Chapter 6

High urinary excretion of Kidney Injury Molecule-1 (KIM-1) is an independent predictor of graft loss in renal transplant recipients

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ABSTRACT

Background
Chronic transplant dysfunction is characterized by renal function decline and proteinuria. Kidney Injury Molecule-1 (KIM-1), a transmembrane tubular protein with unknown function, is undetectable in normal kidneys, but markedly induced after injury. Urinary KIM-1 excretion has been quantified as biomarker of renal damage. We prospectively studied whether urinary KIM-1 predicts graft loss, independent of renal function and proteinuria.

Methods
Renal transplant recipients (n = 145) visiting our out-patient clinic between August 2001 and July 2003 collected 24h urine samples for assessment of baseline urinary KIM-1 excretion (microsphere-based Luminex technology), and were followed for graft loss.

Results
Recipients participated at a median (interquartile range) of 6.0 (2.5-12.0) years post-transplant in baseline measurements. Follow-up beyond baseline was 4.0 (3.2-4.5) years. Urinary KIM-1 excretion was 0.72 (0.42-1.37) ng/24h. Occurrence of graft loss increased over tertiles of KIM-1 excretion: 3 (6.3%), 11 (22.4%), and 17 cases (35.4%) (P = 0.001), respectively. High KIM-1 excretion was associated with proteinuria, low creatinine clearance and high donor age (all P < 0.01). In multivariate Cox regression analyses, prediction of graft loss by KIM-1 appeared independent of creatinine clearance, proteinuria, and donor age. Hazard ratios (95% CI) for the 2nd and 3rd tertile of KIM-1 excretion were 3.6 (0.9-13.5) and 5.1 (1.5-17.8) in the final model.

Conclusions
Urinary excretion of KIM-1 is an independent predictor of long-term graft loss and therefore a promising new biomarker in early prediction of graft loss.
INTRODUCTION
Although the incidence of acute renal allograft rejection has decreased progressively during the last decades, there has been little improvement in long-term graft survival [1]. Chronic transplant dysfunction (CTD), one of the leading causes of late allograft loss [2], is characterized clinically by a gradual decline in renal function with proteinuria and hypertension [3,4]. This is accompanied by histological changes, i.e. interstitial fibrosis, tubular atrophy, vascular occlusion, and glomerulosclerosis [5-7]. Non-invasively, estimates of glomerular filtration rate (e.g. creatinine clearance or plasma creatinine) and proteinuria are used for identification of transplant recipients at increased risk for late renal allograft loss [8-11]. However, once serum creatinine starts to rise, or proteinuria develops, chronic structural lesions are already present, and it is usually too late for intervention [12,13]. Thus, there is a great need for non-invasive markers of CTD that can predict graft loss in an earlier stage [14].

Kidney Injury Molecule-1 (KIM-1) is a recently discovered transmembrane protein with Ig-like and mucin domains in its ectodomain, which is undetectable in normal kidneys, but markedly upregulated in damaged tubular epithelial cells after different types of renal injury [15-19]. KIM-1 ectodomain can be shed into the urine and it was previously shown that urinary excretion of KIM-1 relates to the extent of renal damage in experimental renal disease [19,20] and in various inflammatory and fibrotic human renal diseases [21]. We hypothesized that urinary KIM-1 excretion predicts long-term graft loss in renal transplant patients. Therefore, in this prospective study we investigated whether urinary KIM-1 excretion is a predictor of graft loss independent of creatinine clearance and proteinuria in renal transplant recipients.

