

University of Groningen

Total Body Metabolic Tumor Response in ALK Positive Non-Small Cell Lung Cancer Patients Treated with ALK Inhibition

Kerner, Gerald S. M. A.; Koole, Michel J. B.; Bongaerts, Alphons H. H.; Pruijm, Jan; Groen, Harry J. M.; CTMM Air Force Consortium

Published in:
 PLoS ONE

DOI:
[10.1371/journal.pone.0149955](https://doi.org/10.1371/journal.pone.0149955)

IMPORTANT NOTE: You are advised to consult the publisher's version (publisher's PDF) if you wish to cite from it. Please check the document version below.

Document Version
 Publisher's PDF, also known as Version of record

Publication date:
 2016

[Link to publication in University of Groningen/UMCG research database](#)

Citation for published version (APA):

Kerner, G. S. M. A., Koole, M. J. B., Bongaerts, A. H. H., Pruijm, J., Groen, H. J. M., & CTMM Air Force Consortium (2016). Total Body Metabolic Tumor Response in ALK Positive Non-Small Cell Lung Cancer Patients Treated with ALK Inhibition. *PLoS ONE*, 11(5), [e0149955].
<https://doi.org/10.1371/journal.pone.0149955>

Copyright

Other than for strictly personal use, it is not permitted to download or to forward/distribute the text or part of it without the consent of the author(s) and/or copyright holder(s), unless the work is under an open content license (like Creative Commons).

The publication may also be distributed here under the terms of Article 25fa of the Dutch Copyright Act, indicated by the "Taverne" license. More information can be found on the University of Groningen website: <https://www.rug.nl/library/open-access/self-archiving-pure/taverne-amendment>.

Take-down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

Downloaded from the University of Groningen/UMCG research database (Pure): <http://www.rug.nl/research/portal>. For technical reasons the number of authors shown on this cover page is limited to 10 maximum.

RESEARCH ARTICLE

Total Body Metabolic Tumor Response in ALK Positive Non-Small Cell Lung Cancer Patients Treated with ALK Inhibition

Gerald S. M. A. Kerner^{1*}, Michel J. B. Koole², Alphons H. H. Bongaerts², Jan Pruim^{2,3}, Harry J. M. Groen¹, CTMM Air Force Consortium[¶]

1 University of Groningen and Department of Pulmonary Diseases, University Medical Center Groningen, Groningen, the Netherlands, **2** University of Groningen and Department of Nuclear Medicine and Molecular Imaging, University Medical Center Groningen, Groningen, the Netherlands, **3** Department of Nuclear Medicine, Tygerberg Hospital, Stellenbosch University, Stellenbosch, South-Africa

¶ Membership of the CTMM Air Force Consortium is provided in the Acknowledgments.

* g.s.m.a.kerner@umcg.nl



OPEN ACCESS

Citation: Kerner GSMA, Koole MJB, Bongaerts AHH, Pruim J, Groen HJM, CTMM Air Force Consortium (2016) Total Body Metabolic Tumor Response in ALK Positive Non-Small Cell Lung Cancer Patients Treated with ALK Inhibition. PLoS ONE 11(5): e0149955. doi:10.1371/journal.pone.0149955

Editor: Renato Franco, Istituto dei tumori Fondazione Pascale, ITALY

Received: October 19, 2015

Accepted: January 15, 2016

Published: May 3, 2016

Copyright: © 2016 Kerner et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: All relevant data are available via Figshare (<https://dx.doi.org/10.6084/m9.figshare.3189766.v1>).

Funding: This research was performed within the framework of CTMM, the Center for Translational Molecular Medicine, project AIRFORCE (grant 030-103) (<http://www.ctmm.nl>). CTMM paid for GK's salary and had no role in study design, data collection and analysis, decision to publish or preparation of the manuscript. No other external funding sources for this study.

Abstract

Background

In ALK-positive advanced NSCLC, crizotinib has a high response rate and effectively increases quality of life and survival. CT measurement of the tumor may insufficiently reflect the actual tumor load changes during targeted therapy with crizotinib. We explored whether ¹⁸F-FDG PET measured metabolic changes are different from CT based changes and studied the impact of these changes on disease progression.

Methods

¹⁸F-FDG PET/CT was performed prior to and after 6 weeks of crizotinib treatment. Tumor response on CT was classified with RECIST 1.1, while ¹⁸F-FDG PET response was assessed according to the 1999 EORTC recommendations and PERCIST criteria. Agreement was assessed using McNemars test. During follow-up, patients received additional PET/CT during crizotinib treatment and second generation ALK inhibition. We assessed whether PET was able to detect progression earlier than CT.

Results

In this exploratory study 15 patients were analyzed who were treated with crizotinib. There was a good agreement in the applicability of CT and ¹⁸F-FDG PET/CT using the EORTC recommendations. During first line crizotinib and subsequent second line ALK inhibitors, PET was able to detect progression earlier than CT in 10/22 (45%) events of progression and in the others disease progression was detected simultaneously.

