, Department of Pulmonology, GRIAC Research Institute, Groningen, The Netherlands, 3 University of Groningen, University Medical Center Groningen, Department of Pathology and Medical Biology, GRIAC Research Institute, Groningen, The Netherlands, 4 University of British Columbia James Hogg Research Center, Center for Heart and Lung Health, St. Paul’s Hospital, Vancouver, British Columbia, Canada, 5 Respiratory Division, Department of Medicine, The University of British Columbia, Vancouver, British Columbia, Canada, 6 Department of Pathology and Laboratory Medicine, The University of British Columbia, Vancouver, British Columbia, Canada, 7 Merck & Co. Inc., Rahway, New Jersey, United States of America, 8 Department of Molecular Medicine, Laval University, Québec, Canada

Abstract

Chronic obstructive pulmonary disease (COPD) is the fourth leading cause of mortality worldwide. Recent genome-wide association studies (GWAS) have identified robust susceptibility loci associated with COPD. However, the mechanisms mediating the risk conferred by these loci remain to be found. The goal of this study was to identify causal genes/variants within susceptibility loci associated with COPD. In the discovery cohort, genome-wide gene expression profiles of 500 non-tumor lung specimens were obtained from patients undergoing lung surgery. Blood-DNA from the same patients were genotyped for 1.2 million SNPs. Following genotyping and gene expression quality control filters, 409 samples were analyzed. Lung expression quantitative trait loci (eQTLs) were identified and overlaid onto three COPD susceptibility loci derived from GWAS; 4q31 (HHIP), 4q22 (FAM13A), and 19q13 (RAB4B, EGLN2, MIA, CYP2A6). Significant eQTLs were replicated in two independent datasets (n = 363 and 339). SNPs previously associated with COPD and lung function on 4q31 (rs1828591, rs13118928) were associated with the mRNA expression of HHIP. An association between mRNA expression level of FAM13A and SNP rs2045517 was detected at 4q22, but did not reach statistical significance. At 19q13, significant eQTLs were detected with EGLN2. In summary, this study supports HHIP, FAM13A, and EGLN2 as the most likely causal COPD genes on 4q31, 4q22, and 19q13, respectively. Strong lung eQTL SNPs identified in this study will need to be tested for association with COPD in case-control studies. Further functional studies will also be needed to understand the role of genes regulated by disease-related variants in COPD.

Introduction

Chronic obstructive pulmonary disease (COPD) is the fourth most common cause of death worldwide and is predicted to be the third leading cause of mortality in the world by the year 2030 [1]. COPD is a complex disease characterized by airflow obstruction that is not fully reversible [2]. Cigarette smoking is the most important cause of the rapid decline in pulmonary function that leads to COPD, but not all smokers develop the disease [3]. Moreover, there is familial aggregation of COPD suggesting a genetic contribution [4]. The only well-established genetic risk factors are inherited mutations causing α1-antitrypsin deficiency [5]. However, these mutations occur in only 1–5% of COPD patients [6].

The number of susceptibility genes for COPD is expanding rapidly with lists tabulated at 57 genes in 2009 [7] and at 144 genes in 2012 [8]. Recent genome-wide association studies (GWAS) have identified four susceptibility loci associated with COPD including 4q22 (FAM13A), 4q31 (HHIP), 15q25 (CHRNA5/CHRNB4) and 19q13 (RAB4B, EGLN2, MIA, CYP2A6) [9–11]. The causal genes and genetic mechanisms mediating the risk within these loci remain to be found.

The goal of this study is to identify lung expression quantitative trait loci (eQTL) within COPD susceptibility loci identified by GWAS. As part of the lung eQTL consortium, we have recently performed a genome-wide search for eQTLs in 1,111 human lung samples [12]. A predefined hypothesis of the consortium was that improvement of the localization of causal variants/genes in GWAS-nominated COPD loci [13].

Methods

Ethics Statement

All lung tissue samples were obtained in accordance with Institutional Review Board guidelines at the three sites: Laval...
Clinical characteristics of patients that passed gene expression and genotyping quality control filters.