METHODS
Study design and patients
The current prospective study was part of a larger study and incorporated in the Groningen Renal Transplant Outpatient Program, details of which have been published previously [22-24]. Between August 2001 and July 2003, all adult renal allograft recipients, who had a functioning graft for at least 1 year, were eligible to participate. Patients with known or apparent systemic illnesses (e.g. malignancies or opportunistic infections) were considered not eligible. A total of 606 out of 847 (72%) eligible renal transplant recipients signed written informed consent. Mortality and graft loss were recorded for all renal transplant recipients until June 27, 2006. Graft loss was censored for death and defined as return to dialysis or re-transplantation. For the current study, data processing and analyses are based on a case-cohort design: a random sample of the cohort (n=118) was enriched with remaining cases of graft loss (n=27) in the entire cohort. For patients who died with a functioning graft (n=15 in our study population) duration of follow-up was calculated until date of death. For patients with graft loss (n=31) duration of follow-up was calculated using date of start of dialysis or re-transplantation. The Institutional Review Board approved the study protocol (METc 01/039). Funding sources had neither a role in the collection and analyses of data, nor in publication of the manuscript.
Renal transplant characteristics
Relevant transplant characteristics were taken from the Groningen Renal Transplant Database. This database contains information on all renal transplantations performed at our center since 1968. Current medication was taken from the medical record. Standard immunosuppressive treatment was described previously [22-24].

Processing and storage of urinary samples
After collection, fresh 24h urinary samples were centrifuged. An aliquot was used for routine measurements, i.e. creatinine and protein, and an aliquot was immediately stored (-80°C) until KIM-1 measurement.

Baseline measurements
BMI and blood pressure were measured as described previously [23]. Blood was drawn after an 8- to 12-hour overnight fasting period. Serum total cholesterol, high-density lipoprotein cholesterol (HDL) and non-HDL cholesterol were assessed as described previously [23,24]. Serum creatinine levels were determined using a modified version of the Jaffé method (MEGA AU 510, Merck Diagnostica, Darmstadt, Germany). Creatinine clearance was calculated from 24h urinary creatinine excretion and serum creatinine. Estimated GFR (eGFR) was calculated with the MDRD formula [25]. Total urinary protein concentration was analysed using the Biuret reaction (MEGA AU 510, Merck Diagnostica, Darmstadt, Germany) and proteinuria was defined as urinary protein excretion >0.5 g/24h.

Measurement of urinary KIM-1 level
Urinary KIM-1 measurements were performed using microsphere-based Luminex xMAP technology with polyclonal antibodies raised against the human KIM-1 ectodomain, as described previously [26]. This technique is an adaptation of the sandwich ELISA assay, as described previously [16,20]. For measurements, 30 µl of urine samples were analyzed in duplicate. The lowest limit of detection for this assay is 0.125 ng/ml. The inter- and intra-assay variability is <10%. The urinary KIM-1 level was expressed in absolute terms (ng/ml). Urinary KIM-1 excretion was calculated by multiplication with the volume of the 24h urine collection, resulting in KIM-1 excretion expressed in ng/24h.

Statistical Analyses
Analyses were performed with SPSS version 14.0 (SPSS Inc., Chicago, IL). Parametric variables are given as means ± SD. Non-parametric variables are given as median (interquartile range). Subjects were divided in tertiles based on urinary KIM-1 excretion; differences between the tertiles were tested for statistical significance with one-way ANOVA in case of a parametric variable or the Kruskal-Wallis test in case of a non-parametric variable; the Chi-square test was used in case of a categorical variable. Kaplan-Meier survival analysis with log-rank testing was performed for prospective analysis of graft loss. We then proceeded with univariate and multivariate cox-regression analyses of urinary excretion of KIM-1, creatinine clearance, proteinuria and other
potential determinants of graft loss. In the final multivariate model we included urinary excretion of KIM-1, donor age, creatinine clearance and proteinuria. To further compare the predictive performance of urinary KIM-1 excretion with creatinine clearance, proteinuria and donor age we also generated receiver operator characteristic (ROC) curves. The areas under the ROC curve of KIM-1 and creatinine clearance, proteinuria and donor age were compared nonparametrically by the method of DeLong, DeLong and Clarke-Pearson [27]. A two-sided P-value <0.05 was considered to indicate statistical significance.

