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Combined in-gel tryptic digestion and CNBr cleavage for the generation of peptide maps of an integral membrane protein with MALDI-TOF mass spectrometry

Bart A. van Montfort,1, Mark K. Doeven,1 Benito Canas, Liesbeth M. Veenhoff, Bert Poolman,* George T. Robillard

1Department of Biochemistry, Groningen Biomolecular Sciences and Biotechnology Institute, University of Groningen, Nijenborgh 4, 9747 AG Groningen, The Netherlands
bImperial Cancer Research Fund, London, UK

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Abstract

A limitation of the in-gel approaches for the generation of peptides of membrane proteins is the size and hydrophobicity of the fragments generated. For membrane proteins like the lactose transporter (LacS) of Streptococcus thermophilus, tryptic digestion or CNBr cleavage yields several hydrophobic fragments larger than 3.5 kDa. As a result, the sequence coverage of the membrane domain is low when the in-gel tryptic-digested or CNBr-cleaved fragments are analyzed by matrix-assisted laser desorption/ionization-time-of-flight (MALDI-TOF) mass spectrometry (MS). The combination of tryptic digestion and subsequent CNBr cleavage on the same gel pieces containing LacS approximately doubled the coverage of the hydrophobic membrane domain compared to the individual cleavage methods, while the coverage of the soluble domain remained complete. The fragments formed are predominantly below m/z 2500, which allows accurate mass measurement.

Keywords: Mass spectrometry; Peptide mapping; Membrane protein; Matrix-assisted laser desorption/ionization; CNBr; trypsin

1. Introduction

The generation of peptide maps of proteins by in-gel approaches is used often in proteomics-type studies [1–4]. This approach combines the high separation capacity of sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE), in one or two dimensions, with the speed, ease, and sensitivity of mass spectrometry (MS) to identify proteins by peptide mass fingerprinting at a high throughput. The in-gel approach can also be used for the generation of peptide maps of membrane proteins [5–7]. However, high sequence coverage is difficult to obtain because of the lack—or limited accessibility—of proteolytic cleavage sites in the membrane-spanning segments. This results in large fragments that are highly hydrophobic, which limits the use of MS for protein identification or structure–function analysis of membrane proteins. The latter application is of great interest for the analysis of the structure of membrane proteins such as the lactose transporter (LacS) from Streptococcus thermophilus. The LacS protein has been recently shown to function as a cooperative dimer in proton motive force-driven lactose uptake [8,9]. Peptide maps of LacS with high sequence coverage would help the identification of the dimer interface when analysed by cross-linking methods. The same holds true for the helix packing models of LacS and other members of the GPH family, which are largely based, thus far, on the analysis of second-site suppressor mutants [10].

Recently, we reported in-gel approaches to generate peptide maps of integral membrane proteins, including LacS, using tryptic digestion and CNBr cleavage [11]. The sequence coverage of membrane domains in tryptic
maps was increased when the matrix-assisted laser desorption/ionization-time-of-flight (MALDI-TOF)-compatible detergent octyl-β-glucopyranoside (OBG) was used in the extraction solvents. In-gel CNBr cleavage of the membrane domains resulted in spectra that were 5- to 10-fold more intense than those obtained from in-gel tryptic digestion. A limitation of an in-gel approach is the size of the fragments generated. In general, it is difficult to extract larger fragments from the gel pieces. This disadvantage can be overcome by generating smaller fragments with a combination of different proteases or chemical cleavage methods in one sample.

The combined use of tryptic digestion and CNBr cleavage is presented here for LacS. Peptide mapping of this integral membrane protein with our original procedures resulted in maps, which missed several, predominantly large and hydrophobic peptides [11]. In-gel tryptic digestion followed by CNBr cleavage on the same gel pieces, as presented here, resulted in intense spectra from peptides that were mostly below $m/z$ 2500. The method approximately doubled the sequence coverage of the membrane domain. Moreover, the addition of OBG to the extraction solvents resulted in the recovery of peptides at higher $m/z$ values, which increased the sequence coverage even more.

2. Methods and materials

2.1. Materials

Chemicals and solvents used in this study were of analytical grade. The purification of his-tagged lactose transporter of *S. thermophilus*, in which Cys-320 was replaced with an alanine (LacS-C320A), was done as described [12]. Trypsin for in-gel tryptic digestions was sequencing grade-modified trypsin from Promega. OBG was purchased from Sigma. CNBr was from Fluka.

