

University of Groningen

Dissecting yeast-dependent population differentiation and spatial segregation in *Drosophila melanogaster*

Wang, Xiaocui

DOI:

[10.33612/diss.249063971](https://doi.org/10.33612/diss.249063971)

IMPORTANT NOTE: You are advised to consult the publisher's version (publisher's PDF) if you wish to cite from it. Please check the document version below.

Document Version

Publisher's PDF, also known as Version of record

Publication date:

2022

[Link to publication in University of Groningen/UMCG research database](#)

Citation for published version (APA):

Wang, X. (2022). *Dissecting yeast-dependent population differentiation and spatial segregation in Drosophila melanogaster*. [Thesis fully internal (DIV), University of Groningen]. University of Groningen. <https://doi.org/10.33612/diss.249063971>

Copyright

Other than for strictly personal use, it is not permitted to download or to forward/distribute the text or part of it without the consent of the author(s) and/or copyright holder(s), unless the work is under an open content license (like Creative Commons).

The publication may also be distributed here under the terms of Article 25fa of the Dutch Copyright Act, indicated by the "Taverne" license. More information can be found on the University of Groningen website: <https://www.rug.nl/library/open-access/self-archiving-pure/taverne-amendment>.

Take-down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

Downloaded from the University of Groningen/UMCG research database (Pure): <http://www.rug.nl/research/portal>. For technical reasons the number of authors shown on this cover page is limited to 10 maximum.

References

A

- Agrawal, S., Safarik, S., and Dickinson, M. (2014). The relative roles of vision and chemosensation in mate recognition of *Drosophila melanogaster*. *J Exp Biol* 217(Pt 15), 2796-2805. doi: 10.1242/jeb.105817.
- Allan, C.W., and Matzkin, L.M. (2019). Genomic analysis of the four ecologically distinct cactus host populations of *Drosophila mojavensis*. *BMC Genomics* 20(1), 732. doi: 10.1186/s12864-019-6097-z.
- Amlou, M., Moreteau, B., and David, J.R. (1998). Larval tolerance in the *Drosophila melanogaster* species complex toward the two toxic acids of the *D. sechellia* host plant. *Hereditas* 129(1), 7-14. doi: 10.1111/j.1601-5223.1998.00007.x.
- Anagnostou, C., Dorsch, M., and Rohlf, M. (2010). Influence of dietary yeasts on *Drosophila melanogaster* life-history traits. *Entomol. Exp. Appl.* 136(1), 1-11. doi: 10.1111/j.1570-7458.2010.00997.x.
- Anderson, P., and Anton, S. (2014). Experience-based modulation of behavioural responses to plant volatiles and other sensory cues in insect herbivores. *Plant Cell Environ.* 37(8), 1826-1835. doi: 10.1111/pce.12342.
- Andrade Lopez, J.M., Lanno, S.M., Auerbach, J.M., Moskowitz, E.C., Sligar, L.A., Wittkopp, P.J., et al. (2017). Genetic basis of octanoic acid resistance in *Drosophila sechellia*: functional analysis of a fine-mapped region. *Mol Ecol* 26(4), 1148-1160. doi: 10.1111/mec.14001.
- Anholt, R.R.H. (2020). Chemosensation and evolution of *Drosophila* host plant selection. *iScience* 23(1), 100799. doi: 10.1016/j.isci.2019.100799.
- Anholt, R.R.H., O'Grady, P., Wolfner, M.F., and Harbison, S.T. (2020). Evolution of reproductive behavior. *Genetics* 214(1), 49-73. doi: 10.1534/genetics.119.302263.
- Arguello, J.R., Cardoso-Moreira, M., Grenier, J.K., Gottipati, S., Clark, A.G., and Benton, R. (2016). Extensive local adaptation within the chemosensory system following *Drosophila melanogaster*'s global expansion. *Nat Commun* 7, ncomms11855. doi: 10.1038/ncomms11855.
- Auer, T.O., Khallaf, M.A., Silbering, A.F., Zappia, G., Ellis, K., Alvarez-Ocana, R., et al. (2020). Olfactory receptor and circuit evolution promote host specialization. *Nature* 579(7799), 402-408. doi: 10.1038/s41586-020-2073-7.

References

B

- Ballare, C.L. (2011). Jasmonate-induced defenses: a tale of intelligence, collaborators and rascals. *Trends Plant Sci.* 16(5), 249-257. doi: 10.1016/j.tplants.2010.12.001.
- Bates, D., Mächler, M., Bolker, B., and Walker, S. (2015). Fitting linear mixed-effects models using lme4. *Journal of Statistical Software* 67(1), 1–48. doi: 10.18637/jss.v067.i01.
- Baumberger, J.P. (1919). A nutritional study of insects, with special reference to microorganisms and their substrata. *J. Exp. Zool.* 28(1), 1-81. doi: 10.1002/jez.1400280102.
- Becher, P.G., Flick, G., Rozpędowska, E., Schmidt, A., Hagman, A., Lebreton, S., et al. (2012). Yeast, not fruit volatiles mediate *Drosophila melanogaster* attraction, oviposition and development. *Funct. Ecol.* 26(4), 822-828. doi: 10.1111/j.1365-2435.2012.02006.x.
- Becher, P.G., Hagman, A., Verschut, V., Chakraborty, A., Rozpedowska, E., Lebreton, S., et al. (2018). Chemical signaling and insect attraction is a conserved trait in yeasts. *Ecol. Evol.* 8(5), 2962-2974. doi: 10.1002/ece3.3905.
- Bell, W.J., and Kipp, L.R. (1994). *Drosophila percnosoma* hardy lek sites: spatial and temporal distributions of males and the dynamics of their agonistic behavior (Diptera: Drosophilidae). *J. Kans. Entomol. Soc.* 67(3), 267–276.
- Benton, R. (2008). Chemical sensing in *Drosophila*. *Curr Opin Neurobiol* 18(4), 357-363. doi: 10.1016/j.conb.2008.08.012.
- Berlocher, S.H., and Feder, J.L. (2002). Sympatric speciation in phytophagous insects: moving beyond controversy? *Annu Rev Entomol* 47, 773-815. doi: 10.1146/annurev.ento.47.091201.145312.
- Bernays, E.A. (2001). Neural limitations in phytophagous insects: implications for diet breadth and evolution of host affiliation. *Annu Rev Entomol* 46(46), 703-727. doi: 10.1146/annurev.ento.46.1.703.
- Besson, M., and Martin, J.R. (2005). Centrophobism/thigmotaxis, a new role for the mushroom bodies in *Drosophila*. *J Neurobiol* 62(3), 386-396. doi: 10.1002/neu.20111.

- Billeter, J.C., Atallah, J., Krupp, J.J., Millar, J.G., and Levine, J.D. (2009). Specialized cells tag sexual and species identity in *Drosophila melanogaster*. *Nature* 461(7266), 987-991. doi: 10.1038/nature08495.
- Billeter, J.C., and Wolfner, M.F. (2018). Chemical cues that guide female reproduction in *Drosophila melanogaster*. *J Chem Ecol* 44(9), 750-769. doi: 10.1007/s10886-018-0947-z.
- Brazner, J.C., and Etges, W.J. (1993). Pre-mating isolation is determined by larval rearing substrates in cactophilic *Drosophila mojavensis*. II. Effects of larval substrates on time to copulation, mate choice and mating propensity. *Evol. Ecol.* 7(6), 605-624. doi: 10.1007/bf01237824.
- Breitmeyer, C.M., and Markow, T.A. (1998). Resource availability and population size in cactophilic *Drosophila*. *Funct. Ecol.* 12(1), 14-21. doi: 10.1046/j.1365-2435.1998.00152.x.
- Bruce, T.J., Wadhams, L.J., and Woodcock, C.M. (2005). Insect host location: a volatile situation. *Trends Plant Sci.* 10(6), 269-274. doi: 10.1016/j.tplants.2005.04.003.
- Brucker, R.M., and Bordenstein, S.R. (2012). Speciation by symbiosis. *Trends Ecol Evol* 27(8), 443-451. doi: 10.1016/j.tree.2012.03.011.
- Buser, C.C., Newcomb, R.D., Gaskett, A.C., and Goddard, M.R. (2014). Niche construction initiates the evolution of mutualistic interactions. *Ecol Lett* 17(10), 1257-1264. doi: 10.1111/ele.12331.
- Bush, G.L. (1969). Sympatric host race formation and speciation in frugivorous flies of the genus *Rhagoletis* (Diptera, Tephritidae). *Evolution* 23(2), 237-251. doi: 10.1111/j.1558-5646.1969.tb03508.x.

C

- Caillaud, M.C., and Via, S. (2000). Specialized feeding behavior influences both ecological specialization and assortative mating in sympatric host races of pea aphids. *Am Nat* 156(6), 606-621. doi: 10.1086/316991.
- Capy, P., Veuille, M., Paillette, M., Jallon, J.M., Vouidibio, J., and David, J.R. (2000). Sexual isolation of genetically differentiated sympatric populations of *Drosophila melanogaster* in Brazzaville, Congo: the first step towards speciation? *Heredity (Edinb.)* 84 (Pt 4)(4), 468-475. doi: 10.1046/j.1365-2540.2000.00711.x.