**Table 1.** Clinical characteristics of patients that passed gene expression and genotyping quality control filters.

<table>
<thead>
<tr>
<th></th>
<th>Laval (n = 409)</th>
<th>UBC (n = 339)</th>
<th>Groningen (n = 363)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Male (%)</td>
<td>55.9</td>
<td>53.7</td>
<td>53.2</td>
</tr>
<tr>
<td>Age (years)</td>
<td>63.3±9.9</td>
<td>60.2±14.3</td>
<td>51.5±15.5 [9]</td>
</tr>
<tr>
<td>Body Mass Index (kg/m²)</td>
<td>26.7±5.3</td>
<td>25.6±5.4 [56]</td>
<td>23.2±4.2 [42]</td>
</tr>
<tr>
<td>FEV₁, predicted – pre-BD* (%)</td>
<td>80.5±18.9 [16]</td>
<td>78.2±24.4 [77]</td>
<td>60.5±30.0 [194]</td>
</tr>
<tr>
<td>FVC predicted – pre-BD* (%)</td>
<td>89.8±16.1 [31]</td>
<td>86.9±20.1 [75]</td>
<td>75.0±26.5 [208]</td>
</tr>
<tr>
<td>FEV₁/FVC</td>
<td>0.67±0.10 [32]</td>
<td>0.67±0.13 [77]</td>
<td>0.64±0.19 [189]</td>
</tr>
<tr>
<td>COPD</td>
<td>211 (51.6%) [34]</td>
<td>115 (33.9%) [99]</td>
<td>158 (43.5%) [120]</td>
</tr>
<tr>
<td>Stage 1 : Mild</td>
<td>82 (38.9%)</td>
<td>43 (37.4%)</td>
<td>20 (12.6%)</td>
</tr>
<tr>
<td>Stage 2 : Moderate</td>
<td>117 (55.4%)</td>
<td>60 (52.2%)</td>
<td>38 (23.9%)</td>
</tr>
<tr>
<td>Stage 3 : Severe</td>
<td>11 (5.2%)</td>
<td>2 (1.7%)</td>
<td>21 (13.2%)</td>
</tr>
<tr>
<td>Stage 4 : Very Severe</td>
<td>1 (0.5%)</td>
<td>10 (8.7%)</td>
<td>69 (43.4%)</td>
</tr>
<tr>
<td>Asthma</td>
<td>15 (3.7%)</td>
<td>22 (6.5%)</td>
<td>0 (0.0%)</td>
</tr>
<tr>
<td>Diabetes</td>
<td>41 (10.0%)</td>
<td>13 (3.8%)</td>
<td>27 (7.4%)</td>
</tr>
<tr>
<td>Cardiac diseases</td>
<td>120 (29.3%)</td>
<td>46 (13.6%)</td>
<td>28 (7.7%)</td>
</tr>
<tr>
<td>Smoking</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Smoker</td>
<td>90 (22.0%)</td>
<td>98 (28.9%)</td>
<td>57 (15.7%)</td>
</tr>
<tr>
<td>Ex-Smoker</td>
<td>283 (69.2%)</td>
<td>163 (48.1%)</td>
<td>185 (51.0%)</td>
</tr>
<tr>
<td>Non-Smoker</td>
<td>36 (8.8%)</td>
<td>26 (7.7%)</td>
<td>100 (27.5%)</td>
</tr>
<tr>
<td>Not available</td>
<td>0 (0.0%)</td>
<td>52 (15.3%)</td>
<td>21 (5.8%)</td>
</tr>
<tr>
<td>Pack-years in ever-smokers</td>
<td>48.5±27.5 [37]</td>
<td>44.7±28.5 [58]</td>
<td>31.2±17.4 [51]</td>
</tr>
</tbody>
</table>

FEV₁ : forced expiratory volume in 1 second.
FVC : forced vital capacity.
[-] = missing value.
*pre-BD: pre-bronchodilator.
doi:10.1371/journal.pone.0070220.t001