Competing Interests: The CTMM Air Force Consortium is a private/public consortium with involvement of academia, private companies, and the government. It is not a commercial source of funding. Gerald Kerner was funded by CTMM consortium to perform the research project (translational and imaging research in lung cancer) that is a part of his thesis. The authors are entitled to publish all his work and share all their data publicly. No consultancy, patents, or products in development are involved. All together, this has no impact to the authors' adherence to all the PLOS ONE policies on sharing data and materials. All authors declared not having any competing interests.

Conclusion

In advanced *ALK* positive NSCLC PET was able to detect progressive disease earlier than with CT in nearly half of the assessments while both imaging tests performed similar in the others.

Introduction

In clinical practice, tumor response measurements are performed using the anatomical CT based RECIST criteria[1]. Nowadays, it has been recognized that metabolic tumor changes as measured with ^{18}F -FDG PET can also be used as an indicator of effectiveness (EORTC recommendations[2], PERCIST[3]). Examples of this principle include imatinib treated gastrointestinal stromal tumors and EGFR-TKI treatment for EGFR mutated advanced NSCLC [4–7]. During targeted therapy, early metabolic changes in tumor activity often precedes anatomic tumor lesion size alterations. *ALK* positive advanced NSCLC are treated with different *ALK* inhibitors such as crizotinib, ceritinib and alectinib[8–11]. Whether targeted treatment such as crizotinib induces quick metabolic changes, and whether these metabolic changes are related to lesions size alterations is currently unknown.

The goal of this paper is to describe metabolic responses on crizotinib in *ALK* positive NSCLC patients and compare PET and CT assessments with different tumor response criteria. Furthermore, we also assessed during follow up with *ALK* inhibition whether PET is able to detect progression of disease at an earlier time point compared to regular CT.

Material and Methods

Patients

Patients with advanced *EML4/ALK* positive NSCLC treated with crizotinib were studied with ^{18}F -FDG PET/CT at baseline and after six weeks of treatment. During and after treatment patients underwent PET/CT until progression of disease was determined. If they were eligible for additional treatment (local treatment or systemic treatment with a second generation *ALK* inhibitor), PET/CT was repeated to assess tumor response again until disease progression was determined.

Informed Consent and Ethics

This study was performed using clinical data from previous studies for which informed consent was obtained. For this study all data were anonymized and de-identified prior to analysis. Under the Dutch Law Medical Research Involving Human Subjects Act (WMO), no (additional) informed consent was necessary from the Institutional Review board.

Pathology

Tumor samples were obtained either by bronchoscopy, transthoracic lung biopsies, or from resections. Samples were examined according to the 2011 IASLC/ATS/ERS NSCLC classification[12]. *ALK* status was determined by FISH and/or by immunohistochemistry.

For detecting the *ALK* fusion gene, the Vysis *ALK* Break Apart FISH probe (Abbott 06N43-020) was used. A score of at least 50 tumor cell in an area on the paraffine coupe was marked

by the pathologist and scored by two different observers. For scoring FISH patterns, the criteria were used as described by Thunnissen et al[13].

To detect ALK expression using immunohistochemistry, a fully automated immunohistochemical assay was used on the Ventana BenchMark Ultra with the anti-ALK (D5F3) rabbit monoclonal primary antibody (Ventana Cat. No. 790–4794 / 06679072001). This analysis was performed using the OptiView DAB IHC Detection Kit and the OptiView Amplification Kit. For assessment the Ventana ALK scoring interpretation guide was used (<http://www.uclad.com/newsletters/ALK-LUNG-IHC-INTERPRETATION-GUIDE.pdf>).

CT

The diagnostic CT images were made on a Siemens Biograph/Somatom mCT scanner (Siemens Healthcare, Erlangen, Germany). The CT was performed in 8 seconds, (effective mAs 80, 120 kV with care dose setting active) in a craniocaudal direction at full inspiration. Slice thickness was 0.5 mm, pitch was 14 with a rotation of 0.5 seconds. Patients were injected with 55 ml of Iomeron contrast 350 mg/ml (Bracco Imaging Deutschland GmbH, Konstanz, Germany) at a speed of 2.5 ml/sec 30 seconds prior to scanning.

Tumor response was measured on CT according to RECIST 1.1 criteria by an experienced radiologist[1].

¹⁸F-FDG PET/CT

¹⁸F-FDG PET/CT images were made on the same Siemens Biograph/Somatom mCT time-of-flight scanner according to EANM guidelines.[14, 15] The voxel size of the EANM reconstructions are 4 by 4 by 2.4 mm (38.4 mm³). Prior to tracer injection, a blood sample was drawn to confirm fasting blood glucose level (<11 mmol/l) after a 6 hour fasting period. Patients were dosed at 3 MBq/kg bodyweight intravenously. Sixty minutes after injection, patients were scanned from the upper leg to the brain. Scan times per bed position were dependent on patient weight, 1 minute if less than 60 kg, 2 minutes if between 60–90 kg and 3 minutes if above 90 kg per bed position[16].

