

University of Groningen

## Exploring Redox Biology in physiology and disease

Koning, Anne

**IMPORTANT NOTE: You are advised to consult the publisher's version (publisher's PDF) if you wish to cite from it. Please check the document version below.**

*Document Version*

Publisher's PDF, also known as Version of record

*Publication date:*  
2017

[Link to publication in University of Groningen/UMCG research database](#)

*Citation for published version (APA):*

Koning, A. (2017). *Exploring Redox Biology in physiology and disease*. Rijksuniversiteit Groningen.

**Copyright**

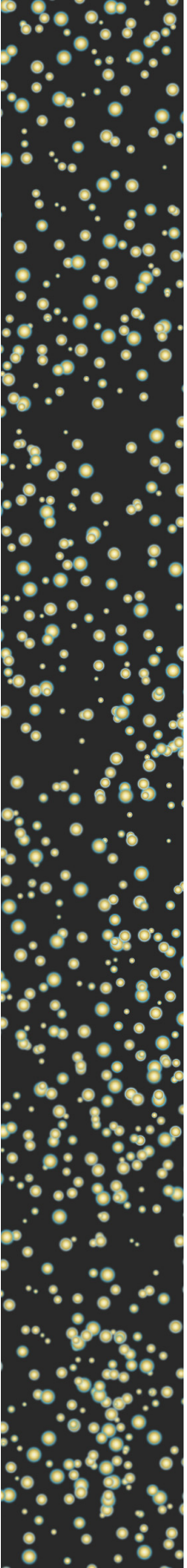
Other than for strictly personal use, it is not permitted to download or to forward/distribute the text or part of it without the consent of the author(s) and/or copyright holder(s), unless the work is under an open content license (like Creative Commons).

The publication may also be distributed here under the terms of Article 25fa of the Dutch Copyright Act, indicated by the "Taverne" license. More information can be found on the University of Groningen website: <https://www.rug.nl/library/open-access/self-archiving-pure/taverne-amendment>.

**Take-down policy**

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

*Downloaded from the University of Groningen/UMCG research database (Pure): <http://www.rug.nl/research/portal>. For technical reasons the number of authors shown on this cover page is limited to 10 maximum.*



# Chapter 1

## Introduction

## Redox biology

Redox reactions - characterized by the transfer of electrons from one chemical species to another - are responsible for the regulation of numerous bodily processes.(1) Under physiological conditions there is a fine-tuned balance between oxidants and reductive capacity. In these circumstances, reactive oxygen, nitrogen and sulfur species (ROS, RNS and RSS) are involved in various physiological cellular processes, including immunity, cell growth and apoptosis.(2–4) Common examples of these species are super oxide ( $O_2^-$ ), hydrogen peroxide ( $H_2O_2$ ), peroxynitrite ( $ONOO^-$ ) and thiyl radical ( $RS\cdot$ ). Unaccompanied by adequate upregulation of antioxidant defence mechanisms, excess production of reactive species leads to oxidative stress, which in turn has been implicated in the development of many disease conditions, including cardiovascular and renal disease.(2,5) The redox biology field has long focussed on the development of oxidative stress biomarkers. Even though these can be useful to determine the degree of oxidative stress and thereby the severity of a certain condition, most of these markers are oxidation products rather than active components of the redox signalling network and therefore provide little mechanistic insight, let alone intervention targets.(6)

## Thiols

Thiols, on the other hand, are critical active components of the antioxidant machinery. These functional groups, consisting of a sulfur and an hydrogen atom, play an important role in transferring signals from distinct elements of the redox network and accordingly are often called redox-switches.(7–11) Since reduced (or free) thiols are readily oxidized by reactive species, their level may be interpreted as a direct reflection of the overall redox status.(11,12) So far, most studies looking into antioxidant machinery have focused on low molecular weight (LMW) thiols, in particular glutathione and cysteine. However, whereas LMW thiols are key actors in intracellular antioxidant defence, in the extracellular compartment these are strongly outnumbered by protein thiols.(7,13) In fact, the majority of thiols in serum (~0.6 mM) is accounted for by the single thiol group of albumin, the most abundant serum protein. Therefore, compared to the concentration of LWM thiols, the total free thiol level may be a more relevant circulatory marker of oxidative stress and, thereby, of disease severity.