RESULTS
Recipients participated at a median (interquartile range) of 6.0 (2.5-12.0) years post-transplant in baseline measurements. Follow-up for graft loss beyond baseline was 4.0 (3.2-4.5) years. KIM-1 excretion was 0.72 (0.42-1.37) ng/24h. Minimum and maximum values were 0.01 and 10.0 ng/24h, respectively. Recipient-related baseline characteristics according to tertiles of KIM-1 excretion are shown in Table 1. There were no significant differences in recipient-related characteristics across the tertiles.

Table 1. Recipient-related baseline parameters of the population subdivided by tertiles of KIM-1 (median with range is given).

<table>
<thead>
<tr>
<th>Tertiles of KIM-1 (ng/24h)</th>
<th>0.30 (0.01-0.48)</th>
<th>0.72 (0.49-1.09)</th>
<th>1.69 (1.15-10.04)</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td>n=48</td>
<td>n=49</td>
<td>n=48</td>
<td></td>
<td></td>
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<tr>
<td><strong>Recipient characteristics</strong></td>
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<tr>
<td>Tx until baseline (years)</td>
<td>5.6 (2.5-11.6)</td>
<td>6.1 (2.0-14.4)</td>
<td>7.4 (3.0-11.2)</td>
<td>0.85</td>
</tr>
<tr>
<td>Dialysis prior Tx (months)</td>
<td>28 (18-46)</td>
<td>24 (11-33)</td>
<td>30 (10-57)</td>
<td>0.18</td>
</tr>
<tr>
<td>Age (years)</td>
<td>52.7 ± 12.4</td>
<td>52.9 ± 11.4</td>
<td>52.7 ± 13.6</td>
<td>1.00</td>
</tr>
<tr>
<td>Male gender, n (%)</td>
<td>24 (50)</td>
<td>30 (61)</td>
<td>26 (54)</td>
<td>0.53</td>
</tr>
<tr>
<td>Body Mass Index (kg/m²)</td>
<td>26.3 ± 4.4</td>
<td>25.6 ± 3.8</td>
<td>26.6 ± 3.7</td>
<td>0.44</td>
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<tr>
<td>Blood pressure</td>
<td></td>
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<tr>
<td>MAP (mm Hg)</td>
<td>110 ± 13</td>
<td>112 ± 14</td>
<td>116 ± 14</td>
<td>0.13</td>
</tr>
<tr>
<td>AGE inhibitor, n (%)</td>
<td>14 (29)</td>
<td>15 (31)</td>
<td>22 (46)</td>
<td>0.17</td>
</tr>
<tr>
<td>β-blocker, n (%)</td>
<td>34 (71)</td>
<td>24 (49)</td>
<td>25 (52)</td>
<td>0.06</td>
</tr>
<tr>
<td>Current smoking, n (%)</td>
<td>12 (25.0)</td>
<td>12 (24.5)</td>
<td>14 (29.2)</td>
<td>0.85</td>
</tr>
<tr>
<td>Glycaemia</td>
<td></td>
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<tr>
<td>Fasting plasma glucose</td>
<td>4.8 ± 1.3</td>
<td>4.9 ± 1.2</td>
<td>5.0 ± 2.0</td>
<td>0.85</td>
</tr>
<tr>
<td>Posttransplant diabetes, n (%)</td>
<td>13 (27)</td>
<td>9 (18)</td>
<td>7 (15)</td>
<td>0.29</td>
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<tr>
<td>Lipids</td>
<td></td>
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<tr>
<td>Cholesterol (mmol/l)</td>
<td>5.6 (5.2-6.0)</td>
<td>5.5 (4.9-6.3)</td>
<td>5.6 (4.8-6.7)</td>
<td>0.90</td>
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<td>HDL cholesterol (mmol/l)</td>
<td>1.2 (0.9-1.3)</td>
<td>1.1 (0.9-1.3)</td>
<td>1.0 (0.8-1.4)</td>
<td>0.31</td>
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<tr>
<td>Non-HDL cholesterol (mmol/l)</td>
<td>4.4 (4.2-4.6)</td>
<td>4.5 (4.1-4.8)</td>
<td>4.6 (4.3-4.9)</td>
<td>0.72</td>
</tr>
<tr>
<td>Statin, n (%)</td>
<td>26 (54)</td>
<td>26 (53)</td>
<td>23 (48)</td>
<td>0.81</td>
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</tbody>
</table>