¹⁸F-FDG PET/ response measurement

All PET based analyses were performed using the IMALYTICS research work station (Philips Technologie GmbH Innovative Technologies Aachen, Aachen, Germany). Using the maximum intensity projection (MIP), each separate metastasis was visually selected and an adaptive threshold algorithm was used to calculate the volume of interest. The threshold was set to 41% based upon the study by Cheebsumon et al[17]. This was performed with the following settings: 20 mm distance of the background shell from the 70% peak/contour and 2.5 threshold for voxels to be excluded.

1. Two different methods of metabolic response measurement were used. Using the previously defined VOI, five lesions with the highest SUV_{max} were selected, and the SUV_{max} averaged. On the response scan, the same 5 lesions were selected and averaged again. The difference in percentage between these two measurements was used for response. The assessment was performed according to the 1999 EORTC recommendations[2].
2. Tumor response assessment according to the PERCIST criteria[3] was performed separately by using MIM version 6.0.2 (MIM software, Cleveland, OH, USA) to assess the SUV_{peak}.

Follow up

Follow-up was performed in all patients. Patients were assessed at regular times every 6–12 weeks. After disease progression patients received a new treatment and a subsequent progression event was recorded.

Statistics

All SUV, except for SUV_{peak} in accordance with the PERCIST criteria [3], were corrected for glucose level. The measure of agreement in the applicability between CT response with the EORTC recommendations and PERCIST criteria, respectively, was assessed using the McNemar test. Progression-free survival (PFS) was defined from date of diagnosis until date of tumor progression on CT or death. If a solitary new lesion was detected and was completely treated with a local treatment such as stereotactic radiotherapy, surgery or radiofrequency ablation and no regrowth was determined for at least 3 months, this single event was not considered as progressive disease.

All statistics were performed using SPSS 22.0 (International Business Machines Corp, Armonk, NY, USA).

Results

Fifteen patients were treated with crizotinib as first line ALK inhibition and were followed with ^{18}F -FDG PET/CT, thirteen had baseline imaging, all had follow up imaging. Median duration of follow up was 11 (2–39) months. Median age of the patients was 57 (21–68) year with 12 females and 3 males. Patient characteristics are given in Table 1. Histology results and ALK status either by FISH and/or immunohistochemistry is given in Table 2.

Table 1. Baseline patient characteristics.

	N = 15*
Median age (range)	57 (21–68)
Male/female	3/12
Line of treatment	
First	3
Second	2
Third	4
Fourth	4
Number of patients with metastases in different organs detected by PET/CT before start of crizotinib treatment (N = 13)	
Pulmonary	12**
Mediastinal	13
Hepatic	8
Bone	12
Brain	3
Median PFS during crizotinib treatment in months (N = 15)	6.93 (0.9–26.1)

*2 patients had no FDG-PET/CT baseline imaging study.

** in 1 patient with no pulmonary metastases, the pulmonary metastases became visible within 2 months of therapy.

doi:10.1371/journal.pone.0149955.t001

Table 2. Baseline ¹⁸F-FDG PET and CT tumor response measurements with PERCIST and EORTC criteria and progression-free survival per patient with ALK positive NSCLC.

Patient	Histology	FISH	IHC	CT	FDG PET		PFS
					RECIST	PERCIST	
1	Adeno	+	+	PR	PMR -38, 8	PMR	>26.1
2	Adeno	-	+	PR	PMR -42, 6	PMR	4.9
3	Adeno	+	+	PR	PMR -57, 9	PMR	8.3
4	Adeno	+	+	PR	PMR -77, 6	PMR	15.7
5	Adeno	+	nd	PR	PMR -66, 9	PMR	7.8
6	Adeno	+	+	PR	PMR -70, 6	PMR	10.4
7	Adeno	+	+	PR	PMR -76, 6	PMR	9.2
8	Adeno	+	+	PR	PMR -42, 6	PMR	6.9
9	Adeno	+	-	PD	SMD -1, 8	SMD	1.8
10	NSCLC NOS	+	-	PD	PMD 7, 4	PMD	0.9
11	Adeno	+	+	PR	nd	PMR	1.8
12	Adeno	+	+	SD	nd	PMR	6.3
13	Adeno	+	-	PD	nd	PMD	2

Histology is according to 2011 IASLC/ATS/ERS NSCLC classification[12].

Immunohistochemistry (IHC) was performed according to the Ventana ALK scoring

CT response is defined according to RECIST 1.1 criteria.

FDG response is defined according to PERCIST criteria and 1999 EORTC recommendations.

Response in PERCIST criteria is response category with percentage change, weeks after start of therapy.

PR = partial response

SD = stable disease

PD = progressive disease

PMR = partial metabolic response

SMD = stable metabolic disease

PMD = progressive metabolic disease.