## Gasotransmitters

Reversible oxidative posttranslational modifications of protein thiols by several small molecules may protect proteins from irreversible oxidative damage and in certain cases alter protein function.(11,13,14) Among these redox-active small molecules are gasotransmitters nitric oxide (NO) and hydrogen sulfide ( $H_2S$ ), which are able to induce S-nitrosylation and S-sulfhydration, respectively.(11) S-sulfhydration is indirectly brought about by  $H_2S$ , either through reactions of oxidized thiols with sulfide or through reactions of sulfide oxidation products with thiols, giving rise to persulfides.(15–17)

While gasotransmitters - further including carbon monoxide - were initially dismissed as toxic gases causing environmental hazard, over the past decades it became apparent that

these molecules are enzymatically produced and exert important regulatory functions in the mammalian body.(18) NO formation from L-arginine is regulated by endothelial, inducible and neuronal NO synthase (NOS), of which endothelial NOS, expressed by endothelial cells, is primarily responsible for NO production in the cardiovascular system. In turn, all H<sub>2</sub>S producing enzymes, cystathionine β-synthase (CBS), cystathionine γ-lyase and 3-mercaptopyruvate, are abundantly present in the vasculature. Both NO and H<sub>2</sub>S have been acknowledged for their versatile role in (patho)physiological processes and feature vasodilatory, antioxidant and anti-inflammatory properties, to name a few.(18)

In short, by constantly relaying signals the redox network is involved in the majority of bodily processes. Studying its components, such as thiols and gaseous signalling molecules, can provide valuable information to advance our understanding of pathophysiological conditions and may even reveal novel targets for therapy.

## Scope of the thesis

The general aim of this thesis was to explore elements of redox signalling in physiology and disease. To this end, we have assessed concentrations of reduced thiols and gasotransmitter metabolites in blood and urine samples from a variety of human populations and have related these to risk or outcome of cardiovascular and renal disease. Also, variants of the *cystathionine β-synthase (CBS)* gene – a gene involved in the regulation of H<sub>2</sub>S production – were investigated in relation to graft survival in human renal transplantation. Finally, the therapeutic potential of H<sub>2</sub>S was studied in rat models of renal disease.

**Chapter 2** comprises a more extensive introduction to redox biology and presents the concept of redox regulation by the reactive species interactome. The remainder of the thesis is divided into two parts.

**Part 1** of this thesis focuses on the role of thiol biology in heart failure. In **Chapter 3** the link between metabolic changes in heart failure and excessive ROS production is reviewed and free thiols are introduced as a potential biomarker and target for therapy. In **Chapter 4** serum free thiol concentrations were determined in a cohort of stable chronic heart failure patients are associated with outcome of disease.

**Part 2** of this thesis is dedicated to gaseous signalling molecules and their metabolites. **Chapter 5** provides an elaborate discussion of the available literature on the potential roles of H<sub>2</sub>S in renal physiology, disease and transplantation. In **Chapter 6** single nucleotide polymorphisms in the *CBS* gene – a gene that regulates the expression of one of the H<sub>2</sub>S producing enzymes – are studied in relation to renal graft survival in humans. H<sub>2</sub>S treatment has previously been shown to protect against renal ischemia-reperfusion injury (IRI). While in mice protection is ascribed to H<sub>2</sub>S-induced metabolic suppression, in larger animals metabolism is generally found to be unaffected by H<sub>2</sub>S.(19–27) However, this gaseous signalling molecule is also considered to be protective independent of metabolic suppression, e.g. by means of its vasodilatory, anti-inflammatory and anti-oxidant properties.(18) In **Chapter 7** we investigated the therapeutic effects of sodium hydrosulfide (NaHS) and sodium thiosulfate (STS) in a model of unilateral renal IRI in rats and the potential of NaHS to induce hypometabolism in isolated rat kidneys. Although thiosulfate is generally