2.2. SDS-PAGE

Approximately 3 μg of LacS (40 pmol) was solubilized in Laemmli sample buffer and heated for 10 min at 50 °C. The samples were separated on Laemmli 10% SDS-poly-

![Fig. 1. The cleavage sites of trypsin and CNBr are indicated in the topology model of LacS. Trypsin cleaves at the C-terminal side of lysines and arginines as indicated by the arrows. CNBr cleavage sites at the C-terminal side of methionines are shown in black.](image-url)
Fig. 2. MALDI-TOF mass spectra of in-gel generated peptides of LacS. (A) Tryptic digest, extracted without OBG; (B) CNBr cleavage, extracted without OBG; (C) combined trypsin/CNBr cleavage, extracted without OBG; (D) combined trypsin/CNBr cleavage, extracted with OBG. (*) Peaks of fragments of the hydrophobic membrane-embedded part of LacS. The insets show the sequence coverage depicted in the topology model of LacS.
acrylamide gels [13] and visualized with Coomassie blue staining.

2.3. In-gel tryptic digestion, CNBr cleavage, or the combination of both

After visualization, the bands containing the protein were excised from SDS-polyacrylamide gels, treated and subjected to digestion with trypsin or CNBr cleavage, following a modified procedure of Hellman et al. [14] as described in Ref. [11]. Tryptic digestion or CNBr cleavage was started by the addition of 5 µl of 75 ng/µl trypsin in 25 mM ammonium bicarbonate, pH 8, or 25 µl of CNBr in 70% TFA (one small crystal was dissolved in 200–300 µl of 70% TFA), respectively, to the dried gel pieces. For the tryptic digestions, the gel pieces were covered with an overlay of 25 mM ammonium bicarbonate (approximately 20 µl) after reswelling and incubated for at least 14 h at 30 °C. The CNBr cleavage was performed for at least 14 h in the dark at room temperature. The combined tryptic and CNBr cleavage was performed as follows. After tryptic digestion, the peptides were not extracted, but the gel pieces were dried in the SpeedVac. These dried gel pieces were washed and dehydrated twice by addition of 100 µl of acetonitrile followed by SpeedVac drying. Subsequently, CNBr cleavage was started by the addition of 25 µl of CNBr in 70% TFA as above. After collection of the supernatant, the peptides were extracted twice by sonication for 5 min in 30 µl of 60% acetonitrile, 1% TFA in the absence or presence of 0.1% OBG. The overlay and extracts were pooled and dried in a SpeedVac. The last traces of ammonium bicarbonate were removed by adding 10 µl of 1% TFA and subsequent drying in the SpeedVac.

2.4. MS

The samples were dissolved in 5 µl of 50% acetonitrile, 0.1% TFA, and sonicated for 5 min. Aliquots of 0.75 µl were applied onto the target and allowed to air dry. Subsequently, 0.75 µl of 10 mg/ml α-cyano-4-hydroxysuccinimide acid in 50% acetonitrile, 0.1% (v/v) TFA was applied to the dried sample and again allowed to dry. MALDI-TOF mass spectra were recorded with a Micromass Tofspec E MALDI time-of-flight mass spectrometer operated in reflectron mode. The spectra were calibrated externally, but if necessary, the spectra were internally recalibrated on trypsin autodigestion products and a matrix peak.

3. Results and discussion

The lactose transporter of S. thermophilus is a 70-kDa integral membrane protein with a 50-kDa N-terminal membrane-embedded part (membrane domain), a 20-res-
which allowed very accurate mass determination and will simplify sequencing by MS/MS.

We suggested previously that the low efficiency of the tryptic in-gel approach was caused by the limited accessibility of the gel-trapped protein for the protease [11]. Upon removal of SDS from the gel pieces, it is likely that the membrane protein aggregates and becomes inaccessible for the protease, which would explain the low intensity of the spectra of the tryptic peptides of the membrane domain. If the accessibility were the problem, the spectrum of the combined method (Fig. 2C and D) would predominantly contain CNBr-cleaved fragments as in Fig. 2B. Since this is not the case, it suggests that the extraction, the solubility, and/or the MALDI MS analysis of the tryptic peptides form a bottleneck and not the accessibility of the membrane domain for the protease. Control experiments excluded the possibility that solely the presence of 70% TFA used in the CNBr cleavage helped the solubilisation of the hydrophobic fragments, since a subsequent incubation of an in-gel tryptic digest in 70% TFA (as in the CNBr cleavage) did not increase the sequence coverage (not shown).

4. Conclusion

The combined use of in-gel tryptic digestion and CNBr cleavage results in intense spectra with a high sequence coverage of the membrane domain and complete coverage of the soluble domain of a large hydrophobic integral membrane protein. The combined method can be used for the identification of structure–function relations as mentioned above. In addition, this method should, in principle, be applicable for many other membrane proteins and make this class of proteins amenable to proteome analysis. Although conventional 2D electrophoresis with isoelectric focusing in the first dimension is often not possible with hydrophobic proteins, the use of blue native electrophoresis in the first dimensional separation could represent a good alternative [15]. On the assumption that individual membrane proteins constitute 0.1–1% of the total membrane protein fraction, 2–20 pmol of protein should be present per spot on a 2D gel, which should be sufficient to identify the protein. Preliminary experiments indicated that it is possible to obtain a high sequence coverage of LacS using the combined method and only 2–4 pmol of protein.

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