PFS = progression-free survival in months.

nd = not determined

doi:10.1371/journal.pone.0149955.t002

Baseline and 6 weeks CT and ¹⁸F-FDG PET/CT measured responses

In 13 patients PET/CT was performed during crizotinib treatment. With CT according to RECIST criteria, there were 9 patients with a partial response, 1 with stable disease and 3 patients had progressive disease after 6 weeks of therapy. Median PFS was 6.9 (range 0.9–26.1) months.

With PET measurements according to 1999 EORTC recommendations, there were 10 partial metabolic responders, 1 stable metabolic disease and 2 progressive metabolic disease. Using the PERCIST criteria in 10 patients, 8 had a partial metabolic response, 1 stable metabolic disease and 1 progressive metabolic disease (Table 2). There were 2 discordant responses between PET and CT, with a more favorable response on PET (i.e. PMR with SD, or SMD with PD). The per patient change in percentage of SUV_{max} was more pronounced than measured with SUV_{peak} (Fig 1). Although the average outcome using the EORTC recommendations or PERCIST criteria were not impressive, all 10 patients had a clinically dramatic response on the 6 week PET/CT with visual assessment (Fig 2).

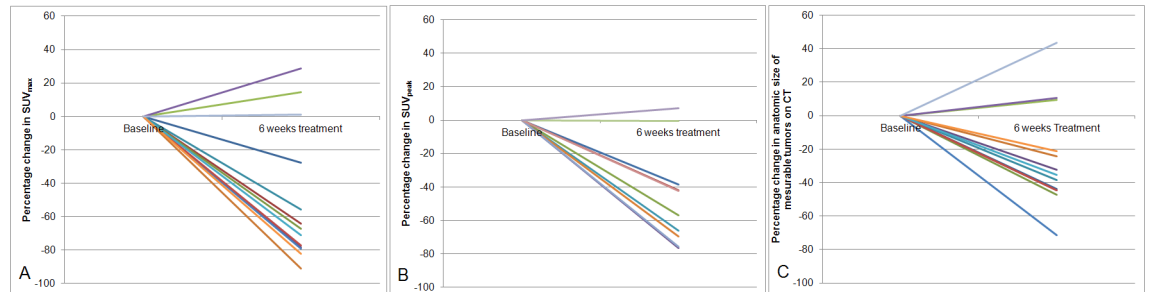


Fig 1. Change in percentage between baseline and after 6 weeks of treatment with crizotinib assessed using SUV_{max} (1A, N = 13), SUV_{peak} (1B, N = 10) and RECIST (1C, N = 13).

doi:10.1371/journal.pone.0149955.g001

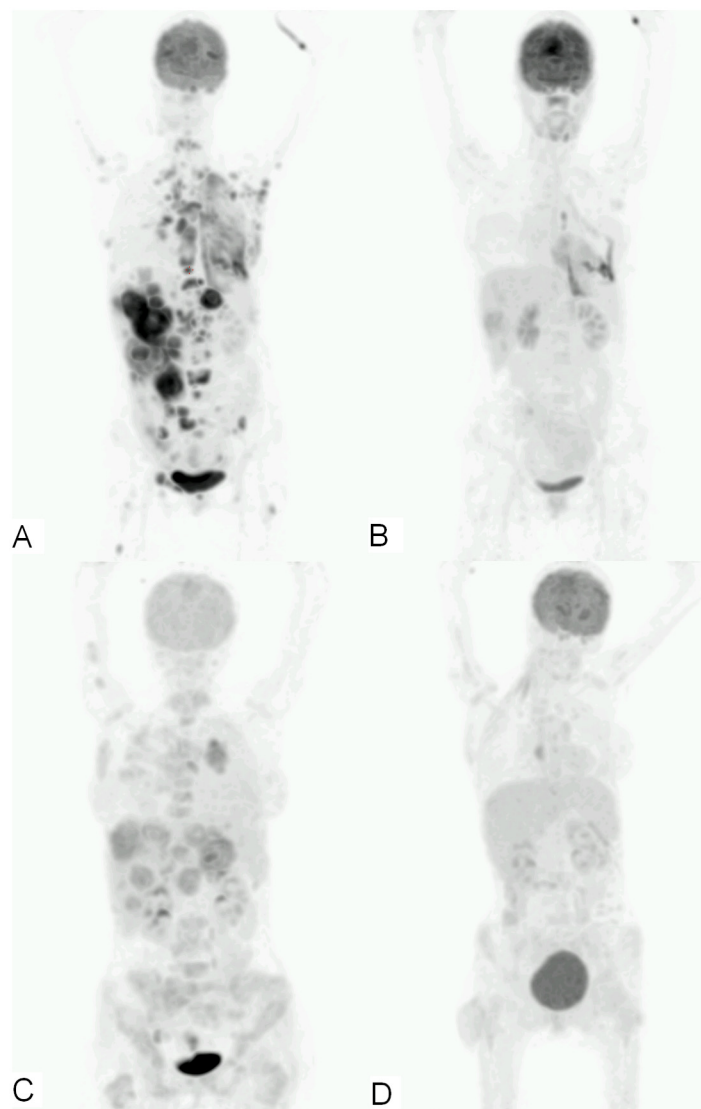


Fig 2. ¹⁸F-FDG maximum intensity projection of patient 2 and 8 prior to (A, B) and after 6 weeks of treatment with crizotinib (C, D). Scale is from 0–15 SUV. These images illustrate the clinically dramatic decrease in ¹⁸F-FDG uptake, with both patients having a PMR according to both PERCIST criteria and the EORTC recommendations.

doi:10.1371/journal.pone.0149955.g002

Overall, there was a good agreement in the applicability between CT and FDG-PET/CT assessed with EORTC recommendations ($N = 13$, $P = 0.37$) at 6 weeks.