References

- Carvalho, M., Schwudke, D., Sampaio, J.L., Palm, W., Riezman, I., Dey, G., et al. (2010). Survival strategies of a sterol auxotroph. *Development* 137(21), 3675-3685. doi: 10.1242/dev.044560.
- Chandler, J.A., Eisen, J.A., and Kopp, A. (2012). Yeast communities of diverse *Drosophila* species: comparison of two symbiont groups in the same hosts. *Appl Environ Microbiol* 78(20), 7327-7336. doi: 10.1128/AEM.01741-12.
- Christiaens, J.F., Franco, L.M., Cools, T.L., De Meester, L., Michiels, J., Wenseleers, T., et al. (2014). The fungal aroma gene ATF1 promotes dispersal of yeast cells through insect vectors. *Cell Rep*. 9(2), 425-432. doi: 10.1016/j.celrep.2014.09.009.
- Cloonan, K.R., Abraham, J., Angeli, S., Syed, Z., and Rodriguez-Saona, C. (2018). Advances in the chemical ecology of the spotted wing *Drosophila* (*Drosophila suzukii*) and its applications. *J Chem Ecol* 44(10), 922-939. doi: 10.1007/s10886-018-1000-y.
- Cooke, J., and Sang, J.H. (1970). Utilization of sterols by larvae of *Drosophila melanogaster*. *J Insect Physiol* 16(5), 801-812. doi: 10.1016/0022-1910(70)90214-3.
- Cooper, D.M. (1960). Food preferences of larval and adult *Drosophila*. *Evolution* 14(1), 41-55. doi: 10.1111/j.1558-5646.1960.tb03055.x.
- Coyne, J.A., Crittenden, A.P., and Mah, K. (1994). Genetics of a pheromonal difference contributing to reproductive isolation in *Drosophila*. *Science* 265(5177), 1461-1464. doi: 10.1126/science.8073292.
- Coyne, J.A., and Orr, H.A. (2004). *Speciation*. Sunderland, MA: Sinauer Associates.
- Craig, T.P., Horner, J.D., and Itami, J.K. (1997). Hybridization studies on the host races of *Eurosta Solidaginis*: implications for sympatric speciation. *Evolution* 51(5), 1552-1560. doi: 10.1111/j.1558-5646.1997.tb01478.x.
- Craig, T.P., and Itami, J.K. (2008). "Evolution of preference and performance relationships," in *Specialization, speciation, and radiation: the evolutionary biology of herbivorous insects*. University of California Press), 20-28.
- Craig, T.P., Itami, J.K., Abrahamson, W.G., and Horner, J.D. (1993). Behavioral evidence for host-race formation in *Eurosta Solidaginis*. *Evolution* 47(6), 1696-1710. doi: 10.1111/j.1558-5646.1993.tb01262.x.
- Crowley-Gall, A., Date, P., Han, C., Rhodes, N., Andolfatto, P., Layne, J.E., et al. (2016). Population differences in olfaction accompany host shift in *Drosophila mojavensis*. *Proc. Biol. Sci.* 283(1837), 20161562. doi: 10.1098/rspb.2016.1562.

D

- Dambroski, H.R., Linn, C., Jr., Berlocher, S.H., Forbes, A.A., Roelofs, W., and Feder, J.L. (2005). The genetic basis for fruit odor discrimination in *Rhagoletis* flies and its significance for sympatric host shifts. *Evolution* 59(9), 1953-1964. doi: 10.1111/j.0014-3820.2005.tb01065.x.
- Das, S., Trona, F., Khallaf, M.A., Schuh, E., Knaden, M., Hansson, B.S., et al. (2017). Electrical synapses mediate synergism between pheromone and food odors in *Drosophila melanogaster*. *Proc Natl Acad Sci U S A* 114(46), E9962-E9971. doi: 10.1073/pnas.1712706114.
- Date, P., Dweck, H.K., Stensmyr, M.C., Shann, J., Hansson, B.S., and Rollmann, S.M. (2013). Divergence in olfactory host plant preference in *D. mojavensis* in response to cactus host use. *PLoS One* 8(7), e70027. doi: 10.1371/journal.pone.0070027.
- Davies, N.B., Krebs, J.R., and West, S.A. (2012). *An introduction to behavioural ecology*. John Wiley & Sons.
- De Moraes, C.M., Mescher, M.C., and Tumlinson, J.H. (2001). Caterpillar-induced nocturnal plant volatiles repel conspecific females. *Nature* 410(6828), 577-580. doi: 10.1038/35069058.
- Dekker, T., Iba, I., Siju, K.P., Stensmyr, M.C., and Hansson, B.S. (2006). Olfactory shifts parallel superspecialism for toxic fruit in *Drosophila melanogaster* sibling, *D. sechellia*. *Curr Biol* 16(1), 101-109. doi: 10.1016/j.cub.2005.11.075.
- Del Campo, M.L., Via, S., and Caillaud, M.C. (2003). Recognition of host-specific chemical stimulants in two sympatric host races of the pea aphid *Acyrtosiphon pisum*. *Ecol. Entomol.* 28(4), 405-412. doi: 10.1046/j.1365-2311.2003.00524.x.
- Dicke, M. (2000). Chemical ecology of host-plant selection by herbivorous arthropods: a multitrophic perspective. *Biochem Syst Ecol* 28(7), 601-617. doi: 10.1016/s0305-1978(99)00106-4.
- Dres, M., and Mallet, J. (2002). Host races in plant-feeding insects and their importance in sympatric speciation. *Philos Trans R Soc Lond B Biol Sci* 357(1420), 471-492. doi: 10.1098/rstb.2002.1059.
- Dumenil, C., Woud, D., Pinto, F., Alkema, J.T., Jansen, I., Van Der Geest, A.M., et al. (2016). Pheromonal cues deposited by mated females convey social information about egg-laying sites in *Drosophila melanogaster*. *J Chem Ecol* 42(3), 259-269. doi: 10.1007/s10886-016-0681-3.

References

E

- Edgecomb, R.S., Harth, C.E., and Schneiderman, A.M. (1994). Regulation of feeding behavior in adult *Drosophila melanogaster* varies with feeding regime and nutritional state. *J Exp Biol* 197(1), 215-235. doi: 10.1242/jeb.197.1.215.
- Engel, P., and Moran, N.A. (2013). The gut microbiota of insects - diversity in structure and function. *FEMS Microbiol. Rev.* 37(5), 699-735. doi: 10.1111/1574-6976.12025.
- Enjin, A., Zaharieva, E.E., Frank, D.D., Mansourian, S., Suh, G.S., Gallio, M., et al. (2016). Humidity sensing in *Drosophila*. *Curr Biol* 26(10), 1352-1358. doi: 10.1016/j.cub.2016.03.049.
- Etges, W.J. (1992). Premating isolation is determined by larval substrates in Cactophilic *Drosophila mojavensis*. *Evolution* 46(6), 1945-1950.
- Etges, W.J., Veenstra, C.L., and Jackson, L.L. (2006). Premating isolation is determined by larval rearing substrates in cactophilic *Drosophila mojavensis*. VII. Effects of larval dietary fatty acids on adult epicuticular hydrocarbons. *J Chem Ecol* 32(12), 2629-2646. doi: 10.1007/s10886-006-9187-8.

F

- Feder, J.L. (1998). The apple maggot fly, *Rhagoletis pomonella*. *Endless forms: species and speciation*. Oxford Univ. Press, New York, 130-144.
- Feder, J.L., and Filchak, K.E. (1999). It's about time: the evidence for host plant-mediated selection in the apple maggot fly, *Rhagoletis pomonella*, and its implications for fitness trade-offs in phytophagous insects. *Entomol. Exp. Appl.* (91), 211-225.
- Feder, J.L., Hunt, T.A., and Bush, L. (1993). The effects of climate, host plant phenology and host fidelity on the genetics of apple and hawthorn infesting races of *Rhagoletis pomonella*. *Entomol. Exp. Appl.* 69(2), 117-135. doi: 10.1111/j.1570-7458.1993.tb01735.x.
- Feder, J.L., Opp, S.B., Wlazlo, B., Reynolds, K., Go, W., and Spisak, S. (1994). Host fidelity is an effective premating barrier between sympatric races of the apple maggot fly. *Proc Natl Acad Sci U S A* 91(17), 7990-7994. doi: 10.1073/pnas.91.17.7990.

- Fedina, T.Y., Kuo, T.H., Dreisewerd, K., Dierick, H.A., Yew, J.Y., and Pletcher, S.D. (2012). Dietary effects on cuticular hydrocarbons and sexual attractiveness in *Drosophila*. *PLoS One* 7(12), e49799. doi: 10.1371/journal.pone.0049799.
- Fei, M., Gols, R., and Harvey, J.A. (2014). Seasonal phenology of interactions involving short-lived annual plants, a multivoltine herbivore and its endoparasitoid wasp. *J Anim Ecol* 83(1), 234-244. doi: 10.1111/1365-2656.12122.
- Filchak, K.E., Roethele, J.B., and Feder, J.L. (2000). Natural selection and sympatric divergence in the apple maggot *Rhagoletis pomonella*. *Nature* 407(6805), 739-742. doi: 10.1038/35037578.
- Fischer, C.N., Trautman, E.P., Crawford, J.M., Stabb, E.V., Handelsman, J., and Broderick, N.A.J.E. (2017). Metabolite exchange between microbiome members produces compounds that influence *Drosophila* behavior. 6, e18855.
- Fogleman, J.C., and Danielson, P.B. (2001). Chemical interactions in the cactus-microorganism-*Drosophila* model system of the Sonoran Desert. *Am. Zool.* 41(4), 877-889. doi: 10.1093/icb/41.4.877.
- Fox, J., and Weisberg, S. (2019). *An {R} companion to applied regression*.
- Fry, J.D., Heinsohn, S.L., and Mackay, T.F.C. (1996). The contribution of new mutations to genotype-environment interaction for fitness in *Drosophila Melanogaster*. *Evolution* 50(6), 2316-2327. doi: 10.1111/j.1558-5646.1996.tb03619.x.
- Fujii, S., Krishnan, P., Hardin, P., and Amrein, H. (2007). Nocturnal male sex drive in *Drosophila*. *Curr Biol* 17(3), 244-251. doi: 10.1016/j.cub.2006.11.049.
- Funk, D.J., Filchak, K.E., and Feder, J.L. (2002). Herbivorous insects: model systems for the comparative study of speciation ecology. *Genetica* 116(2-3), 251-267. doi: 10.1023/a:1021236510453.
- Futuyma, D.J., and Moreno, G. (1988). The evolution of ecological specialization. *Annu. Rev. Ecol. Syst.* 19, 207-233.

G

- Gadenne, C., Barrozo, R.B., and Anton, S. (2016). Plasticity in insect olfaction: to smell or not to smell? *Annu Rev Entomol* 61, 317-333. doi: 10.1146/annurev-ento-010715-023523.