Parametric variables are expressed as mean ± SD, non-parametric variables are given as median (interquartile range). Statistical analyses were performed with one-way ANOVA or the Kruskal-Wallis test in case of a non-parametric variable, and the Chi-square test in case of a categorical variable. Abbreviations: Tx, transplantation; MAP, mean arterial pressure; ACE, angiotensin-converting enzyme; HDL, high-density lipoprotein.

Transplant-related characteristics are shown in Table 2. Donor age, serum creatinine, and proteinuria increased significantly with increasing KIM-1 excretion. Creatinine clearance, eGFR, and duration of follow-up beyond baseline measurements decreased significantly
with increasing KIM-1 excretion. Occurrence of graft loss resulting in return to dialysis or re-transplantation (death-censored graft loss) increased significantly with increasing urinary KIM-1 excretion at baseline: 3 (6.3%), 11 (22.4%) and 17 (35.4%), for the consecutive tertiles respectively, $P = 0.001$. A corresponding Kaplan-Meier curve for the tertiles of urinary KIM-1 excretion is shown in Figure 1.

Table 2. Transplant-related parameters of the population subdivided by tertiles of KIM-1 (median with range is given).

<table>
<thead>
<tr>
<th>Tertiles of KIM-1 (ng/24h)</th>
<th>0.30 (0.01-0.48)</th>
<th>0.72 (0.49-1.09)</th>
<th>1.69 (1.15-10.04)</th>
<th>n=48</th>
<th>n=49</th>
<th>n=49</th>
<th>P</th>
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<tr>
<td><strong>Donor characteristics</strong></td>
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<tr>
<td>Age (years)</td>
<td>34.3 ± 15.2</td>
<td>37.5 ± 15.5</td>
<td>44.4 ± 16.0</td>
<td>0.006</td>
<td></td>
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<tr>
<td>Male gender, n (%)</td>
<td>28 (58)</td>
<td>30 (64)</td>
<td>21 (44)</td>
<td>0.13</td>
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<td>Deceased donor transplant, n (%)</td>
<td>42 (86)</td>
<td>43 (88)</td>
<td>40 (83)</td>
<td>0.78</td>
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<tr>
<td><strong>Time</strong></td>
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<tr>
<td>Warm ischemia times (min)</td>
<td>36.0 (32.0-44.5)</td>
<td>36.0 (30.0-45.0)</td>
<td>35.0 (30.0-48.8)</td>
<td>0.86</td>
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<td>Cold ischemia times (h)</td>
<td>20.0 (13.5-26.8)</td>
<td>22.0 (14.0-25.5)</td>
<td>22.5 (10.3-27.0)</td>
<td>0.80</td>
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<td><strong>Acute rejection treatment</strong></td>
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<td>High dose corticosteroids, n (%)</td>
<td>13 (27)</td>
<td>20 (41)</td>
<td>17 (35)</td>
<td>0.36</td>
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<tr>
<td>Other rejection therapy, n (%)</td>
<td>4 (8)</td>
<td>8 (16)</td>
<td>3 (6)</td>
<td>0.23</td>
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<td><strong>Immunosuppression</strong></td>
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<tr>
<td>Prednisolone dose (mg/day)</td>
<td>10 (7.5-10.0)</td>
<td>10 (7.5-10)</td>
<td>10 (7.5-10)</td>
<td>0.90</td>
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<tr>
<td>Cyclosporine, n (%)</td>
<td>29 (60)</td>
<td>23 (47)</td>
<td>31 (65)</td>
<td>0.19</td>
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<td>Tacrolimus, n (%)</td>
<td>8 (17)</td>
<td>14 (29)</td>
<td>6 (13)</td>
<td>0.11</td>
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<tr>
<td>Mycophenolate mofetil, n (%)</td>
<td>23 (48)</td>
<td>18 (37)</td>
<td>15 (31)</td>
<td>0.23</td>
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<tr>
<td>Azathioprine, n (%)</td>
<td>15 (31)</td>
<td>18 (37)</td>
<td>18 (38)</td>
<td>0.78</td>
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<td><strong>Renal allograft function</strong></td>
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<tr>
<td>Serum creatinine (μmol/l)</td>
<td>138 ± 60</td>
<td>157 ± 65</td>
<td>195 ± 86</td>
<td>&lt;0.001</td>
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<tr>
<td>Creatinine clearance (ml/min)</td>
<td>64 ± 22</td>
<td>60 ± 24</td>
<td>49 ± 21</td>
<td>0.004</td>
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<td></td>
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<tr>
<td>eGFR (ml/min)</td>
<td>49.1 ± 14.3</td>
<td>44.7 ± 15.8</td>
<td>35.7 ± 15.3</td>
<td>&lt;0.001</td>
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<tr>
<td>Proteinuria (g/24h)</td>
<td>0.1 (0.0-0.3)</td>
<td>0.2 (0.0-0.5)</td>
<td>0.5 (0.2-1.1)</td>
<td>&lt;0.001</td>
<td></td>
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<td></td>
</tr>
<tr>
<td>Proteinuria, n (%)</td>
<td>8 (17)</td>
<td>12 (25)</td>
<td>24 (50)</td>
<td>0.001</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Follow up since baseline (years)</td>
<td>4.2 (3.6-4.5)</td>
<td>3.8 (3.0-4.3)</td>
<td>3.7 (2.1-4.5)</td>
<td>0.03</td>
<td></td>
<td></td>
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</tr>
<tr>
<td>Death-censored graft loss, n (%)</td>
<td>3 (6.3)</td>
<td>11 (22.4)</td>
<td>17 (35.4)</td>
<td>0.001</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Parametric variables are expressed as mean ± SD, non-parametric variables are given as median (interquartile range). Statistical analyses were performed with one-way ANOVA or the Kruskal-Wallis test in case of a non-parametric variable, and the Chi-square test in case of a categorical variable. Abbreviations: eGFR, estimated glomerular filtration rate.