Follow-up with CT and ^{18}F -FDG PET/CT

Fifteen patients were included for follow-up, in which additional PET/CT scans were performed. A total of 78 PET/CT were available for evaluation. In 8 out of 15 patients, local progression was detected. Local oligometastatic progression was treated with radio frequency ablation (RFA) in 1 patient, with surgery in 3 patients, and with (stereotactic) radiation in 5 patients. In 6 patients with systemic progression, 4 were treated with ceritinib, 1 with alectinib, and 1 was treated with pemetrexed before receiving ceritinib treatment.

PET/CT was used to detect an increase in metabolic activity at places with previous solid tumors on CT or new lesions that were very small or not yet visible on CT. Comparison of PET and CT according to EORTC criteria at first, second, third line of *ALK* targeted treatment (either systemic or localized treatment) revealed in 5/12, 3/7, 2/3 patients that progressive disease was detected earlier on PET compared with CT. Under first and fourth line treatment one and two patients, respectively, showed no disease progression. This means that in 10/22 (45%) events of progressive disease PET was superior compared to CT. Compared with all assessments, in 10 out of 78 PET/CT, PET alone provided evidence of progression, whereas in 12 out of 78, PET/CT and CT both provided evidence of progression at the same time point.

Discussion

In this exploratory study the metabolic activity in the primary tumor and metastases decreased dramatically soon after starting crizotinib. There was a good agreement in the applicability between CT and PET based response assessment at 6 weeks. However the metabolic activity decreased to a larger extent than the corresponding tumor size on CT. This result was in line with the good agreement between the measurements according to EORTC recommendations and those measured with PERCIST criteria. The SUV_{max} changes showed the largest absolute decrease in activity. To the best of our knowledge, this is also the first study to compare ^{18}F -FDG PET/CT related outcome with *ALK* immunohistochemistry.

Previously, a study with a murine *ALK* positive NSCLC model in which the *ALK* kinase inhibitor TAE684 was administered, a substantially diminished tumor metabolic activity was detected within 24 hours of starting therapy[18]. One clinical study showed that *ALK* positive NSCLC patients had a higher SUV_{max} than *ALK* negative NSCLC patients, but this difference disappeared in larger tumors[19].

Crizotinib treatment is clearly superior to chemotherapy in treating *ALK* positive NSCLC patients, with a PFS of 7.7 months[9] yet unfortunately, treatment with targeted therapy commonly leads to acquired resistance. To overcome crizotinib resistance, different therapeutic strategies have been developed [20]. Identifying resistance to treatment at an early moment in individual patients is important, because in solitary or oligometastases localized treatment options such as stereotactic radiotherapy, video-assisted resections or radiofrequency ablation can be applied. Response assessment with ^{18}F -FDG PET/CT could represent a method with the ability to identify early resistance to treatment and to identify patients with solitary, oligo or “systemic” metastases. Future research should focus on whether such strategy will improve survival, quality of life and cost-effectiveness. What time point is the best to evaluate an early tumor response? We performed the assessments at 6 weeks but that time point may be too late. At that time there was no difference in test performance between PET and CT. In other targeted treatment modalities with advanced NSCLC, early responses on PET preceded anatomic tumor size alterations[4, 6]. A recent study with surgical resections showed that response

monitoring with ^{18}F -FDG PET within 1 week of starting treatment with erlotinib in an unselected NSCLC population identified 64% of histopathological responders[21]. The same study also showed that a decrease in ^{18}F -FDG activity seen after 1 week of therapy is likely to continue after 3 weeks.

Assessing tumor responses at follow up was easier with PET/CT than with CT. In 10/22 events of disease progression in 15 patients, PET was capable of detecting progression earlier than CT. An additional advantage of PET is that progression is detectable outside of the field of view of a CT. These advantages should be taken into account in cost-effectiveness studies using ^{18}F -FDG PET/CT in response assessment during follow-up of oligometastasis.