References

- Gleason, J.M., Jallon, J.M., Rouault, J.D., and Ritchie, M.G. (2005). Quantitative trait loci for cuticular hydrocarbons associated with sexual isolation between *Drosophila simulans* and *D. sechellia*. *Genetics* 171(4), 1789-1798. doi: 10.1534/genetics.104.037937.
- Gleason, J.M., James, R.A., Wicker-Thomas, C., and Ritchie, M.G. (2009). Identification of quantitative trait loci function through analysis of multiple cuticular hydrocarbons differing between *Drosophila simulans* and *Drosophila sechellia* females. *Heredity (Edinb.)* 103(5), 416-424. doi: 10.1038/hdy.2009.79.
- Gloss, A.D., Groen, S.C., and Whiteman, N.K. (2016). A genomic perspective on the generation and maintenance of genetic diversity in herbivorous insects. *Annu Rev Ecol Syst* 47, 165-187. doi: 10.1146/annurev-ecolsys-121415-032220.
- Gloss, A.D., Nelson Dittrich, A.C., Lapoint, R.T., Goldman-Huertas, B., Verster, K.I., Pelaez, J.L., et al. (2019). Evolution of herbivory remodels a *Drosophila* genome. *bioRxiv*, 767160. doi: 10.1101/767160.
- Goldman-Huertas, B., Mitchell, R.F., Lapoint, R.T., Faucher, C.P., Hildebrand, J.G., and Whiteman, N.K. (2015). Evolution of herbivory in Drosophilidae linked to loss of behaviors, antennal responses, odorant receptors, and ancestral diet. *Proc Natl Acad Sci U S A* 112(10), 3026-3031. doi: 10.1073/pnas.1424656112.
- Goodman, J.M., Scott, C.W., Donahue, P.N., and Atherton, J.P. (1984). Alcohol oxidase assembles post-translationally into the peroxisome of *Candida boidinii*. *The Journal of biological chemistry* 259(13).
- Gorter, J.A., and Billeter, J.C. (2017). A method to test the effect of environmental cues on mating behavior in *Drosophila melanogaster*. *J Vis Exp* (125). doi: 10.3791/55690.
- Gorter, J.A., Jagadeesh, S., Gahr, C., Boonekamp, J.J., Levine, J.D., and Billeter, J.C. (2016). The nutritional and hedonic value of food modulate sexual receptivity in *Drosophila melanogaster* females. *Sci. Rep.* 6, 19441. doi: 10.1038/srep19441.
- Gould, A.L., Zhang, V., Lamberti, L., Jones, E.W., Obadia, B., Korasidis, N., et al. (2018). Microbiome interactions shape host fitness. *Proc Natl Acad Sci U S A* 115(51), E11951-E11960. doi: 10.1073/pnas.1809349115.
- Grangeteau, C., Yahou, F., Everaerts, C., Dupont, S., Farine, J.P., Beney, L., et al. (2018). Yeast quality in juvenile diet affects *Drosophila melanogaster* adult life traits. *Sci. Rep.* 8(1), 13070. doi: 10.1038/s41598-018-31561-9.

- Grant, P.R. (2017). *Ecology and evolution of Darwin's finches (Princeton Science Library Edition)*. Princeton, NJ: Princeton University Press.
- Grenier, J.K., Arguello, J.R., Moreira, M.C., Gottipati, S., Mohammed, J., Hackett, S.R., et al. (2015). Global diversity lines - a five-continent reference panel of sequenced *Drosophila melanogaster* strains. *G3 (Bethesda)* 5(4), 593-603. doi: 10.1534/g3.114.015883.
- Grosjean, Y., Rytz, R., Farine, J.P., Abuin, L., Cortot, J., Jefferis, G.S., et al. (2011). An olfactory receptor for food-derived odours promotes male courtship in *Drosophila*. *Nature* 478(7368), 236-240. doi: 10.1038/nature10428.
- Günther, C.S., and Goddard, M.R. (2019). Do yeasts and *Drosophila* interact just by chance? *Fungal Ecol.* 38, 37-43. doi: 10.1016/j.funeco.2018.04.005.

H

- Haerty, W., Gibert, P., Capy, P., Moreteau, B., and David, J.R. (2003). Microspatial structure of *Drosophila melanogaster* populations in Brazzaville: evidence of natural selection acting on morphometrical traits. *Heredity (Edinb.)* 91(5), 440-447. doi: 10.1038/sj.hdy.6800305.
- Hansson, B.S., and Stensmyr, M.C. (2011). Evolution of insect olfaction. *Neuron* 72(5), 698-711. doi: 10.1016/j.neuron.2011.11.003.
- Hardy, N.B., Kaczvinsky, C., Bird, G., and Normark, B.B. (2020). What we don't know about diet-breadth evolution in herbivorous insects. *Annu. Rev. Ecol. Evol. Syst.* 51(1), 103-122. doi: 10.1146/annurev-ecolsys-011720-023322.
- Heed, W.B. (1978). "Ecology and genetics of Sonoran desert *Drosophila*," in *Ecological genetics: The interface*. Springer, 109-126.
- Hendrichs, J., Katsoyannos, B.I., Papaj, D.R., and Prokopy, R.J. (1991). Sex differences in movement between natural feeding and mating sites and tradeoffs between food consumption, mating success and predator evasion in Mediterranean fruit flies (Diptera: Tephritidae). *Oecologia* 86(2), 223-231. doi: 10.1007/BF00317534.
- Heys, C., Lizé, A., Colinet, H., Price, T.A.R., Prescott, M., Ingleby, F., et al. (2018). Evidence that the microbiota counteracts male outbreeding strategy by inhibiting sexual signaling in females. *Frontiers in Ecology and Evolution* 6, 365. doi: 10.3389/fevo.2018.00029.

References

- Hoang, D., Kopp, A., and Chandler, J.A. (2015). Interactions between *Drosophila* and its natural yeast symbionts-Is *Saccharomyces cerevisiae* a good model for studying the fly-yeast relationship? *PeerJ* 3, e1116. doi: 10.7717/peerj.1116.
- Holzinger, F., Frick, C., and Wink, M. (1992). Molecular basis for the insensitivity of the Monarch (*Danaus plexippus*) to cardiac glycosides. *FEBS Lett.* 314(3), 477-480. doi: 10.1016/0014-5793(92)81530-y.
- Hood, G.R., Powell, T.H.Q., Doellman, M.M., Sim, S.B., Glover, M., Yee, W.L., et al. (2020). Rapid and repeatable host plant shifts drive reproductive isolation following a recent human-mediated introduction of the apple maggot fly, *Rhagoletis pomonella*. *Evolution* 74(1), 156-168. doi: 10.1111/evo.13882.
- Hungate, E.A., Earley, E.J., Boussy, I.A., Turissini, D.A., Ting, C.T., Moran, J.R., et al. (2013). A locus in *Drosophila sechellia* affecting tolerance of a host plant toxin. *Genetics* 195(3), 1063-1075. doi: 10.1534/genetics.113.154773.
- Hussain, A., Ucpunar, H.K., Zhang, M., Loschek, L.F., and Grunwald Kadow, I.C. (2016). Neuropeptides modulate female chemosensory processing upon mating in *Drosophila*. *PLoS Biol* 14(5), e1002455. doi: 10.1371/journal.pbio.1002455.

J

- Jaenike, J. (1978). On optimal oviposition behavior in phytophagous insects. *Theor Popul Biol* 14(3), 350-356. doi: 10.1016/0040-5809(78)90012-6.
- Jaenike, J. (1990). Host specialization in phytophagous insects. *Annu. Rev. Ecol. Syst.* 21(1), 243-273. doi: 10.1146/annurev.ecolsys.21.1.243.
- Janz, N. (2011). Ehrlich and raven revisited: mechanisms underlying codiversification of plants and enemies. *Annu. Rev. Ecol. Evol. Syst.* 42(1), 71-89. doi: 10.1146/annurev-ecolsys-102710-145024.
- Jefferis, G.S., Potter, C.J., Chan, A.M., Marin, E.C., Rohlfsing, T., Maurer, C.R., Jr., et al. (2007). Comprehensive maps of *Drosophila* higher olfactory centers: spatially segregated fruit and pheromone representation. *Cell* 128(6), 1187-1203. doi: 10.1016/j.cell.2007.01.040.
- Jiang, L., Zhan, Y., and Zhu, Y. (2018). Combining quantitative food-intake assays and forcibly activating neurons to study appetite in *Drosophila*. *J Vis Exp* (134), e56900. doi: 10.3791/56900.

- Johnson, R.M., Harpur, B.A., Dogantzis, K.A., Zayed, A., and Berenbaum, M.R. (2018). Genomic footprint of evolution of eusociality in bees: floral food use and CYPome “blooms”. *Insectes Soc.* 65(3), 445-454. doi: 10.1007/s00040-018-0631-x.
- Jones, C.D. (1998). The genetic basis of *Drosophila sechellia*'s resistance to a host plant toxin. *Genetics* 149(4), 1899-1908. doi: 10.1093/genetics/149.4.1899.
- Joseph, R.M., and Carlson, J.R. (2015). *Drosophila* chemoreceptors: a molecular interface between the chemical world and the brain. *Trends Genet* 31(12), 683-695. doi: 10.1016/j.tig.2015.09.005.
- Joseph, R.M., Devineni, A.V., King, I.F., and Heberlein, U. (2009). Oviposition preference for and positional avoidance of acetic acid provide a model for competing behavioral drives in *Drosophila*. *Proc Natl Acad Sci U S A* 106(27), 11352-11357. doi: 10.1073/pnas.0901419106.
- Jousselin, E., Cruaud, A., Genson, G., Chevenet, F., Footitt, R.G., and Coeur d'acier, A. (2013). Is ecological speciation a major trend in aphids? Insights from a molecular phylogeny of the conifer-feeding genus *Cinara*. *Front Zool* 10(1), 56. doi: 10.1186/1742-9994-10-56.
- K**
- Kakioka, R., Sutra, N., Kobayashi, H., Ansai, S., Masengi, K.W.A., Nagano, A.J., et al. (2021). Resource partitioning is not coupled with assortative mating in sympatrically divergent ricefish in a Wallacean ancient lake. *J Evol Biol* 34(7), 1133-1143. doi: 10.1111/jeb.13874.
- Kambach, S., Kuhn, I., Castagneyrol, B., and Bruelheide, H. (2016). The impact of tree diversity on different aspects of insect herbivory along a global temperature gradient - a meta-analysis. *PLoS One* 11(11), e0165815. doi: 10.1371/journal.pone.0165815.
- Kang, L., Rashkovetsky, E., Michalak, K., Garner, H.R., Mahaney, J.E., Rzigalinski, B.A., et al. (2019). Genomic divergence and adaptive convergence in *Drosophila simulans* from Evolution Canyon, Israel. *Proc Natl Acad Sci U S A* 116(24), 11839-11844. doi: 10.1073/pnas.1720938116.
- Karageorgi, M., Bracker, L.B., Lebreton, S., Minervino, C., Cavey, M., Siju, K.P., et al. (2017). Evolution of multiple sensory systems drives novel egg-laying behavior in the fruit pest *Drosophila suzukii*. *Curr Biol* 27(6), 847-853. doi: 10.1016/j.cub.2017.01.055.