dismissed as a break down product of H<sub>2</sub>S, there is evidence to support its reconversion into H<sub>2</sub>S *in vivo*.(28–31) As a treatment modality, STS is of particular interest as it can safely be administered to humans. In **Chapter 8** the renoprotective properties of NaHS and STS were studied in a rat model of Angiotensin II-induced hypertensive renal disease.

A logical first step towards translation of H<sub>2</sub>S research would be to study endogenous H<sub>2</sub>S metabolism in various human populations, ranging from healthy subjects to patients with severe disease. However, unfortunately a reliable method for direct detection of H<sub>2</sub>S in biological samples is lacking.(32,33) Alternatively, quantification of its metabolites, thiosulfate and sulfate, may be used to evaluate changes in H<sub>2</sub>S metabolism, although other sources of these metabolites as well as their potential direct (patho)physiological roles should be taken into account when doing so. In **Chapter 9** urinary excretion of these sulfur metabolites is assessed in relation to risk of cardiovascular events and all-cause mortality in a large sample of subjects from the general population. **Chapter 10** explores changes in sulfate clearance in CHF patients compared to healthy individuals, as well as the relationship between sulfate clearance and outcome of disease.

Whereas H<sub>2</sub>S has only recently been recognised as a cardiovascular signalling molecule, NO has been acknowledged for its role in cardiovascular physiology ever since its identification as endothelial derived relaxing factor back in 1987.(34–37) Circulating concentrations of its metabolites, nitrite and nitrate, are often used as markers of endogenous NO production. However, the potential influence of the renal handling of these anions on circulating levels has largely been disregarded. In fact, surprisingly little is known about the handling of either anion by the kidneys. In **Chapter 11** we therefore aimed to elucidate the renal handling of nitrite and nitrate and to explore its relevance to clinical outcome in CHF.

In **Chapter 12** the content of this thesis is summarized and discussed. To conclude, we provide our perspective on the future of redox biology research and the therapeutic potential of redox modulation in aging and disease.