One problem we encountered, is the discordance between the dramatic results on visual clinical assessment and the less dramatic results using SUV_{max} and SUV_{peak} . The weakness of the traditional PET based measurement assessment are based upon the lesion with single highest uptake value, or as we did, with 5 lesions with the highest SUV_{max} . It does not take into account the sometimes dramatic decrease of all lesions. Furthermore, it does not take into account lesions that become metabolically inactive. Both response assessment techniques determine progression, with either the appearance of a new lesion or the increased uptake of one lesion to at least a certain percentage compared to previous PET scans. Importantly, the comparison in increased uptake is between the two highest measurable lesions, which does not necessarily need to be the same lesion. An example of the discordance can be described in this example: a patient has 3 lesions. After 6 weeks of treatment the main tumor has a SUV_{max} of 5, the liver lesion a SUV_{max} of 2 and a bone lesion with a SUV_{max} of 3. At the next response scan after 12 weeks of treatment, in the main tumor SUV_{max} decreased to 4, the liver lesion SUV_{max} increased to 6 and the bone lesion remains 3. Because the highest SUV_{max} of the lesions is originally 5 and at the last assessment 6, according to the EORTC recommendations and PERCIST criteria, the patient is not progressive, yet the liver lesion has a clear threefold increase in uptake and clinically the patient has progressive disease. Such a patient is eligible for other forms of targeted therapy and/or a local treatment such as surgery or RFA. With new targeted therapy such as crizotinib, the need to identify examples as the above from patients with systemic disease will become more necessary. It is therefore imperative to reconsider our response criteria as is done for immunotherapy.

Conclusion

This explorative study of ^{18}F -FDG PET/CT in *ALK* positive NSCLC patients treated with crizotinib showed a good agreement between CT and PET measurements at 6 weeks. However, follow up with PET increases early detection of metastases. In 45% of detection of progressive disease events in 15 patients treated with *ALK* inhibitors, PET detected progression of disease earlier than CT did.

Acknowledgments

This research was performed within the framework of CTMM, the Center for Translational Molecular Medicine, project AIRFORCE (grant 030–103), project leader prof.dr. G. van Dongen (email g.van.dongen@vumc.nl).

Members: E. Caldenhoven (eric.caldenhoven@ctmm.nl); V.M.H. Coupé; (georgy.shakirin@philips.com); A. Fischer (alexander.fischer@philips.com); H.J.M. Groen (h.j.m.groen@umcg.nl); L. Perk (l.perk@cyclotron.nl); M. van Herk (portal@nki.nl); P.H. Elsinga (p.h.elsinga@umcg.nl); P. Lambin (philippe.lambin@maastro.nl); R.H. Brakenhoff (r.brakenhoff@vumc.nl); R. Boellaard (r.boellaard@vumc.nl); C. Uyl (uyl@bmg.eur.nl); W. van Criekinge (wim.vancrieking@ugent.be)

We are grateful for the help provided by J.H. van Snick with the logistics and execution of this study.

Author Contributions

Conceived and designed the experiments: HG. Performed the experiments: GK. Analyzed the data: GK MK AB JP HG. Contributed reagents/materials/analysis tools: MK AB. Wrote the paper: GK MK AB JP HG.