References

- Karageorgi, M., Groen, S.C., Sumbul, F., Pelaez, J.N., Verster, K.I., Aguilar, J.M., et al. (2019). Genome editing retraces the evolution of toxin resistance in the monarch butterfly. *Nature* 574(7778), 409-412. doi: 10.1038/s41586-019-1610-8.
- Kaspi, R., and Yuval, B. (1999). Mediterranean fruit fly leks: factors affecting male location. *Funct. Ecol.* 13, 539–545.
- Keesey, I.W., Knaden, M., and Hansson, B.S. (2015). Olfactory specialization in *Drosophila suzukii* supports an ecological shift in host preference from rotten to fresh fruit. *J Chem Ecol* 41(2), 121-128. doi: 10.1007/s10886-015-0544-3.
- Kessler, A., and Baldwin, I.T. (2001). Defensive function of herbivore-induced plant volatile emissions in nature. *Science* 291(5511), 2141-2144. doi: 10.1126/science.291.5511.2141.
- Khallaf, M.A., Auer, T.O., Grabe, V., Depetris-Chauvin, A., Ammagarahalli, B., Zhang, D.D., et al. (2020). Mate discrimination among subspecies through a conserved olfactory pathway. *Sci Adv* 6(25), eaba5279. doi: 10.1126/sciadv.aba5279.
- Kirkpatrick, M., and Ravigne, V. (2002). Speciation by natural and sexual selection: models and experiments. *Am Nat* 159 Suppl 3, S22-35. doi: 10.1086/338370.
- Ko, K.I., Root, C.M., Lindsay, S.A., Zaninovich, O.A., Shepherd, A.K., Wasserman, S.A., et al. (2015). Starvation promotes concerted modulation of appetitive olfactory behavior via parallel neuromodulatory circuits. *Elife* 4. doi: 10.7554/eLife.08298.
- Koerte, S., Keesey, I.W., Easson, M.L.A.E., Gershenson, J., Hansson, B.S., and Knaden, M. (2020). Variable dependency on associated yeast communities influences host range in *Drosophila* species. *Oikos* 129(7), 964-982. doi: 10.1111/oik.07180.
- Kohlmeier, P., Zhang, Y., Gorter, J.A., Su, C.-Y., Billeter, J.-C.J.N.e., and evolution (2021). Mating increases *Drosophila melanogaster* females' choosiness by reducing olfactory sensitivity to a male pheromone. 5(8), 1165-1173.
- Kromann, S.H., Saveer, A.M., Binyameen, M., Bengtsson, M., Birgersson, G., Hansson, B.S., et al. (2015). Concurrent modulation of neuronal and behavioural olfactory responses to sex and host plant cues in a male moth. *Proc. Biol. Sci.* 282(1799), 20141884. doi: 10.1098/rspb.2014.1884.
- Kubli, E., and Bopp, D. (2012). Sexual behavior: how Sex Peptide flips the postmating switch of female flies. *Curr Biol* 22(13), R520-522. doi: 10.1016/j.cub.2012.04.058.

- Kuo, T.H., Yew, J.Y., Fedina, T.Y., Dreisewerd, K., Dierick, H.A., and Pletcher, S.D. (2012). Aging modulates cuticular hydrocarbons and sexual attractiveness in *Drosophila melanogaster*. *J Exp Biol* 215(Pt 5), 814-821. doi: 10.1242/jeb.064980.
- Kurtzman, C.P., Mateo, R.Q., Kolecka, A., Theelen, B., Robert, V., and Boekhout, T. (2015). Advances in yeast systematics and phylogeny and their use as predictors of biotechnologically important metabolic pathways. *FEMS Yeast Res.* 15(6). doi: 10.1093/femsyr/fov050.
- Kvarnemo, C., and Simmons, L.W. (2013). Polyandry as a mediator of sexual selection before and after mating. *Philos Trans R Soc Lond B Biol Sci* 368(1613), 20120042. doi: 10.1098/rstb.2012.0042.
- Kwan, L., and Rundle, H.D. (2010). Adaptation to desiccation fails to generate pre- and postmating isolation in replicate *Drosophila melanogaster* laboratory populations. *Evolution* 64(3), 710-723. doi: 10.1111/j.1558-5646.2009.00864.x.
- L**
- Lam, S.S., and Howell, K.S. (2015). *Drosophila*-associated yeast species in vineyard ecosystems. *FEMS Microbiol. Lett.* 362(20). doi: 10.1093/femsle/fnv170.
- Lande, R. (1979). Quantitative genetic analysis of multivariate evolution, applied to brain:body size allometry. *Evolution* 33(1Part2), 402-416. doi: 10.1111/j.1558-5646.1979.tb04694.x.
- Landolt, P.J., and Phillips, T.W. (1997). Host plant influences on sex pheromone behavior of phytophagous insects. *Annu Rev Entomol* 42, 371-391. doi: 10.1146/annurev.ento.42.1.371.
- Landry, C.R., and Aubin-Horth, N. (eds.). (2014). *Ecological genomics*. Dordrecht: Springer.
- Lanno, S.M., Gregory, S.M., Shimshak, S.J., Alverson, M.K., Chiu, K., Feil, A.L., et al. (2017). Transcriptomic analysis of octanoic acid response in *Drosophila sechellia* using RNA-sequencing. *G3 (Bethesda)* 7(12), 3867-3873. doi: 10.1534/g3.117.300297.
- Larter, N.K., Sun, J.S., and Carlson, J.R. (2016). Organization and function of *Drosophila* odorant binding proteins. *Elife* 5, e20242. doi: 10.7554/eLife.20242.

References

- Lavista-Llanos, S., Svatos, A., Kai, M., Riemensperger, T., Birman, S., Stensmyr, M.C., et al. (2014). Dopamine drives *Drosophila sechellia* adaptation to its toxic host. *Elife* 3. doi: 10.7554/eLife.03785.
- Le Goff, G., and Hilliou, F. (2017). Resistance evolution in *Drosophila*: the case of CYP6G1. *Pest Manag Sci* 73(3), 493-499. doi: 10.1002/ps.4470.
- Leal, W.S. (2013). Odorant reception in insects: roles of receptors, binding proteins, and degrading enzymes. *Annu Rev Entomol* 58, 373-391. doi: 10.1146/annurev-ento-120811-153635.
- Lebreton, S., Carlsson, M.A., and Witzgall, P. (2017). Insulin Signaling in the Peripheral and Central Nervous System Regulates Female Sexual Receptivity during Starvation in *Drosophila*. *Front. Physiol.* 8, 685. doi: 10.3389/fphys.2017.00685.
- Lebreton, S., Trona, F., Borrero-Echeverry, F., Bilz, F., Grabe, V., Becher, P.G., et al. (2015). Feeding regulates sex pheromone attraction and courtship in *Drosophila* females. *Sci. Rep.* 5(1), 13132. doi: 10.1038/srep13132.
- Lee, J.C., Bruck, D.J., Dreves, A.J., Ioriatti, C., Vogt, H., and Baufeld, P. (2011). In Focus: Spotted wing drosophila, *Drosophila suzukii*, across perspectives. *Pest Manag Sci* 67(11), 1349-1351. doi: 10.1002/ps.2271.
- Legal, L., Chappe, B., and Jallon, J.M. (1994). Molecular basis of *Morinda citrifolia* (L.): toxicity on *Drosophila*. *J Chem Ecol* 20(8), 1931-1943. doi: 10.1007/BF02066234.
- Legal, L., David, J.R., and Jallon, J.M. (1992). Toxicity and attraction effects produced by *Morinda citrifolia* fruits on the *Drosophila melanogaster* complex of species. *Chemoecology* 3(3-4), 125-129. doi: 10.1007/bf01370140.
- Liang, L., and Luo, L. (2010). The olfactory circuit of the fruit fly *Drosophila melanogaster*. *Sci China Life Sci* 53(4), 472-484. doi: 10.1007/s11427-010-0099-z.
- Lightle, D., Ambrosino, M., and Lee, J.C. (2010). Sugar in moderation: sugar diets affect short-term parasitoid behaviour. *Physiol. Entomol.* 35(2), 179-185. doi: 10.1111/j.1365-3032.2009.00718.x.
- Lin, C.-c., Prokop-prigge, K.A., Preti, G., and Potter, C.J. (2015). Food odors trigger *Drosophila* males to deposit a pheromone that guides aggregation and female oviposition decisions. *eLife*.

- Linz, J., Baschwitz, A., Strutz, A., Dweck, H.K., Sachse, S., Hansson, B.S., et al. (2013). Host plant-driven sensory specialization in *Drosophila erecta*. *Proc. Biol. Sci.* 280(1760), 20130626. doi: 10.1098/rspb.2013.0626.
- Liu, H., and Kubli, E. (2003). Sex-peptide is the molecular basis of the sperm effect in *Drosophila melanogaster*. *Proc Natl Acad Sci U S A* 100(17), 9929-9933. doi: 10.1073/pnas.1631700100.
- Luo, S., Michaud, J.P., Li, J., Liu, X., and Zhang, Q. (2013). Odor learning in *Microplitis mediator* (Hymenoptera: Braconidae) is mediated by sugar type and physiological state. *Biol. Control* 65(2), 207-211. doi: 10.1016/j.biocontrol.2013.02.010.

M

- MacWilliam, D., Kowalewski, J., Kumar, A., Pontrello, C., and Ray, A. (2018). Signaling mode of the broad-spectrum conserved CO₂ Receptor is one of the important determinants of odor valence in *Drosophila*. *Neuron* 97(5), 1153-1167 e1154. doi: 10.1016/j.neuron.2018.01.028.
- Mansourian, S., Enjin, A., Jirle, E.V., Ramesh, V., Rehermann, G., Becher, P.G., et al. (2018). Wild African *Drosophila melanogaster* Are Seasonal Specialists on Marula Fruit. *Curr Biol* 28(24), 3960-3968 e3963. doi: 10.1016/j.cub.2018.10.033.
- Marcillac, F., and Ferveur, J.F. (2004). A set of female pheromones affects reproduction before, during and after mating in *Drosophila*. *J Exp Biol* 207(Pt 22), 3927-3933. doi: 10.1242/jeb.01236.
- Marie Curie, S.N., Butlin, R., Debelle, A., Kerth, C., Snook, R.R., Beukeboom, L.W., et al. (2012). What do we need to know about speciation? *Trends Ecol Evol* 27(1), 27-39. doi: 10.1016/j.tree.2011.09.002.
- Markow, T.A. (1988). Reproductive behavior of *Drosophila melanogaster* and *D. nigrospiracula* in the field and in the laboratory. *J Comp Psychol* 102(2), 169-173. doi: 10.1037/0735-7036.102.2.169.
- Markow, T.A. (2015). The secret lives of *Drosophila* flies. *Elife* 4. doi: 10.7554/eLife.06793.
- Markow, T.A., and O'Grady, P. (2008). Reproductive ecology of *Drosophila*. *Funct. Ecol.* 22(5), 747-759. doi: 10.1111/j.1365-2435.2008.01457.x.