COPD Susceptibility Loci

Lung eQTLs were overlaid onto COPD susceptibility loci identified by previous GWAS except for the 15q25-CHRNA5/CHRNA3/IREB2 locus that we have reported on previously [15]. Three COPD loci were considered; 4q22 (FAMI3A, 4q31 (HHIP) and 19q13 (RAB4B, EGLN2, MIA, CYP2A6). SNPs associated with COPD from previous GWAS were tabulated for the three loci (Table 2). SNPs genotyped in the lung eQTL consortium located 1 Mb up and downstream of the most distant associated SNPs in both directions were evaluated. Chromosomes 4q22 (88,875,909-90,886,297), 4q31 (144,480,780-146,506,456) and 19q13 (40,292,404-42,302,706) include 718, 412 and 739 SNPs, respectively. Genes residing in the same regions were tested as cis-eQTLs for probe sets for 14 genes on 4q22 (PPPI, PKD2, ABCG2, PPM1K, HERC6, HERC5, PIGT, HERC3, NAPIL3, FAM13A, TIGD2, GPRIN3, SVCA, MMIREN7), 9 genes on 4q31 (FREM3, GYPE, GYPB, GIPA, HHIP, ANAPC10, ABCE1, OTUD4, SMAD1) and 45...
genes on 19q13 (DYM, FBL, FCGP, PSMC4, ZNF546, ZNF780B, ZNF780A, MAP3K10, TTCTB, CNTD2, ART2, C1orf92, PLD3, PRX, SERTAD1, SERTAD2, BLVRB, SFTBN4, SHKBPI, LTBP4, NUMBL, ADCK4, ITPKC, C1orf95, SNRPA, EGLN2, CYP2G1P, CYP2B6, CYP2A13, CYP2S1, AXL, NHRNPU1, TGFBI, B9D2, TMEH, EXOSC3, BCKDHA, B5GNT3, ATP5XL, LOC100505495, CEACAM21, CEACAM4, CEACAM7, CEACAM3, CEACAM6).

Statistical Analysis

Lung eQTLs were identified using a different model than in our previous genome-wide lung eQTL mapping study [12]. Expression data were adjusted for age, sex, and smoking status using robust residuals obtained with the robust fitting of linear models function (rlm) in the R statistical package MASS. Residuals values deviating from the median by more than three standard deviations were filtered as outliers. Association tests between adjusted expression traits and SNPs were performed using quantitative association tests implemented in PLINK [16] (version 1.07). Association tests were performed with the “assoc” command and association tests implemented in PLINK [16] (version 1.07).

Table 2. SNPs associated with COPD in previous GWAS.

<table>
<thead>
<tr>
<th>Locus</th>
<th>SNP</th>
<th>SNP position</th>
<th>Study</th>
</tr>
</thead>
<tbody>
<tr>
<td>4q22</td>
<td>rs1964516</td>
<td>89,875,909</td>
<td>Cho et al. 2012. Human Molecular Genetics.1</td>
</tr>
<tr>
<td></td>
<td>rs7671167</td>
<td>89,883,979</td>
<td>Cho et al. 2010. Nature Genetics.10</td>
</tr>
<tr>
<td>4q31</td>
<td>rs1828591</td>
<td>145,480,780</td>
<td>Cho et al. 2010. Nature Genetics.10</td>
</tr>
<tr>
<td></td>
<td>rs1930300</td>
<td>89,886,297</td>
<td>Cho et al. 2010. Nature Genetics.10</td>
</tr>
<tr>
<td></td>
<td>rs7671167</td>
<td>89,883,979</td>
<td>Cho et al. 2010. Nature Genetics.10</td>
</tr>
<tr>
<td>19q13</td>
<td>rs2604894</td>
<td>41,292,404</td>
<td>Cho et al. 2012. Human Molecular Genetics.11</td>
</tr>
<tr>
<td></td>
<td>rs7937</td>
<td>41,302,706</td>
<td>Cho et al. 2012. Human Molecular Genetics.11</td>
</tr>
</tbody>
</table>