References

1. Eisenhauer EA, Therasse P, Bogaerts J, Schwartz LH, Sargent D, Ford R, et al. New response evaluation criteria in solid tumours: revised RECIST guideline (version 1.1). *Eur J Cancer*. 2009; 45(2):228–47. Epub 2008/12/23. doi: S0959-8049(08)00873-3 [pii] doi: [10.1016/j.ejca.2008.10.026](https://doi.org/10.1016/j.ejca.2008.10.026) PMID: [19097774](https://pubmed.ncbi.nlm.nih.gov/19097774/).
2. Young H, Baum R, Cremerius U, Herholz K, Hoekstra O, Lammertsma AA, et al. Measurement of clinical and subclinical tumour response using [18F]-fluorodeoxyglucose and positron emission tomography: review and 1999 EORTC recommendations. European Organization for Research and Treatment of Cancer (EORTC) PET Study Group. *Eur J Cancer*. 1999; 35(13):1773–82. Epub 2000/02/16. doi: S0959804999002294 [pii]. PMID: [10673991](https://pubmed.ncbi.nlm.nih.gov/10673991/).
3. Wahl RL, Jacene H, Kasamon Y, Lodge MA. From RECIST to PERCIST: Evolving Considerations for PET response criteria in solid tumors. *J Nucl Med*. 2009; 50 Suppl 1:122S–50S. Epub 2009/06/24. doi: 50/Suppl_1/122S [pii] doi: [10.2967/jnumed.108.057307](https://doi.org/10.2967/jnumed.108.057307) PMID: [19403881](https://pubmed.ncbi.nlm.nih.gov/19403881/); PubMed Central PMCID: PMC2755245.
4. Sunaga N, Oriuchi N, Kaira K, Yanagitani N, Tomizawa Y, Hisada T, et al. Usefulness of FDG-PET for early prediction of the response to gefitinib in non-small cell lung cancer. *Lung Cancer*. 2008; 59(2):203–10. Epub 2007/10/05. doi: S0169-5002(07)00482-5 [pii] doi: [10.1016/j.lungcan.2007.08.012](https://doi.org/10.1016/j.lungcan.2007.08.012) PMID: [17913282](https://pubmed.ncbi.nlm.nih.gov/17913282/).
5. Kobe C, Scheffler M, Holstein A, Zander T, Nogova L, Lammertsma AA, et al. Predictive value of early and late residual 18F-fluorodeoxyglucose and 18F-fluorothymidine uptake using different SUV measurements in patients with non-small-cell lung cancer treated with erlotinib. *Eur J Nucl Med Mol Imaging*. 2012; 39(7):1117–27. Epub 2012/04/25. doi: [10.1007/s00259-012-2118-8](https://doi.org/10.1007/s00259-012-2118-8) PMID: [22526960](https://pubmed.ncbi.nlm.nih.gov/22526960/).
6. Tiseo M, Ippolito M, Scarlattei M, Spadaro P, Cosentino S, Latteri F, et al. Predictive and prognostic value of early response assessment using 18FDG-PET in advanced non-small cell lung cancer patients treated with erlotinib. *Cancer Chemother Pharmacol*. 2014; 73(2):299–307. Epub 2013/11/22. doi: [10.1007/s00280-013-2356-x](https://doi.org/10.1007/s00280-013-2356-x) PMID: [24258456](https://pubmed.ncbi.nlm.nih.gov/24258456/).
7. Treglia G, Mirk P, Stefanelli A, Rufini V, Giordano A, Bonomo L. 18F-Fluorodeoxyglucose positron emission tomography in evaluating treatment response to imatinib or other drugs in gastrointestinal stromal tumors: a systematic review. *Clin Imaging*. 2012; 36(3):167–75. Epub 2012/05/01. doi: S0899-7071(11)00172-0 [pii] doi: [10.1016/j.clinimag.2011.08.012](https://doi.org/10.1016/j.clinimag.2011.08.012) PMID: [22542374](https://pubmed.ncbi.nlm.nih.gov/22542374/).
8. Kwak EL, Bang YJ, Camidge DR, Shaw AT, Solomon B, Maki RG, et al. Anaplastic lymphoma kinase inhibition in non-small-cell lung cancer. *N Engl J Med*. 2010; 363(18):1693–703. Epub 2010/10/29. doi: [10.1056/NEJMoa1006448](https://doi.org/10.1056/NEJMoa1006448) PMID: [20979469](https://pubmed.ncbi.nlm.nih.gov/20979469/); PubMed Central PMCID: PMC3014291.
9. Shaw AT, Kim DW, Nakagawa K, Seto T, Crino L, Ahn MJ, et al. Crizotinib versus chemotherapy in advanced ALK-positive lung cancer. *N Engl J Med*. 2013; 368(25):2385–94. Epub 2013/06/04. doi: [10.1056/NEJMoa1214886](https://doi.org/10.1056/NEJMoa1214886) PMID: [23724913](https://pubmed.ncbi.nlm.nih.gov/23724913/).
10. Gadgeel SM, Gandhi L, Riely GJ, Chiappori AA, West HL, Azada MC, et al. Safety and activity of alectinib against systemic disease and brain metastases in patients with crizotinib-resistant ALK-rearranged non-small-cell lung cancer (AF-002JG): results from the dose-finding portion of a phase 1/2 study. *Lancet Oncol*. 2014; 15(10):1119–28. Epub 2014/08/26. doi: S1470-2045(14)70362-6 [pii] doi: [10.1016/S1470-2045\(14\)70362-6](https://doi.org/10.1016/S1470-2045(14)70362-6) PMID: [25153538](https://pubmed.ncbi.nlm.nih.gov/25153538/).
11. Shaw AT, Kim DW, Mehra R, Tan DS, Felip E, Chow LQ, et al. Ceritinib in ALK-rearranged non-small-cell lung cancer. *N Engl J Med*. 2014; 370(13):1189–97. Epub 2014/03/29. doi: [10.1056/NEJMoa1311107](https://doi.org/10.1056/NEJMoa1311107) PMID: [24670165](https://pubmed.ncbi.nlm.nih.gov/24670165/); PubMed Central PMCID: PMC4079055.
12. Travis WD, Brambilla E, Noguchi M, Nicholson AG, Geisinger KR, Yatabe Y, et al. International association for the study of lung cancer/american thoracic society/european respiratory society international multidisciplinary classification of lung adenocarcinoma. *J Thorac Oncol*. 2011; 6(2):244–85. Epub 2011/01/22. doi: [10.1097/JTO.0b013e318206a221](https://doi.org/10.1097/JTO.0b013e318206a221) 01243894-201102000-00004 [pii]. PMID: [21252716](https://pubmed.ncbi.nlm.nih.gov/21252716/).