References

- Matsubayashi, K.W., Ohshima, I., and Nosil, P. (2010). Ecological speciation in phytophagous insects. *Entomol. Exp. Appl.* 134(1), 1-27. doi: 10.1111/j.1570-7458.2009.00916.x.
- Matsunaga, T., Reisenman, C.E., Goldman-Huertas, B., Brand, P., Miao, K., Suzuki, H.C., et al. (2021). Olfactory receptors tuned to volatile mustard oils in drosophilid flies. *bioRxiv*. doi: 10.1101/2019.12.27.889774.
- Matsuo, T., Sugaya, S., Yasukawa, J., Aigaki, T., and Fuyama, Y. (2007). Odorant-binding proteins OBP57d and OBP57e affect taste perception and host-plant preference in *Drosophila sechellia*. *PLoS Biol* 5(5), e118. doi: 10.1371/journal.pbio.0050118.
- Matzkin, L.M. (2012). Population transcriptomics of cactus host shifts in *Drosophila mojavensis*. *Mol Ecol* 21(10), 2428-2439. doi: 10.1111/j.1365-294X.2012.05549.x.
- Matzkin, L.M. (2014). "Ecological genomics of host shifts in *Drosophila mojavensis*," in *Ecological genomics*, eds. C.R. Landry & N. Aubin-Horth. (Dordrecht: Springer), 233-247.
- Matzkin, L.M., Markow, T.A., and Michalak, P. (2013). *Transcriptional differentiation across the four subspecies of Drosophila mojavensis*. New York: Nova Scientific Publishers.
- McBride, C.S., Baier, F., Omondi, A.B., Spitzer, S.A., Lutomiah, J., Sang, R., et al. (2014). Evolution of mosquito preference for humans linked to an odorant receptor. *Nature* 515(7526), 222-227. doi: 10.1038/nature13964.
- Mercier, D., Tsuchimoto, Y., Ohta, K., and Kazama, H. (2018). Olfactory Landmark-Based Communication in Interacting *Drosophila*. *Curr Biol* 28(16), 2624-2631 e2625. doi: 10.1016/j.cub.2018.06.005.
- Missbach, C., Dweck, H.K., Vogel, H., Vilcinskas, A., Stensmyr, M.C., Hansson, B.S., et al. (2014). Evolution of insect olfactory receptors. *Elife* 3, e02115. doi: 10.7554/eLife.02115.
- Mithofer, A., and Boland, W. (2012). Plant defense against herbivores: chemical aspects. *Annu. Rev. Plant Biol.* 63, 431-450. doi: 10.1146/annurev-arplant-042110-103854.
- Mohammad, F., Aryal, S., Ho, J., Stewart, J.C., Norman, N.A., Tan, T.L., et al. (2016). Ancient anxiety pathways influence *Drosophila* defense behaviors. *Curr Biol* 26(7), 981-986. doi: 10.1016/j.cub.2016.02.031.

- Montgomery, S.L. (1975). Comparative breeding site ecology and the adaptive radiation of picture-winged *Drosophila* (Diptera: Drosophilidae) in Hawaii.
- Moreau, J., Desouhant, E., Louâpre, P., Goubault, M., Rajon, E., Jarrige, A., et al. (2017). "How host plant and fluctuating environments affect insect reproductive strategies?," in *Insect-Plant Interactions in a Crop Protection Perspective*, eds. N. Sauvion, D. Thiéry & P.-A. Calatayud. Academic Press), 259-287.
- Morrow, E.H., Stewart, A.D., and Rice, W.R. (2005). Patterns of sperm precedence are not affected by female mating history in *Drosophila Melanogaster*. *Evolution* 59(12), 2608-2615. doi: 10.1111/j.0014-3820.2005.tb00973.x.
- Murgier, J., Everaerts, C., Farine, J.P., and Ferveur, J.F. (2019). Live yeast in juvenile diet induces species-specific effects on *Drosophila* adult behaviour and fitness. *Sci. Rep.* 9(1), 8873. doi: 10.1038/s41598-019-45140-z.
- N**
- Nevo, E., Rashkovetsky, E., Pavlicek, T., and Korol, A. (1998). A complex adaptive syndrome in *Drosophila* caused by microclimatic contrasts. *Heredity (Edinb.)* 80 (Pt 1)(1), 9-16. doi: 10.1046/j.1365-2540.1998.00274.x.
- Newby, B.D., and Etges, W.J. (1998). Host preference among populations of *Drosophila mojavensis* (Diptera: Drosophilidae) that use different host cacti. *J. Insect Behav.* 11(5), 691-712. doi: 10.1023/a:1022398809881.
- Nosil, P. (2012). *Ecological speciation*. Oxford University Press.
- Nyman, T., Linder, H.P., Pena, C., Malm, T., and Wahlberg, N. (2012). Climate-driven diversity dynamics in plants and plant-feeding insects. *Ecol Lett* 15(8), 889-898. doi: 10.1111/j.1461-0248.2012.01782.x.
- Nyman, T., Vikberg, V., Smith, D.R., and Boeve, J.L. (2010). How common is ecological speciation in plant-feeding insects? A 'Higher' Nematinae perspective. *BMC Evol Biol* 10(1), 266. doi: 10.1186/1471-2148-10-266.

O

References

- Oakeshott, J.G., Vacek, D.C., and Anderson, P.R. (1989). Effects of microbial floras on the distributions of five domestic *Drosophila* species across fruit resources. *Oecologia* 78(4), 533-541. doi: 10.1007/BF00378745.
- Otte, T., Hilker, M., and Geiselhardt, S. (2016). Phenotypic plasticity of mate recognition systems prevents sexual interference between two sympatric leaf beetle species. *Evolution* 70(8), 1819-1828. doi: 10.1111/evo.12976.
- Otte, T., Hilker, M., and Geiselhardt, S. (2018). Phenotypic plasticity of cuticular hydrocarbon profiles in insects. *J Chem Ecol* 44(3), 235-247. doi: 10.1007/s10886-018-0934-4.

P

- Pegoraro, M., Flavell, L.M.M., Menegazzi, P., Colombi, P., Dao, P., Helfrich-Forster, C., et al. (2020). The genetic basis of diurnal preference in *Drosophila melanogaster*. *BMC Genomics* 21(1), 596. doi: 10.1186/s12864-020-07020-z.
- Pelosi, P., Iovinella, I., Felicioli, A., and Dani, F.R. (2014). Soluble proteins of chemical communication: an overview across arthropods. *Front. Physiol.* 5, 320. doi: 10.3389/fphys.2014.00320.
- Piper, M.D., Blanc, E., Leitao-Goncalves, R., Yang, M., He, X., Linford, N.J., et al. (2014). A holidic medium for *Drosophila melanogaster*. *Nat. Methods* 11(1), 100-105. doi: 10.1038/nmeth.2731.
- Prieto-Godino, L.L., Rytz, R., Cruchet, S., Bargeton, B., Abuin, L., Silbering, A.F., et al. (2017). Evolution of acid-sensing olfactory circuits in Drosophilids. *Neuron* 93(3), 661-676 e666. doi: 10.1016/j.neuron.2016.12.024.

Q

- Quan, A.S., and Eisen, M.B. (2018). The ecology of the *Drosophila*-yeast mutualism in wineries. *PLoS One* 13(5), e0196440. doi: 10.1371/journal.pone.0196440.

R

- Raghu, S., and Clarke, A.R. (2003). Spatial and temporal partitioning of behaviour by adult dacines: direct evidence for methyl eugenol as a mate rendezvous cue for *Bactrocera cacuminata*. *Physiol. Entomol.* (28), 175–184.
- Ramasamy, S., Ometto, L., Crava, C.M., Revadi, S., Kaur, R., Horner, D.S., et al. (2016). The Evolution of olfactory gene families in *Drosophila* and the genomic basis of chemical-ecological adaptation in *Drosophila suzukii*. *Genome Biol. Evol.* 8(8), 2297-2311. doi: 10.1093/gbe/evw160.
- R Core Team. (2020). R: A language and environment for statistical computing. R Foundation for Statistical Computing. <https://www.R-project.org/>
- Rehermann, G., Spitaler, U., Sahle, K., Cossu, C.S., Donne, L.D., Bianchi, F., et al. (2022). Behavioral manipulation of *Drosophila suzukii* for pest control: high attraction to yeast enhances insecticide efficacy when applied on leaves. *78(3)*, 896-904.
- Revadi, S., Vitagliano, S., Rossi Stacconi, M.V., Ramasamy, S., Mansourian, S., Carlin, S., et al. (2015). Olfactory responses of *Drosophila suzukii* females to host plant volatiles. *Physiol. Entomol.* 40(1), 54-64. doi: 10.1111/phen.12088.
- Ribeiro, C., and Dickson, B.J. (2010). Sex peptide receptor and neuronal TOR/S6K signaling modulate nutrient balancing in *Drosophila*. *Curr Biol* 20(11), 1000-1005. doi: 10.1016/j.cub.2010.03.061.
- Ribeiro, I.M.A., Drews, M., Bahl, A., Machacek, C., Borst, A., and Dickson, B.J. (2018). Visual projection neurons mediating directed courtship in *Drosophila*. *Cell* 174(3), 607-621 e618. doi: 10.1016/j.cell.2018.06.020.
- Rice, W.R. (1987). Speciation via habitat specialization: the evolution of reproductive isolation as a correlated character. *Evol. Ecol.* 1(4), 301-314. doi: 10.1007/bf02071555.
- Rimal, S., and Lee, Y. (2018). The multidimensional ionotropic receptors of *Drosophila melanogaster*. *Insect Mol Biol* 27(1), 1-7. doi: 10.1111/imb.12347.
- Rio, B., Couturier, G., Lemeunier, F., and Lachaise, D. (1983). Evolution d'une spécialisation saisonnière chez *Drosophila erecta* (Dipt., Drosophilidae). 235–248.
- R'Kha, S., Capy, P., and David, J.R. (1991). Host-plant specialization in the *Drosophila melanogaster* species complex: a physiological, behavioral, and genetical analysis. *Proc Natl Acad Sci U S A* 88(5), 1835-1839. doi: 10.1073/pnas.88.5.1835.