![Figure 1](image-url)  
Kaplan-Meier survival curve of tertiles of urinary KIM-1 excretion (in ng/24h) at baseline for death-censored graft survival. P-value = 0.001 according to log-rank test.
We subsequently investigated whether KIM-1 excretion is an independent predictor of graft loss (Table 3). After adjustment for donor age and creatinine clearance, hazard ratios of the 2nd and 3rd tertiles of KIM-1 excretion for graft loss decreased, but remained significant. Further adjustment for proteinuria in addition to donor age and creatinine clearance did not materially change the results. Hazard ratios for graft loss of the KIM-1 tertiles in the final model were 3.6 (0.9-13.5) and 5.1 (1.5-17.8), with P = 0.06 and P = 0.01 respectively. These hazard ratios are comparable to hazard ratios for proteinuria: 3.1 ((1.0-9.2), P = 0.04) and 6.0 ((2.3-15.8), P < 0.001) for urinary protein excretion > 0.5 and ≤1 g/24h and for urinary protein excretion > 1 g/24h respectively, in the final model. Adjustment for eGFR and serum creatinine instead of creatinine clearance did not materially change the results. In univariate analyses, donor age was a significant predictor of graft loss, but this association was no longer significant after adjustment for creatinine clearance, serum creatinine, or eGFR in the Cox-regression models (data not shown). Subsequently we investigated whether the relationship between KIM-1 excretion and graft survival persists if analysed in relation to time since transplantation rather than time since baseline measurements. We first investigated whether KIM-1 excretion at baseline is related to time since transplantation. It appeared that there is no significant correlation between KIM-1 excretion and time since transplantation (r=0.05; P = 0.53). Also, the relation between KIM-1 excretion and graft survival persists if analysed in relation to time since transplantation rather than time since baseline measurements.