Results

Lung eQTLs in the 4q22 Locus

718 SNPs and 50 probesets covering 14 genes were located in the defined region on chromosome 4q22. 91 eQTLs were detected in the Laval set (Figure 1, Table S1). These 91 eQTLs consisted of 64 unique SNPs, 8 probesets and 4 genes (PPM1K, GPRIN3, SNCA, MMRI). Significant linkage disequilibrium (LD) was observed among the 64 SNPs (Figure S1). 61 of the 91 eQTLs replicated in both replication cohorts (P<0.05). The strongest association detected in all cohorts was rs17013978 with PPM1K (Figure 1). The expression level of this gene decreased with the number of T alleles (Figure 2). In this cohort, this SNP explained 50.2 to 57.1% of the expression variance of PPM1K and the direction of the effect was the same in the three cohorts. None of the SNPs previously associated with COPD on 4q22 (Table 2) were genotyped in our eQTL dataset, but five of them were found in LD (r^2>0.5) (Figure 3). These five SNPs did not significantly associated with the expression of genes at that locus, but a trend was observed with FAM13A. In fact, three of those five polymorphisms, in complete LD with each other (rs2045517, rs2869967, rs2869966) and in modest LD (r^2 = 0.53-0.69) with COPD SNPs were nominally associated with the expression levels of FAM13A (P = 4.1 x 10^-5). The FAM13A-rs2045517 eQTL was replicated in UBC, but not in Groningen (Figure 4).

Lung eQTLs in the 4q31 Locus

412 SNPs and 34 probesets interrogating 9 unique genes were tested around previously COPD associated SNPs on chromosome 4q31. Significant eQTLs found in the Laval dataset are shown in Figure 5 and Table S2. 55 unique SNPs, 6 probesets and 4 genes (FREM3, BC029578, HHIP, OTUD4) were involved in the significant eQTLs. Only eQTLs associated with BC029578 (35) and OTUD4 (1) were replicated in the two replication sets. eQTL-SNPs on chromosome 4q31 are subdivided in two strong LD blocks (Figure S2). The strongest eQTL in Laval dataset, validated in both replication sets, was rs7667092 with BC029578 (Figure 6). The expression levels of the gene increased with the number of T alleles in all cohorts. In the three cohorts, this SNP explained 7.6 to 12.5% of the gene expression variance of BC029578. However, this polymorphism was not in LD with SNPs previously associated with COPD (r^2 = 0.016). Two SNPs (rs1828591, rs13118928) previously associated with COPD were found to affect the expression of HHIP. Rs1828591 was the most significant SNP associated with HHIP in the Laval dataset. This eQTL was replicated in UBC, but not in Groningen (Figure 7). The G allele was associated with lower expression of HHIP in the Laval and UBC datasets. The same pattern was observed in the Groningen set, but the association was not significant.

Lung eQTLs in the 19q13 Locus

On 19q13, 739 SNPs and 95 probesets covering 45 different genes were tested. The expression levels of RAB4B, MIA and CYP2A6 were not available in our datasets. 222 eQTLs were detected (Figure 8 and Table S3). 174 SNPs were regulating 11 genes. The strongest eQTL detected was rs17013978 with PPM1K (Figure 1). This SNP explained 50.2 to 57.1% of the expression variance of PPM1K and the direction of the effect was the same in the three cohorts. None of the SNPs previously associated with COPD on 4q22 (Table 2) were genotyped in our eQTL dataset, but five of them were found in LD (r^2>0.5) (Figure 3). These five SNPs did not significantly associated with the expression of genes at that locus, but a trend was observed with FAM13A. In fact, three of those five polymorphisms, in complete LD with each other (rs2045517, rs2869967, rs2869966) and in modest LD (r^2 = 0.53-0.69) with COPD SNPs were nominally associated with the expression levels of FAM13A (P = 4.1 x 10^-5). The FAM13A-rs2045517 eQTL was replicated in UBC, but not in Groningen (Figure 4).
EGLN2/MIA/CYP2A6 (Figure 8). These eQTL-SNPs were not in LD with the COPD SNPs rs7937 and rs2604894. The latter two SNPs were in strong LD ($r^2 = 0.82$) and rs7937 was genotyped in our lung eQTL dataset. Rs7937 was not associated with

Figure 1. Lung eQTLs on 4q22 in the Laval dataset. Each dot represents an association test between one SNP and one probeset. Only dots above the red line are significant ($p < 5.10^{-5}$). Significant SNPs were regulating the expression levels of PPM1K in red, GPRIN3 in blue, SNCA in green and MMRN1 in purple. The SNP with the smaller p-value is indicated. SNPs previously associated with COPD are presented at the bottom. doi:10.1371/journal.pone.0070220.g001