References

- Romero-Ferrero, F., Bergomi, M.G., Hinz, R.C., Heras, F.J., and De Polavieja, G.G. (2019). Idtracker. ai: tracking all individuals in small or large collectives of unmarked animals. *Nat. Methods* 16(2), 179-182.
- Root, C.M., Ko, K.I., Jafari, A., and Wang, J.W. (2011). Presynaptic facilitation by neuropeptide signaling mediates odor-driven food search. *Cell* 145(1), 133-144. doi: 10.1016/j.cell.2011.02.008.
- Rosquete, M.R., and Kleine-Vehn, J. (2013). Halotropism: turning down the salty date. *Curr Biol* 23(20), R927-929. doi: 10.1016/j.cub.2013.08.020.
- Rothwell, E.M., and Holeski, L.M.J.E.E. (2020). Phytochemical defences and performance of specialist and generalist herbivores: A meta-analysis. 45(3), 396-405.
- Rundle, H.D., Chenoweth, S.F., and Blows, M.W. (2009). The diversification of mate preferences by natural and sexual selection. *J Evol Biol* 22(8), 1608-1615. doi: 10.1111/j.1420-9101.2009.01773.x.
- Rundle, H.D., and Dyer, K.A. (2015). Reproductive character displacement of female mate preferences for male cuticular hydrocarbons in *Drosophila subquinaria*. *Evolution* 69(10), 2625-2637. doi: 10.1111/evo.12761.
- Rundle, H.D., and Nosil, P. (2005). Ecological speciation. *Ecol. Lett.* 8(3), 336-352. doi: 10.1111/j.1461-0248.2004.00715.x.

S

- Sauvion, N., Thiéry, D., and Calatayud, P.-A. (eds.). (2017). *Advances in botanical research : insect-plant interactions in a crop protection perspective*. Academic Press.
- Saveer, A.M., Kromann, S.H., Birgersson, G., Bengtsson, M., Lindblom, T., Balkenius, A., et al. (2012). Floral to green: mating switches moth olfactory coding and preference. *Proc. Biol. Sci.* 279(1737), 2314-2322. doi: 10.1098/rspb.2011.2710.
- Scaplen, K.M., Mei, N.J., Bounds, H.A., Song, S.L., Azanchi, R., and Kaun, K.R. (2019). Automated real-time quantification of group locomotor activity in *Drosophila melanogaster*. *Sci. Rep.* 9(1), 4427. doi: 10.1038/s41598-019-40952-5.
- Scheidler, N.H., Liu, C., Hamby, K.A., Zalom, F.G., and Syed, Z. (2015). Volatile codes: correlation of olfactory signals and reception in *Drosophila*-yeast chemical communication. *Sci. Rep.* 5, 14059. doi: 10.1038/srep14059.

- Schluter, D. (2009). Evidence for ecological speciation and its alternative. *Science* 323(5915), 737-741. doi: 10.1126/science.1160006.
- Scott, K. (2018). Gustatory processing in *Drosophila melanogaster*. *Annu Rev Entomol* 63, 15-30. doi: 10.1146/annurev-ento-020117-043331.
- Scriber, J.M., and Slansky, F. (1981). The nutritional ecology of immature insects. *Annu. Rev. Entomol.* 26(1), 183-211. doi: 10.1146/annurev.en.26.010181.001151.
- Seeholzer, L.F., Seppo, M., Stern, D.L., and Ruta, V. (2018). Evolution of a central neural circuit underlies *Drosophila* mate preferences. *Nature* 559(7715), 564-569. doi: 10.1038/s41586-018-0322-9.
- Servedio, M.R., and Boughman, J.W. (2017). The role of sexual selection in local adaptation and speciation. *Annu. Rev. Ecol. Evol. Syst.* 48(1), 85-109. doi: 10.1146/annurev-ecolsys-110316-022905.
- Servedio, M.R., Van Doorn, G.S., Kopp, M., Frame, A.M., and Nosil, P. (2011). Magic traits in speciation: 'magic' but not rare? *Trends Ecol Evol* 26(8), 389-397. doi: 10.1016/j.tree.2011.04.005.
- Shahandeh, M.P., Brock, C., and Turner, T.L. (2020). Light dependent courtship behavior in *Drosophila simulans* and *D. melanogaster*. *PeerJ* 8, e9499. doi: 10.7717/peerj.9499.
- Shahandeh, M.P., Pischedda, A., and Turner, T.L. (2018). Male mate choice via cuticular hydrocarbon pheromones drives reproductive isolation between *Drosophila* species. *Evolution* 72(1), 123-135. doi: 10.1111/evo.13389.
- Shahandeh, M.P., and Turner, T.L. (2020). The complex genetic architecture of male mate choice evolution between *Drosophila* species. *Heredity (Edinb.)* 124(6), 737-750. doi: 10.1038/s41437-020-0309-9.
- Sharon, G., Segal, D., Ringo, J.M., Hefetz, A., Zilber-Rosenberg, I., and Rosenberg, E. (2010). Commensal bacteria play a role in mating preference of *Drosophila melanogaster*. *Proc Natl Acad Sci U S A* 107(46), 20051-20056. doi: 10.1073/pnas.1009906107.
- Sharon, G., Segal, D., Zilber-Rosenberg, I., and Rosenberg, E. (2011). Symbiotic bacteria are responsible for diet-induced mating preference in *Drosophila melanogaster*, providing support for the hologenome concept of evolution. *Gut Microbes* 2(3), 190-192. doi: 10.4161/gmic.2.3.16103.

References

- Shelly, T.E. (2018). Sexual selection on leks: a fruit fly primer. *J Insect Sci* 18(3). doi: 10.1093/jisesa/iey048.
- Shropshire, J.D., and Bordenstein, S.R. (2016). Speciation by symbiosis: the microbiome and behavior. *mBio* 7(2), e01785. doi: 10.1128/mBio.01785-15.
- Silbering, A.F., and Benton, R. (2010). Ionotropic and metabotropic mechanisms in chemoreception: 'chance or design'? *EMBO Rep* 11(3), 173-179. doi: 10.1038/embor.2010.8.
- Simon, J.C., and Dickinson, M.H. (2010). A new chamber for studying the behavior of *Drosophila*. *PLoS One* 5(1), e8793. doi: 10.1371/journal.pone.0008793.
- Singer, M.C., and Parmesan, C. (2010). Phenological asynchrony between herbivorous insects and their hosts: signal of climate change or pre-existing adaptive strategy? *Philos Trans R Soc Lond B Biol Sci* 365(1555), 3161-3176. doi: 10.1098/rstb.2010.0144.
- Singer, M.S. (2008). "Evolutionary Ecology of Polyphagy," in *Specialization, Speciation, and Radiation The Evolutionary Biology of Herbivorous Insects*, ed. K. Tilmon. University of California Press), 29-42.
- Smadja, C., and Butlin, R.K. (2009). On the scent of speciation: the chemosensory system and its role in premating isolation. *Heredity (Edinb.)* 102(1), 77-97. doi: 10.1038/hdy.2008.55.
- Snowberg, L.K., and Bolnick, D.I. (2012). Partitioning the effects of spatial isolation, nest habitat, and individual diet in causing assortative mating within a population of threespine stickleback. *Evolution* 66(11), 3582-3594. doi: 10.1111/j.1558-5646.2012.01701.x.
- Soibam, B., Mann, M., Liu, L., Tran, J., Lobaina, M., Kang, Y.Y., et al. (2012). Open-field arena boundary is a primary object of exploration for *Drosophila*. *Brain Behav* 2(2), 97-108. doi: 10.1002/brb3.36.
- Soto-Yeber, L., Soto-Ortiz, J., Godoy, P., and Godoy-Herrera, R. (2018). The behavior of adult *Drosophila* in the wild. *PLoS One* 13(12), e0209917. doi: 10.1371/journal.pone.0209917.
- Stamps, J., Buechner, M., Alexander, K., Davis, J., and Zuniga, N. (2005). Genotypic differences in space use and movement patterns in *Drosophila melanogaster*. *Anim. Behav.* 70(3), 609-618. doi: 10.1016/j.anbehav.2004.11.018.

- Stamps, J.A., Yang, L.H., Morales, V.M., and Boundy-Mills, K.L. (2012). *Drosophila* regulate yeast density and increase yeast community similarity in a natural substrate. *PLoS One* 7(7), e42238. doi: 10.1371/journal.pone.0042238.
- Starmer, W.T., and Aberdeen, V. (1990). "The nutritional importance of pure and mixed cultures of yeasts in the development of *Drosophila mulleri* larvae in *Opuntia* tissues and its relationship to host plant shifts," in *Ecological and evolutionary genetics of Drosophila*. Springer), 145–160.
- Starmer, W.T., Peris, F., and Fontdevila, A. (1988). The transmission of yeasts by *Drosophila buzzatii* during courtship and mating. *Anim. Behav.* 36(6), 1691-1695. doi: 10.1016/s0003-3472(88)80109-x.
- Steck, K., Walker, S.J., Itskov, P.M., Baltazar, C., Moreira, J.M., and Ribeiro, C. (2018). Internal amino acid state modulates yeast taste neurons to support protein homeostasis in *Drosophila*. *Elife* 7. doi: 10.7554/eLife.31625.
- Stefanini, I. (2018). Yeast-insect associations: It takes guts. *Yeast* 35(4), 315-330. doi: 10.1002/yea.3309.
- Stevens, M. (2013). *Sensory ecology, behaviour, and evolution*. Oxford University Press.
- T**
- Team, R.C. (2020). *R: A language and environment for statistical computing*.
- Terashima, J., and Bownes, M. (2004). Translating available food into the number of eggs laid by *Drosophila melanogaster*. *Genetics* 167(4), 1711-1719. doi: 10.1534/genetics.103.024323.
- Thornhill, R., and Alcock, J. (1983). *The evolution of insect mating systems*. s.l.: Harvard University Press.
- Tomioka, S., Aigaki, T., and Matsuo, T. (2012). Conserved cis-regulatory elements of two odorant-binding protein genes, *Obp57d* and *Obp57e*, in *Drosophila*. *Genes Genet Syst* 87(5), 323-329. doi: 10.1266/ggs.87.323.
- Tsuchida, T., Koga, R., and Fukatsu, T. (2004). Host plant specialization governed by facultative symbiont. *Science* 303(5666), 1989. doi: 10.1126/science.1094611.