Table 3. Univariate and multivariate Cox regression analyses of determinants of death-censored graft loss in renal transplant recipients.

<table>
<thead>
<tr>
<th>KIM-1 tertiles</th>
<th>Tertile I</th>
<th>Tertile II</th>
<th>Tertile III</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Reference</td>
<td>HR (95% CI)</td>
<td>P</td>
</tr>
<tr>
<td>Unadjusted model</td>
<td>1.0</td>
<td>4.2 (1.2-15.0)</td>
<td>0.03</td>
</tr>
<tr>
<td>Adjusted for donor age</td>
<td>1.0</td>
<td>3.9 (1.1-13.9)</td>
<td>0.04</td>
</tr>
<tr>
<td>Adjusted for donor age, CrCl</td>
<td>1.0</td>
<td>3.2 (0.9-11.8)</td>
<td>0.08</td>
</tr>
<tr>
<td>Adjusted for donor age, CrCl, Uprot</td>
<td>1.0</td>
<td>3.6 (0.9-13.5)</td>
<td>0.06</td>
</tr>
</tbody>
</table>

Proteinuria was divided in three groups, the first ≤ 0.5 g/24; the second >0.5 and ≤ 1.0 g/24; the third >1.0 g/24 h. Abbreviations: CrCl, creatinine clearance; Uprot, proteinuria.

In addition, we performed a multivariate analysis with continuous variables. In this model creatinine clearance (HR=0.92 per ml/min, P < 0.001), proteinuria (HR=1.34 per g/24h, P = 0.0001) and KIM-1 (HR=1.31 per ng/24h, P = 0.02) appeared independent predictors of graft loss. Donor age (HR=1.01 per year, P = 0.40) was not an independent predictor of graft loss in this model. These results for continuous variables did not materially differ from the analyses with categorical variables.

Finally, we compared the predictive performance of urinary KIM-1 excretion with creatinine clearance, proteinuria and donor age by generating ROC curves (Figure 2). ROC analysis of prediction of graft loss by KIM-1 excretion revealed a mean (SE) area under the curve of 0.71 (0.05; P < 0.001). The area under the curve for creatinine clearance was 0.89 (0.03; P < 0.001), which was significantly higher than for KIM-1 (P = 0.002 for creatinine clearance versus KIM-1). The area under the curve for proteinuria was 0.82 (0.05;
P < 0.001) and for donor age 0.65 (0.05; P < 0.001). Proteinuria and donor age were equal predictors of graft loss when compared to KIM-1 (P = ns).

**Figure 2**
Areas under the Receiver Operating Curves (ROC) with standard error (SE) of KIM-1, creatinine clearance, proteinuria and donor age at baseline for graft loss were: 0.71 (0.05; P<0.001), 0.89 (0.03; P<0.001), 0.82 (0.05; P<0.001), and 0.65 (0.05; P<0.001), respectively. The areas under the ROC curve were compared nonparametrically by the method of DeLong and Clarke-Pearson. The area under the curve for creatinine clearance was significantly higher than for KIM-1 (P=0.002), for proteinuria and donor age this did not differ (P=ns).