Figure 2. Boxplots of gene expression levels in the lung for PPM1K according to genotype groups for SNP rs17013978. The left y-axis shows the mRNA expression levels for PPM1K. The x-axis represents the three genotyped groups for SNP rs17013978. The right y-axis shows the proportion of the gene expression variance explained by the SNP (black bar). Each panel represents a different cohort: Laval (n = 392), UBC (n = 287), Groningen (n = 342). The eQTL p-values were $5.6 \times 10^{-51}$, $2.8 \times 10^{-51}$ and $3.8 \times 10^{-55}$, respectively. doi:10.1371/journal.pone.0070220.g002
expression of genes located in this predefined 19q13 locus. The
gene most significantly regulated by rs7937 was
NUMBL (p = 0.0187). Three SNPs were regulating the expression of
EGLN2, a gene previously associated with COPD. The most
significant association with EGLN2 was with rs4803369
(p = 8.9 × 10^{-7}). Rs4803369 is located at 13,274 bp away from
rs7937 and is in modest LD (r^2 = 0.33) with the latter. The eQTL
results for rs4803369-EGLN2 from the three cohorts are illustrated
in Figure 9. This eQTL was significant and had the same direction
of effect in all 3 cohorts.

Discussion

The goal of this study was to identify causal variants and genes
within susceptibility loci associated with COPD. GWAS have
indicated four loci associated with this disease as defined by lung
function measures [9–11]. However, GWAS could not fully
revealed the genetic mechanisms mediating the risk within these
coli. In this study, we used genotypes and expression values of a
large number of lung samples derived from three independent
populations to identify eQTLs. Our analyses were centered on
three loci previously associated with COPD: 4q22 (FAM13A, 4q31
(HHIP) and 19q13 (RAB4B, EGLN2, MIA, CYP2A6). We identified
genetic variants influencing gene expression at each locus and
replicated findings in two independent cohorts.

The first study to identify an association between the 4q22 locus
and COPD was published in 2010 [19]. Three other studies
confirmed an association between this locus and COPD/lung
function [10,11,20]. In this study, we found 91 eQTLs on 4q22 in
the discovery cohort and 61 of them were replicated in both
replication sets. The majority of the SNPs were located in introns
(n = 30) and intergenic regions (n = 27). Other SNPs were located
in the 3’ UTR (n = 4) and upstream of the gene (n = 2). Only one
missense SNP was found to regulate the expression of
MMRN1.

Lung eQTLs on 4q22 were found and validated with four genes:
PMM1K, SNCA, PPMA1 and GPRIN3 genes. A SNP located in
SNCA (rs2035268) has been associated with accelerated FEV1/
FVC decline [21]. Three SNPs in our dataset were in perfect LD
with rs2035268: rs3889917, rs7684637 and rs3773461.

However, none of those SNPs were significantly associated with
the expression level of a gene. The best r^2 between one of our
significant eQTL-SNP and rs2035268 was 0.047. No SNP
previously associated with COPD within and near FAM13A
[10,11,19] were available in our dataset. However, some
polymorphisms were in LD with the disease associated SNPs
(r^2>0.5). The latter were nominally associated with the expression
of FAM13A and validated in one replication set. Accordingly, our
results provide some support that FAM13A is the COPD causal
gene on 4q22.

The 4q31 locus was first associated with COPD and lung
function in two studies in 2009 [9,22], and then replicated in four
other GWAS [10,11,19,23]. In our discovery set, 55 SNPs, 6
probesets covering 4 genes were significant. Interestingly, some of
the eQTL-SNPs have been previously associated with COPD

---

Figure 3. Linkage disequilibrium plot of selected SNPs on the 4q22 locus in the 1000 Genome Project. The white horizontal bar on the
upper part of the figure illustrates the location of SNPs on a physical scale. LD values (r^2) are indicated in each box. The color of the squares illustrates
the strength of pairwise r^2 values on a black and white scale where black indicates perfect LD (r^2 = 1) and white indicates perfect equilibrium (r^2 = 0).
The genotypes are from the 1000 Genome Project interim phase 1 release (2010/11/23). Red rectangles are SNPs previously associated with COPD
(Table 2). Blue rectangles are the most significant eQTL-SNPs for the four regulated genes found on 4q22 (Figure 1). The other illustrated SNPs were
genotyped in our study and in LD (r^2>0.5) with COPD SNPs.
doi:10.1371/journal.pone.0070220.g003
Figure 4. Boxplots of gene expression levels in the lung for FAM13A according to genotype groups for SNP rs2045517. The left y-axis shows the mRNA expression levels for FAM13A. The x-axis represents the three genotype groups for SNP rs2045517. The right y-axis shows the proportion of the gene expression variance explained by the SNP (black bar). Each panel represents a different cohort: Laval (n = 407), UBC (n = 287), Groningen (n = 342). The eQTL p values were 4.1 \times 10^{-5}, 0.009 and 0.218, respectively. doi:10.1371/journal.pone.0070220.g004