References

Tsukamoto, Y., Kataoka, H., Nagasawa, H., and Nagata, S. (2014). Mating changes the female dietary preference in the two-spotted cricket, *Gryllus bimaculatus*. *Front. Physiol.* 5, 95. doi: 10.3389/fphys.2014.00095.

V

Verschut, T.A., and Hamback, P.A. (2018). A random survival forest illustrates the importance of natural enemies compared to host plant quality on leaf beetle survival rates. *BMC Ecol* 18(1), 33. doi: 10.1186/s12898-018-0187-7.

Vogt, R.G., Prestwich, G.D., and Lerner, M.R. (1991). Odorant-binding-protein subfamilies associate with distinct classes of olfactory receptor neurons in insects. *J Neurobiol* 22(1), 74-84. doi: 10.1002/neu.480220108.

Vosshall, L.B., Amrein, H., Morozov, P.S., Rzhetsky, A., and Axel, R. (1999). A spatial map of olfactory receptor expression in the *Drosophila* antenna. *Cell* 96(5), 725-736. doi: 10.1016/s0092-8674(00)80582-6.

Voudibio, J., Capy, P., Defaye, D., Pla, E., Sandrin, J., Csink, A., et al. (1989). Short-range genetic structure of *Drosophila melanogaster* populations in an Afrotropical urban area and its significance. *Proc Natl Acad Sci U S A* 86(21), 8442-8446. doi: 10.1073/pnas.86.21.8442.

W

Wagner, D.L., and Van Driesche, R.G. (2010). Threats posed to rare or endangered insects by invasions of nonnative species. *Annu Rev Entomol* 55, 547-568. doi: 10.1146/annurev-ento-112408-085516.

Walter, T., and Couzin, I.D. (2021). TRex, a fast multi-animal tracking system with markerless identification, and 2D estimation of posture and visual fields. *Elife* 10. doi: 10.7554/eLife.64000.

Wang, X., Billeter, J.C., and Maan, M.E. (2022). Lack of alignment across yeast-dependent life-history traits may limit *Drosophila melanogaster* dietary specialization. *J. Evol. Biol.*

- Wang, X., Verschut, T.A., Billeter, J.-C., and Maan, M.E. (2021). Seven questions on the chemical ecology and neurogenetics of resource-mediated speciation. *Frontiers in Ecology and Evolution* 9, 7. doi: 10.3389/fevo.2021.640486.
- Warburg, M.S., and Yuval, B. (1997). Circadian patterns of feeding and reproductive activities of Mediterranean fruit flies (Diptera: Tephritidae) on various hosts in Israel. *Ann. Entomol. Soc. Am.* 90(4), 487-495. doi: 10.1093/aesa/90.4.487.
- Webster, B., and Carde, R.T. (2017). Use of habitat odour by host-seeking insects. *Biol. Rev. Camb. Philos. Soc.* 92(2), 1241-1249. doi: 10.1111/brv.12281.
- Weddle, C.B., Steiger, S., Hamaker, C.G., Ower, G.D., Mitchell, C., Sakaluk, S.K., et al. (2013). Cuticular hydrocarbons as a basis for chemosensory self-referencing in crickets: a potentially universal mechanism facilitating polyandry in insects. *Ecol Lett* 16(3), 346-353. doi: 10.1111/ele.12046.
- Wertheim, B., Allemand, R., Vet, L.E.M., and Dicke, M. (2006). Effects of aggregation pheromone on individual behaviour and food web interactions: a field study on *Drosophila*. *Ecol. Entomol.* 31(3), 216-226. doi: 10.1111/j.1365-2311.2006.00757.x.
- Wetzel, W.C., Kharouba, H.M., Robinson, M., Holyoak, M., and Karban, R. (2016). Variability in plant nutrients reduces insect herbivore performance. *Nature* 539(7629), 425-427. doi: 10.1038/nature20140.
- Wilson, D.A., Best, A.R., and Sullivan, R.M. (2004). Plasticity in the olfactory system: lessons for the neurobiology of memory. *Neuroscientist* 10(6), 513-524. doi: 10.1177/1073858404267048.
- Wisotsky, Z., Medina, A., Freeman, E., and Dahanukar, A. (2011). Evolutionary differences in food preference rely on Gr64e, a receptor for glycerol. *Nat Neurosci* 14(12), 1534-1541. doi: 10.1038/nn.2944.
- Wood, T.K., and Keese, M.C. (1990). Host-plant-induced assortative mating in *Enchenopa* treehoppers. *Evolution* 44(3), 619-628. doi: 10.1111/j.1558-5646.1990.tb05942.x.

X

- Xu, H., and Turlings, T.C.J. (2018). Plant volatiles as mate-finding cues for insects. *Trends Plant Sci.* 23(2), 100-111. doi: 10.1016/j.tplants.2017.11.004.

References

Xu, K., Zheng, X., and Sehgal, A. (2008). Regulation of feeding and metabolism by neuronal and peripheral clocks in *Drosophila*. *Cell Metab.* 8(4), 289-300. doi: 10.1016/j.cmet.2008.09.006.

Xue, H.-J., Segraves, K.A., Wei, J., Zhang, B., Nie, R.-E., Li, W.-Z., et al. (2018). Chemically mediated sexual signals restrict hybrid speciation in a flea beetle. *Behav. Ecol.* 29(6), 1462–1471. doi: 10.1093/beheco/ary105.

Y

Yapici, N., Kim, Y.J., Ribeiro, C., and Dickson, B.J. (2008). A receptor that mediates the post-mating switch in *Drosophila* reproductive behaviour. *Nature* 451(7174), 33-37. doi: 10.1038/nature06483.

Yassin, A., Debat, V., Bastide, H., Gidaszewski, N., David, J.R., and Pool, J.E. (2016). Recurrent specialization on a toxic fruit in an island *Drosophila* population. *Proc Natl Acad Sci U S A* 113(17), 4771-4776. doi: 10.1073/pnas.1522559113.

Yukilevich, R. (2012). Asymmetrical patterns of speciation uniquely support reinforcement in *Drosophila*. *Evolution* 66(5), 1430-1446. doi: 10.1111/j.1558-5646.2011.01534.x.

Z

Zaninovich, O.A., Kim, S.M., Root, C.R., Green, D.S., Ko, K.I., and Wang, J.W. (2013). A single-fly assay for foraging behavior in *Drosophila*. *J Vis Exp* (81), e50801. doi: 10.3791/50801.

English Summary

Adaptation to different environments is a major mechanism in phenotypic evolution and species diversification. Among the different environmental factors, food is a key driver of local adaptation and speciation. Numerous cases of food-mediated speciation are seen in birds and insects, but how populations exploiting different food resources differentiate and become reproductively isolated from each other remain poorly understood. The research presented in this thesis aimed at dissecting how heterogeneous food resources can drive population differentiation and assortative mating between populations, using the model organism *Drosophila melanogaster* and its essential food resource – yeast.

The chemosensory system is the most ubiquitous sensory channel in animals to detect and respond to food resources. Chemosensory divergence may thus play a crucial role in food-mediated adaptation and speciation. In **Chapter 2**, we formulated seven key questions to explore how the chemosensory system can facilitate food-mediated ecological speciation. We used examples of insect research and integrated approaches from different scientific disciplines. We started by identifying which aspects of food resources are heterogeneous in a given environment (Question 1), as this is the starting point of divergent selection. We then discussed which aspects of food exert selection on consumers (Question 2), and explored how consumers detect (Question 3), exploit (Question 4) and adapt to these resources (Question 5) to understand how local adaptation proceeds. Finally, we discussed whether successful exploitation of new food resources is genetically inherited and/or acquired during an individual's lifetime (Question 6) and reviewed the mechanisms that reduce gene flow between populations that specialize on alternative resources (Question 7). The formulated seven questions allowed us to find some knowledge gaps. Notably, different approaches are needed for different questions. There is rarely one model system in which all seven questions have been thoroughly answered. It is often not clear what kind of heterogeneity, or which aspect of a particular resource, will generate divergence in consumers. Because of the important roles gut microbes play in resource exploitation ability and mate choice of hosts, we proposed microbes in the gut as a promising direction for future research on food-mediated ecological speciation.

The successful exploitation of a food resource is a multifactorial phenotype that involves a range of traits. Thus, dietary specialization will be facilitated by the correlated evolution of these traits. In **Chapter 3**, we used *Drosophila melanogaster* and yeast to explore the scope for dietary specialization. We quantified how different *D. melanogaster* strains from around the globe respond to different yeast species, across multiple yeast-dependent life history traits including feeding, mating, egg-laying, egg development and survival. We found fly strains varied in their responses to different yeast species: some strains performed well on a specific yeast species, while other strains did not. We did not detect the trade-offs in performance on different yeast species: performance on alternative yeast species is positively correlated. Yeast-dependent trait responses were not aligned: different life-history traits were maximized on different yeast species. Our results confirmed the existing insight that *D. melanogaster* is a resource generalist: it can grow, reproduce and survive on all the yeast species we tested.

Taken together, our findings suggest that there are evolutionary constraints for these important life-history traits to adapt in concert, possibly providing a mechanistic explanation of the limited extent of dietary specialization in *D. melanogaster* strains across the globe.

Heterogeneity in food resources can facilitate assortative mating: when mating takes place on food resources, assortative mating can arise as a by-product of food choice. In **Chapter 4 & 5**, we studied the spatial coupling of food and mates, the sensory and behavioural mechanisms underlying this spatial coupling, and whether this coupling would potentially promote assortative mating. To quantify the location of foraging and sexual behaviour of individuals in a heterogeneous environment, we built a system which combined the low-cost Raspberry Pi video recording system with the fast and efficient TRex tracking software and Matlab scripts to provide an automated, fast, efficient and easy-to-use method for fly tracking in **Chapter 4**. By tracking the location of fruit flies in environments containing heterogeneous food patches, we observed that *D. melanogaster*, either alone or in pairs with the opposite sex, stayed on yeast at night, but individuals' tendencies to be on the yeast during the day depended on several variables including sex, light conditions and presence of the other sex. To further explore the spatial coupling of food and mates and the underlying mechanism mediating the dynamics of the spatial coupling of food and mates, we tracked the mating location of both virgin and mated flies under different light conditions for 24h. We found that *D. melanogaster* and several of its sibling species generally chose to mate on patches containing yeast. In *D. melanogaster* however, virgin females primarily mated away from yeast, but previously mated females re-mated on yeast. Using experimental manipulation of the chemical composition of the yeast-containing patches and mutant flies lacking sex peptide (males; SP) and sex peptide receptor (females; SPR), we established the sensory and behavioural mechanisms underlying this spatial coupling of food and mates. We found that mating location preference involved attraction to yeast-derived chemical cues (the combination of acetic acid and protein) and was modulated by the male-derived sex peptide received by females during mating. This preference for mating on yeast-containing patches was stronger at night than during the day, and increased with the passing of time since the first mating. In choice experiments with two different yeast species, we discovered that *D. melanogaster* pairs exerted preferences for mating on one yeast species over another. Together, our study suggests that some level of assortative mating may result from the preference for mating on yeast, but the strength of such assortative mating will depend on several variables including the presence of multiple yeast species and timing of (re)mating with respect to light cycle.