13. Thunnissen E, Bubendorf L, Dietel M, Elmberger G, Kerr K, Lopez-Rios F, et al. EML4-ALK testing in non-small cell carcinomas of the lung: a review with recommendations. *Virchows Arch.* 2012; 461(3):245–57. Epub 2012/07/25. doi: [10.1007/s00428-012-1281-4](https://doi.org/10.1007/s00428-012-1281-4) PMID: [22825000](https://pubmed.ncbi.nlm.nih.gov/22825000/); PubMed Central PMCID: [PMC3432214](https://pubmed.ncbi.nlm.nih.gov/PMC3432214/).
14. Boellaard R, O'Doherty MJ, Weber WA, Mottaghy FM, Lonsdale MN, Stroobants SG, et al. FDG PET and PET/CT: EANM procedure guidelines for tumour PET imaging: version 1.0. *Eur J Nucl Med Mol Imaging.* 2010; 37(1):181–200. Epub 2009/11/17. doi: [10.1007/s00259-009-1297-4](https://doi.org/10.1007/s00259-009-1297-4) PMID: [19915839](https://pubmed.ncbi.nlm.nih.gov/19915839/); PubMed Central PMCID: [PMC2791475](https://pubmed.ncbi.nlm.nih.gov/PMC2791475/).
15. Boellaard R, Oyen WJ, Hoekstra CJ, Hoekstra OS, Visser EP, Willemsen AT, et al. The Netherlands protocol for standardisation and quantification of FDG whole body PET studies in multi-centre trials. *Eur J Nucl Med Mol Imaging.* 2008; 35(12):2320–33. Epub 2008/08/16. doi: [10.1007/s00259-008-0874-2](https://doi.org/10.1007/s00259-008-0874-2) PMID: [18704407](https://pubmed.ncbi.nlm.nih.gov/18704407/).
16. de Groot EH, Post N, Boellaard R, Wagenaar NR, Willemsen AT, van Dalen JA. Optimized dose regimen for whole-body FDG-PET imaging. *EJNMMI Res.* 2013; 3(1):63. Epub 2013/08/14. doi: 2191-219X-3-63 [pii] doi: [10.1186/2191-219X-3-63](https://doi.org/10.1186/2191-219X-3-63) PMID: [23938036](https://pubmed.ncbi.nlm.nih.gov/23938036/).
17. Cheebsumon P, Boellaard R, de Ruyscher D, van Elmpt W, van Baardwijk A, Yaqub M, et al. Assessment of tumour size in PET/CT lung cancer studies: PET- and CT-based methods compared to pathology. *EJNMMI Res.* 2012; 2(1):56. Epub 2012/10/05. doi: 2191-219X-2-56 [pii] doi: [10.1186/2191-219X-2-56](https://doi.org/10.1186/2191-219X-2-56) PMID: [23034289](https://pubmed.ncbi.nlm.nih.gov/23034289/).
18. Chen Z, Sasaki T, Tan X, Carretero J, Shimamura T, Li D, et al. Inhibition of ALK, PI3K/MEK, and HSP90 in murine lung adenocarcinoma induced by EML4-ALK fusion oncogene. *Cancer Res.* 2010; 70(23):9827–36. Epub 2010/10/19. doi: 0008-5472.CAN-10-1671 [pii] doi: [10.1158/0008-5472.CAN-10-1671](https://doi.org/10.1158/0008-5472.CAN-10-1671) PMID: [20952506](https://pubmed.ncbi.nlm.nih.gov/20952506/); PubMed Central PMCID: [PMC3043107](https://pubmed.ncbi.nlm.nih.gov/PMC3043107/).
19. Choi H, Paeng JC, Kim DW, Lee JK, Park CM, Kang KW, et al. Metabolic and metastatic characteristics of ALK-rearranged lung adenocarcinoma on FDG PET/CT. *Lung Cancer.* 2013; 79(3):242–7. Epub 2012/12/25. doi: S0169-5002(12)00642-3 [pii] doi: [10.1016/j.lungcan.2012.11.021](https://doi.org/10.1016/j.lungcan.2012.11.021) PMID: [23261227](https://pubmed.ncbi.nlm.nih.gov/23261227/).
20. Katayama R, Khan TM, Benes C, Lifshits E, Ebi H, Rivera VM, et al. Therapeutic strategies to overcome crizotinib resistance in non-small cell lung cancers harboring the fusion oncogene EML4-ALK. *Proc Natl Acad Sci U S A.* 2011; 108(18):7535–40. Epub 2011/04/20. doi: 1019559108 [pii] doi: [10.1073/pnas.1019559108](https://doi.org/10.1073/pnas.1019559108) PMID: [21502504](https://pubmed.ncbi.nlm.nih.gov/21502504/); PubMed Central PMCID: [PMC3088626](https://pubmed.ncbi.nlm.nih.gov/PMC3088626/).
21. van Gool MH, Aukema TS, Schaake EE, Rijna H, Codrington HE, Valdes Olmos RA, et al. (18)F-fluorodeoxyglucose positron emission tomography versus computed tomography in predicting histopathological response to epidermal growth factor receptor-tyrosine kinase inhibitor treatment in resectable non-small cell lung cancer. *Ann Surg Oncol.* 2014; 21(9):2831–7. Epub 2014/05/23. doi: [10.1245/s10434-014-3791-6](https://doi.org/10.1245/s10434-014-3791-6) PMID: [24845729](https://pubmed.ncbi.nlm.nih.gov/24845729/).