Figure 5. Lung eQTLs on 4q31 in the Laval dataset. Each dot represents an association test between one SNP and one probeset. Only dots above the red line are significant (p < 1.5 \times 10^{-5}). Significant SNPs were regulating the expression levels of BC029578 in green, FREM3 in purple, HHIP in blue, and OTUD4 in red. The SNP with the smaller p-value is indicated. SNPs previously associated with COPD are presented at the bottom. doi:10.1371/journal.pone.0070220.g005
SNPs were mainly located in intergenic regions (n = 49). Others were in introns (n = 4), coding-synonymous region (n = 1) and 3’UTR region (n = 1). The only eQTLs replicating in the two replication sets are those associated with the BC029578 transcript and another associated with OTUD4. This transcript is located between the GYPE and GYPB genes. SNPs regulating BC029578 are distributed

(rs1828591 [9,24], rs13118928 [9,24]) and lung function (rs1828591 [22], rs7655625 [22], rs1980057 [19,22]). SNPs were mainly located in intergenic regions (n = 49). Others were in introns (n = 4), coding-synonymous region (n = 1) and 3’UTR region (n = 1). The only eQTLs replicating in the two replication sets are those associated with the BC029578 transcript and another associated with OTUD4. This transcript is located between the GYPE and GYPB genes. SNPs regulating BC029578 are distributed

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(rs1828591 [9,24], rs13118928 [9,24]) and lung function (rs1828591 [22], rs7655625 [22], rs1980057 [19,22]). SNPs were mainly located in intergenic regions (n = 49). Others were in introns (n = 4), coding-synonymous region (n = 1) and 3’UTR region (n = 1). The only eQTLs replicating in the two replication sets are those associated with the BC029578 transcript and another associated with OTUD4. This transcript is located between the GYPE and GYPB genes. SNPs regulating BC029578 are distributed
across a 400 kb region. Further studies are needed to understand the function of BC029578. eQTLs were also associated with FREM3 and HHIP, a member of the hedgehog-interacting protein family. HHIP has been associated with COPD in three GWAS [9–11]. Significant eQTLs in this gene only replicated in UBC, but the same direction of effect was also observed in the Goningen set. These results supported that HHIP is the most likely causal gene in the region.

There are many genes present in the 19q13 locus. This locus was recently associated with COPD and so far no replication study has been published [11]. 222 eQTLs were detected in our original set and 210 of them were validated in the replication sets. Ten genes were regulated by SNPs in the Laval dataset, which were all validated in replication sets. Some SNPs have been previously associated with COPD (rs2302188 [25], rs4803481 [25], rs1800469 [26,27]) and lung function (rs2941718 [26], rs96957 [26]). Interestingly, CEACAM21 was associated with COPD susceptibility in a sputum eQTLs study on COPD patients [25]. This gene encodes carcinoembryonic antigen, which has been found to be overexpressed in heavy smokers [28,29]. To the best of our knowledge, no studies have to date supported the contribution of AXL, NUMBL, SERTAD3, B3GNT8, CEACAM4, CYP2G1P, LOC100505495 or ZNF780A to the development of COPD or related phenotypes. Rs7937, a SNP located in RAB4B, EGLN2 and MIA-RAB4B and identified in previous GWAS, was genotyped in our datasets. However, no association was detected between this SNP and the expression level of any gene. The strongest association with a suspected COPD gene is EGLN2-rs4803369. This gene is known to be involved in regulating hypoxia tolerance and apoptosis in cardiac and skeletal muscle. These results support that EGLN2 is a potential causal COPD gene on 19q13.