**DISCUSSION**
In this longitudinal prospective study we investigated whether urinary KIM-1 excretion could be of use in the prediction of graft loss in renal transplant recipients in addition to established markers as creatinine clearance and proteinuria. We found that high KIM-1 excretion is a predictor of graft loss, independent of donor age, creatinine clearance and proteinuria, with hazard ratios of similar magnitude as proteinuria.
To the best of our knowledge this study is the first prospective study on urinary KIM-1 excretion as potential predictor of graft loss in renal transplant recipients. The only other prospective study concerning KIM-1 to date is by Liangos et al [26]. They have shown in a cohort of 201 hospitalized patients with established acute renal failure that urinary KIM-1 excretion was predictive for adverse clinical outcomes, i.e. patients in the highest KIM-1 quartile had 3.2-fold higher odds for dialysis requirement or hospital death than patients in the lowest quartile.
For the association between KIM-1 and progressive decline in renal function several potential explanations are possible. KIM-1 is a type 1 transmembrane glycoprotein that is expressed in humans and rodents when injured renal tubules adopt a dedifferentiated phenotype [17,21]. Expression of KIM-1 is also found in renal tumor cells and in tubular cells adjacent to carcinoma cells; these cells may be compressed by the tumor cells and consequently adopt a dedifferentiated phenotype [28,29]. Cleavage of the KIM-1 ectodomain by matrix metalloproteinases results in KIM-1 excretion into the urine [30]. Urinary excretion of KIM-1 is therefore a reflection of the degree of histopathological tubular damage that exists at a certain timepoint [19-21]. Since renal damage and tubular
dedifferentiation have been shown to predict renal function decline and graft loss [31,32].
KIM-1 may be a non-invasively assessable marker of these two entities.
Up to now the function of KIM-1 is unclear. Based on its protein structure, with Ig- and
mucin-domains in the extracellular part, and its close resemblance to adhesion molecules,
KIM-1 could function in cel/cel or cel/matrix interactions. KIM-1 is also known as T cell
immunoglobulin mucin-like domain 1 (TIM-1), and as such it is expressed on T cells.
Meyers et al [33] have demonstrated that Tim-4, which is exclusively expressed on
antigen-presenting cells in the mouse, is the natural ligand for Tim-1. This suggests that
KIM-1 can interact with other proteins. KIM-1 expression is induced after different types of
renal injury, but we can only speculate whether tubular KIM-1 expression is a harmful or
protective mechanism. From ischemia/reperfusion studies it is known that KIM-1
expression persists during the first recovery phase, indicating that KIM-1 might not always
be associated with a deleterious outcome. Furthermore, when renal damage is reduced
with renoprotective interventions KIM-1 expression is also reduced [34,35].
In our Cox regression analysis model we included – besides donor age, proteinuria and
KIM-1 – creatinine clearance, which was also an independent predictor of graft loss.
However, as 24h urine samples are not readily available in many clinics and/or studies, we
also tested the regression model with the eGFR (calculated with the MDRD formula) that
can be calculated from serum creatinine. When substituting creatinine clearance with
eGFR, KIM-1 was still an independent predictor of graft loss.
Donor age is a commonly accepted predictor of renal function decline, and also in our
study we found that higher KIM-1 levels were associated with a higher donor age.
However, donor age did not independently predict graft loss in the multivariate analysis.
Renal function, i.e. creatinine clearance, is highly associated with age, and because renal
function is a strong risk factor for graft failure this can explain why donor age did not
significantly contribute to the multivariate model for prediction of graft loss. This is in
concordance with other studies showing that donor age alone is not a risk factor for worse
long-term outcomes as long as there was no history of hypertension and kidney function
was normal [36-38].
The present study has some limitations. First, the present study is a single-center study
and the predictive value of KIM-1 excretion needs to be confirmed in other centra and/or
multi-center studies. Second, the renal transplant recipients were included at different
timepoints after transplantation, this could possibly induce a healthy survivor bias, and
therefore it is advisable to study the predictive value of urinary KIM-1 excretion on graft
survival at a fixed timepoint, for example 1 year, after transplantation. Furthermore,
urinary KIM-1 excretion should be analysed in relation to the histological diagnosis. Also,
in our study, creatinine clearance, proteinuria, and KIM-1 excretion were assessed from
samples taken at one time-point in each patient. It would be interesting to investigate in a
future study whether sequential measurements of KIM-1 excretion could be used as an
even earlier marker, and can be used to predict development of proteinuria and rises in
plasma creatinine.
In conclusion, elevated urinary levels of KIM-1 independently predict long-term graft loss
in renal transplant recipients. Therefore, measuring urinary KIM-1 excretion is of additional
value, next to measurement of creatinine clearance and proteinuria, for identification of subjects at risk for renal allograft failure who may benefit from intervention.

REFERENCES
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