## References

1. Jones DP, Sies H. The Redox Code. *Antioxid Redox Signal*. 2015 Sep 20;23(9):734–46.
2. Valko M, Leibfritz D, Moncol J, Cronin MTD, Mazur M, Telser J. Free radicals and antioxidants in normal physiological functions and human disease. *Int J Biochem Cell Biol*. 2007;39(1):44–84.
3. Goszcz K, Deakin SJ, Duthie GG, Stewart D, Leslie SJ, Megson IL. Antioxidants in Cardiovascular Therapy: Panacea or False Hope? *Front Cardiovasc Med* 2015 Jul 6;2:29.
4. DeLeon ER, Gao Y, Huang E, Arif M, Arora N, Divietro A, et al. A case of mistaken identity: are reactive oxygen species actually reactive sulfide species? *Am J Physiol Regul Integr Comp Physiol*. 2016 Apr 1;310(7):R549–560.
5. Poulianiti KP, Kaltsatou A, Mitrou GI, Jamurtas AZ, Koutedakis Y, Maridaki M, et al. Systemic Redox Imbalance in Chronic Kidney Disease: A Systematic Review. *Oxid Med Cell Longev*. 2016;2016:1–19.
6. Margaritelis NV, Cobley JN, Paschalis V, Veskoukis AS, Theodorou AA, Kyparos A, et al. Going retro: Oxidative stress biomarkers in modern redox biology. *Free Radic Biol Med*. 2016 Sep;98:2–12.
7. Turell L, Radi R, Alvarez B. The thiol pool in human plasma: The central contribution of albumin to redox processes. *Free Radic Biol Med*. 2013 Dec;65:244–53.
8. Avery SV. Molecular targets of oxidative stress. *Biochem J*. 2011 Mar 1;434(2):201–10.
9. Imlay JA. Pathways of oxidative damage. *Annu Rev Microbiol*. 2003;57:395–418.
10. Cooper CE, Patel RP, Brookes PS, Darley-Usmar VM. Nanotransducers in cellular redox signaling: modification of thiols by reactive oxygen and nitrogen species. *Trends Biochem Sci*. 2002 Oct;27(10):489–92.
11. Chung HS, Wang S-B, Venkatraman V, Murray CI, Van Eyk JE. Cysteine Oxidative Posttranslational Modifications: Emerging Regulation in the Cardiovascular System. *Circ Res*. 2013 Jan 18;112(2):382–92.
12. Banne AF, Amiri A, Pero RW. Reduced level of serum thiols in patients with a diagnosis of active disease. *J Anti-Aging Med*. 2003;6(4):327–34.
13. Mansoor MA, Svardal AM, Ueland PM. Determination of the in vivo redox status of cysteine, cysteinylglycine, homocysteine, and glutathione in human plasma. *Anal Biochem*. 1992 Feb 1;200(2):218–29.
14. Hochgrafe F, Mostertz J, Pother D-C, Becher D, Helmann JD, Hecker M. S-Cysteinylolation Is a General Mechanism for Thiol Protection of *Bacillus subtilis* Proteins after Oxidative Stress. *J Biol Chem*. 2007 Jul 5;282(36):25981–5.
15. Nagy P. Mechanistic Chemical Perspective of Hydrogen Sulfide Signaling. *Methods Enzymol*. 2015;554:3–29.
16. Ida T, Sawa T, Ihara H, Tsuchiya Y, Watanabe Y, Kumagai Y, et al. Reactive cysteine persulfides and S-polysulfination regulate oxidative stress and redox signaling. *Proc Natl Acad Sci*. 2014 May 27;111(21):7606–11.
17. Greiner R, Pálkink Z, Bäsell K, Becher D, Antelmann H, Nagy P, et al. Polysulfides link H<sub>2</sub>S to protein thiol oxidation. *Antioxid Redox Signal*. 2013 Nov 20;19(15):1749–65.
18. Szabo C. Medicinal Chemistry and Therapeutic Applications of the Gasotransmitters NO, CO, and H<sub>2</sub>S and their Prodrugs. In: *Burger's Medicinal Chemistry and Drug Discovery*. Seventh Edition. Hoboken, NJ, USA: John Wiley & Sons, Inc.; 2010. p. 265–368.
19. Blackstone E, Morrison M, Roth MB. H<sub>2</sub>S Induces a Suspended Animation–Like State in Mice. *Science*. 2005 Apr 22;308(5721):518–518.
20. Bos EM, Leuvenink HGD, Snijder PM, Kloosterhuis NJ, Hillebrands J-L, Leemans JC, et al. Hydrogen Sulfide-Induced Hypometabolism Prevents Renal Ischemia/Reperfusion Injury. *J Am Soc Nephrol*. 2009 Jul 23;20(9):1901–5.
21. Haouzi P, Notet V, Chenuel B, Chalou B, Sponne I, Ogier V, et al. H<sub>2</sub>S induced hypometabolism in mice is missing in sedated sheep. *Respir Physiol Neurobiol*. 2008 Jan 1;160(1):109–15.
22. Haouzi P, Bell HJ, Notet V, Bihain B. Comparison of the metabolic and ventilatory response to hypoxia and H<sub>2</sub>S in unsedated mice and rats. *Respir Physiol Neurobiol*. 2009 Jul 31;167(3):316–22.
23. Aslami H, Heinen A, Roelofs JJTH, Zuurbier CJ, Schultz MJ, Juffermans NP. Suspended animation inducer hydrogen sulfide is protective in an in vivo model of ventilator-induced lung injury. *Intensive Care Med*. 2010 Nov;36(11):1946–52.