eQTLs obtained in this study are derived from non-tumor lung parenchymal samples. As all organs, the lung is multicellular. The cellular heterogeneity constitutes an inherent limitation of our study and will inevitably reduce the power to detect eQTLs. It is known that many eQTLs will be missed by studying heterogeneous tissues [30]. Although many eQTLs are shared across tissues [31,32], a relatively large portion of eQTLs are cell type- and tissue-specific [33,34]. eQTL mapping results are also known to be affected by environmental cues as well as the development stage and differentiation states of cells [35,36]. Due to the spatiotemporal characteristics of eQTLs [30–38], the lung eQTL results in this study will need to be verified in others disease-relevant tissues.
and cell types as well as in tissues from diseased and healthy individuals.

In conclusion, we used a large collection of human lung specimens from patients with and without COPD to identify SNPs that regulated gene expression in three COPD susceptibility loci derived from GWAS. Strong lung eQTLs were detected in the three COPD loci. However, the eQTL-SNPs were not necessarily the SNPs associated with COPD. On 4q22, SNPs associated with COPD near the FAM13A gene were indirectly (in LD) associated with the mRNA expression levels of FAM13A. On 4q31, the suspected candidate in this region, HHIP, was found to be regulated by SNPs previously associated with COPD. On 19q13, SNPs associated with COPD were consistently associated with the expression level of EGLN2. Further functional studies will be needed to verify the contribution of susceptibility genes in COPD.

Strong lung eQTL SNPs will also need to be tested for association with COPD in case-control studies and in additional eQTL mapping studies in other disease-relevant tissues and cell types. This study is an important step forward to better understanding the underlying biology of the COPD susceptibility loci. It also shows the potential of eQTLs in a relevant tissue to leverage the results of previous GWAS and extend their functional meaning to gene expression.

Supporting Information

Figure S1 Linkage disequilibrium plot of significant SNPs on the 4q22 locus in the 1000 Genome Project. The white horizontal bar on the upper part of the figure illustrates the location of SNPs on a physical scale. LD values ($r^2$) are indicated in each box. The color of the squares illustrate the strength of pairwise $r^2$ values on a black and white scale where black indicates perfect LD ($r^2 = 1$) and white indicates perfect equilibrium ($r^2 = 0$). Red rectangles are SNPs previously associated with COPD (Table 2). The genotypes are from the 1000 Genome Project interim phase 1 release (2010/11/23).

Figure S2 Linkage disequilibrium plot of significant SNPs on the 4q31 locus in the 1000 Genome Project. The white horizontal bar on the upper part of the figure illustrates the location of SNPs on a physical scale. LD values ($r^2$) are indicated in each box. The color of the squares illustrate the strength of pairwise $r^2$ values on a black and white scale where black indicates perfect LD ($r^2 = 1$) and white indicates perfect equilibrium ($r^2 = 0$). Red rectangles are SNPs previously associated with COPD (Table 2). The genotypes are from the 1000 Genome Project interim phase 1 release (2010/11/23).

Figure S3 Linkage disequilibrium plot of significant SNPs on the 19q13 locus in the 1000 Genome Project. The white horizontal bar on the upper part of the figure illustrates the location of SNPs on a physical scale. LD values ($r^2$) are indicated in each box. The color of the squares illustrate the strength of pairwise $r^2$ values on a black and white scale where black indicates perfect LD ($r^2 = 1$) and white indicates perfect equilibrium ($r^2 = 0$). Red rectangles are SNPs previously associated with COPD (Table 2). The genotypes are from the 1000 Genome Project interim phase 1 release (2010/11/23).

Table S1 Significant eQTLs at the 4q22 locus in the Laval dataset and replication in UBC and Groningen datasets.

Table S2 Significant eQTLs at the 4q31 locus in the Laval dataset and replication in UBC and Groningen datasets.
Table S3 Significant eQTLs at the 19q13 locus in the Laval dataset and replication in UBC and Groningen datasets.

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Author Contributions

Conceived and designed the experiments: M. Lamontagne YB. Performed the experiments: DN. Analyzed the data: M. Lamontagne. Contributed reagents/materials/analysis tools: M. Laviolette CC PP DS JH DP WT. Wrote the paper: M. Lamontagne YB.

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