24. Seitz DH, Fröba JS, Niesler U, Palmer A, Veltkamp HA, Braumüller ST, et al. Inhaled hydrogen sulfide attenuates endotoxemia-induced organ injury via stimulation of anti-inflammatory pathways, but does not alter the inflammatory response after blunt chest trauma. *Shock* Augusta Ga. 2012 Feb;37(2):197–204.
25. Aslami H, Beurskens CJP, de Beer FM, Kuipers MT, Roelofs JJTH, Hegeman MA, et al. A short course of infusion of a hydrogen sulfide-donor attenuates endotoxemia-induced organ injury via stimulation of anti-inflammatory pathways, with no additional protection from prolonged infusion. *Cytokine*. 2013 Feb;61(2):614–21.
26. Li J, Zhang G, Cai S, Redington AN. Effect of inhaled hydrogen sulfide on metabolic responses in anesthetized, paralyzed, and mechanically ventilated piglets. *Pediatr Crit Care Med J Soc Crit Care Med World Fed Pediatr Intensive Crit Care Soc*. 2008 Jan;9(1):110–2.
27. Drabek T, Kochanek PM, Stezoski J, Wu X, Bayir H, Morhard RC, et al. Intravenous hydrogen sulfide does not induce hypothermia or improve survival from hemorrhagic shock in pigs. *Shock* Augusta Ga. 2011 Jan;35(1):67–73.
28. Mikami Y, Shibuya N, Kimura Y, Nagahara N, Ogasawara Y, Kimura H. Thioredoxin and dihydrolipoic acid are required for 3-mercaptopyruvate sulfurtransferase to produce hydrogen sulfide. *Biochem J*. 2011 Nov 1;439(3):479–85.
29. Villarejo M, Westley J. Mechanism of Rhodanese catalysis of thiosulfate-lipoate oxidation-reduction. *J Biol Chem*. 1963 Dec;238:4016–20.
30. Koj A, Frendo J, Janik Z. [35S]thiosulphate oxidation by rat liver mitochondria in the presence of glutathione. *Biochem J*. 1967 Jun;103(3):791–5.
31. Olson KR, DeLeon ER, Gao Y, Hurley K, Sadauskas V, Batz C, et al. Thiosulfate: a readily accessible source of hydrogen sulfide in oxygen sensing. *Am J Physiol Regul Integr Comp Physiol*. 2013 Sep 15;305(6):R592–603.
32. Nagy P, Pálkás Z, Nagy A, Budai B, Tóth I, Vasas A. Chemical aspects of hydrogen sulfide measurements in physiological samples. *Biochim Biophys Acta*. 2014 Feb;1840(2):876–91.
33. Olson KR, DeLeon ER, Liu F. Controversies and conundrums in hydrogen sulfide biology. *Nitric Oxide Biol Chem Off J Nitric Oxide Soc*. 2014 Sep 15;41C:11–26.
34. Loscalzo J. The identification of nitric oxide as endothelium-derived relaxing factor. *Circ Res*. 2013 Jul 5;113(2):100–3.
35. Furchgott RF, Zawadzki JV. The obligatory role of endothelial cells in the relaxation of arterial smooth muscle by acetylcholine. *Nature*. 1980 Nov 27;288(5789):373–6.
36. Ignarro LJ, Buga GM, Wood KS, Byrns RE, Chaudhuri G. Endothelium-derived relaxing factor produced from artery and vein is nitric oxide. *Proc Natl Acad Sci U S A*. 1987 Dec;84(24):9265–9.
37. Massion PB, Feron O, Dessy C, Balligand J-L. Nitric oxide and cardiac function: ten years after, ... continuing. *Circ Res*. 2003 Sep 5;93(5):388–98.