Overall, this thesis dissects the mechanisms underlying food-mediated population differentiation and assortative mating. On one hand, we find that the inconsistency of responses between life-history traits forms a possible limitation to dietary specialization and population differentiation. On the other hand, we discover that food preference may directly lead to assortative mating through its spatial coupling with mates. Our findings highlight the importance of analyzing multiple life-history traits involved in food exploitation to explore

English summary

the scope of food-mediated population differentiation and examining several variables including mating status, the presence of multiple food resources and timing of (re)mating with respect to light cycle to assess the potential strength of assortative mating that may result from the preference for mating on food resources.

Nederlandse Samenvatting

Translation by Gerrit Potkamp

Adaptatie aan verschillende omgevingen is een belangrijk mechanisme in fenotypische evolutie en diversificatie van soorten. Voedsel is, te midden van verschillende omgevingsfactoren, een belangrijke aanjager van lokale adaptie en soortvorming. In vogels en insecten zijn tal van voorbeelden van voedsel-gemedieerde soortvorming aan te wijzen. Hoe populaties die verschillende voedselbronnen exploiteren differentiëren en reproductief geïsoleerd raken is echter niet goed begrepen. Het onderzoek gepresenteerd in dit proefschrift richtte zich op het ontleden hoe heterogene voedselbronnen populatiedifferentiatie en assortatieve paring tussen populaties kunnen aandrijven, door gebruik te maken van het modelorganisme *Drosophila melanogaster* en zijn essentiële voedselbron – gist.

Het chemosensorisch systeem is bij dieren het belangrijkste zintuiglijk kanaal in het detecteren van en reageren op voedselbronnen. Chemosensorische divergentie zou daarom een cruciale rol kunnen spelen in voedsel-gemedieerde adaptie en soortvorming. In **Hoofdstuk 2** formuleerden we zeven sleutelvragen om te ontdekken hoe het chemosensorisch systeem voedsel-gemedieerde ecologische soortvorming kan mediëren. We begonnen met het identificeren welke aspecten van voedselbronnen heterogeen zijn in een bepaalde omgeving (Vraag 1), omdat dit het startpunt van divergente selectie is. Daarna bespraken we welke aspecten van voedsel selectie uitoefenen op consumenten (Vraag 2), en verkenden hoe consumenten deze bronnen detecteren (Vraag 3), exploiteren (Vraag 4) en zich er aan adapteren (Vraag 5) om te begrijpen hoe lokale adaptie te werk gaat. Tenslotte bespraken we of succesvolle exploitatie van nieuwe voedselbronnen genetisch is geërfd en/of verworven is gedurende het leven van een individu (Vraag 6) en bekeken we de mechanismen die genoverdracht tussen populaties die zich op verschillende bronnen specialiseren reduceren (Vraag 7). De zeven geformuleerde vragen liet ons enkele gaten in onze kennis zien. In het bijzonder, verschillende benaderingen zijn nodig voor het beantwoorden van de verschillende vragen. Alle zeven vragen zijn maar zelden volledig beantwoord in een enkel modelsysteem. Het is vaak niet duidelijk wat voor soort heterogeniteit, of welk aspect van een bepaalde bron, divergentie in consumenten zal genereren. Vanwege de belangrijke rol die darmmicroben spelen in het vermogen bronnen te exploiteren en de partnerkeuze van gastheren wezen we deze microben in het darmstelsel aan als een veelbelovende richting voor toekomstig onderzoek naar voedsel-gemedieerde ecologische soortvorming.

De succesvolle exploitatie van een voedselbron is een fenotype met meerder facetten waar een scala aan eigenschappen bij betrokken is. Dieetspecialisatie zal daarom worden gefaciliteerd door de gecorreleerde evolutie van deze eigenschappen. In **Hoofdstuk 3** gebruikten we *Drosophila melanogaster* en gist om het domein van dieetspecialisatie te verkennen. We kwantificeerden hoe verschillende *D. melanogaster*-stammen van over de hele wereld reageren op verschillende gistsoorten, over meerdere gist-afhankelijke levensgeschiedenis-eigenschappen waaronder voeden, paren, het leggen van eitjes, de ontwikkeling van eitjes en overleving. We vonden dat vliegenstammen varieerden in hun respons op verschillende gist soorten: sommige stammen presteerden goed verschillende gistsoorten, andere stammen niet. We detecteerden geen compromis in prestatie op

verschillende gistsoorten: de prestatie op alternatieve gistsoorten is positief gecorreleerd. Gist-afhankelijke reacties waren niet afgestemd: verschillende levensgeschiedenis-eigenschappen waren gemaximaliseerd op verschillende gistsoorten. Onze resultaten bevestigden het bestaande idee dat *D. melanogaster* een generalist is: hij kan groeien, reproduceren en overleven op alle geteste gistsoorten. Onze bevindingen suggereren bij elkaar genomen dat er evolutionaire beperkingen voor deze belangrijke levensgeschiedenis-eigenschappen om zich gezamenlijk aan te passen bestaan, wat mogelijk een mechanistische verklaring biedt van de beperkte dieetspecialisatie van *D. melanogaster*-stammen verspreid over de wereld.

Heterogeniteit in voedselbronnen kan assortatieve paring faciliteren: wanneer paring plaatsvindt op voedselbronnen kan assortatieve paring ontstaan als een bijproduct van voedselkeuze. In **Hoofdstuk 4 & 5** bestudeerden we de ruimtelijke koppeling tussen voedsel en partners, de onderliggende sensorische en gedragsmatige mechanismen hiervan, en of deze koppeling assortatieve paring zou kunnen bevorderen. Om de locatie van foerageren en seksueel gedrag van individuen in een heterogene omgeving te kwantificeren hebben we in **Hoofdstuk 4** een systeem gebouwd dat een geautomatiseerde, snelle, efficiënte en gebruiksvriendelijke methode voor het volgen van vliegen biedt door het goedkope Raspberry Pi video-opnamesysteem te combineren met de snelle en efficiënte TRex tracking software en Matlab scripts. Door de locatie van fruitvliegen in omgevingen met heterogene voedselplekken te volgen observeerden we dat *D. melanogaster*, ofwel alleen of in paren met het andere geslacht, 's nachts op gist verbleef, maar dat de neiging van individuen zich overdag op gist te begeven afhankelijk was van verschillende variabelen, waaronder geslacht, de lichtcondities en de aanwezigheid van het andere geslacht. Om de ruimtelijke koppeling tussen voedsel en partners en de onderliggende mechanismen die de dynamiek van deze koppeling mediëren verder te onderzoeken volgden we de locatie van zowel maagdelijke als gepaarde vliegen in verschillende lichtcondities gedurende 24 uur. We vonden dat *D. melanogaster* en verschillende van zijn zustersoorten er in het algemeen voor kiezen te paren op plekken waar gist aanwezig is. In *D. melanogaster* echter paarden maagdelijke vrouwtjes voornamelijk weg van gist, terwijl eerder gepaarde vrouwtjes op gist opnieuw paarden. Door gebruik te maken van experimentele manipulatie van de chemische compositie van de plekken die gist bevatten en gemuteerde vliegen zonder geslachtspeptide (mannetjes; SP) en de receptor voor geslachtspeptide (vrouwtjes; SPR) stelden we de sensorische en gedragsmatige mechanismen die ten grondslag liggen aan de ruimtelijke koppeling van voedsel en partners vast. We vonden dat de voorkeur voor de paringslocatie te maken had met de aantrekkingskracht van chemische, van gist afgeleide, signalen (de combinatie van azijnzuur en eiwit) en werd gemoduleerd door de mannelijke geslachtspeptide die door vrouwtjes tijdens de paring wordt ontvangen. De voorkeur om te paren op plekken met gist was 's nachts sterker dan overdag, en nam toe met de tijd na de eerste paring. In keuze-experimenten met twee verschillende soorten gist ontdekten we dat *D. melanogaster*-paren voorkeur hadden om te paren op één gistsoort in plaats van op de andere soort. Samengevoegd suggereert onze studie dat een bepaald niveau van assortatieve paring het

Nederlandse samenvatting

gevolg kan zijn van de voorkeur te paren op gist, maar dat de sterkte van deze assortatieve paring afhankelijk zal zijn van verschillende variabelen, waaronder de aanwezigheid van verschillende gistsoorten en de timing van (opnieuw) paren ten opzichte van de lichtcyclus.

Samenvattend ontleedt dit proefschrift de mechanismen die ten grondslag liggen aan voedsel-gemedieerde populatiedifferentiatie en assortatieve paring. Aan de ene kant vinden we dat de inconsistentie van reacties tussen levensgeschiedenis-eigenschappen een mogelijke beperking van dieetspecialisatie en populatiedifferentiatie vormen. Aan de andere kant ontdekken we dat voedselvoorkeur direct tot assortatieve paring zou kunnen leiden door de ruimtelijke koppeling met partners. Onze bevindingen benadrukken het belang meerdere levensgeschiedenis-eigenschappen die betrokken zijn met de exploitatie van voedsel te bestuderen om het domein van voedsel-gemedieerde populatiedifferentiatie te ontdekken, en meerdere variabelen te onderzoeken, waaronder paringsstatus, de aanwezigheid van meerder voedselbronnen en de timing van (opnieuw) paren ten opzichte van de lichtcyclus, om de potentiële sterkte van assortatieve paring die het gevolg kan zijn van de voorkeur te paren op voedsel vast te stellen.

Acknowledgements

Acknowledgements

Time flies. It was almost five years ago that I took the flight from China to the Netherlands and started my new life in a very different world. As I naively planned, everything would be fine and I could definitely get my PhD. Indeed, everything turns out fine and I will get my PhD in the coming month. Look back, I feel so grateful for many people being in my PhD life. Without the support, encouragement and help of these people, I could not make such an achievement of getting my doctorate.

First and foremost, I would like to thank my two amazing PhD supervisors: **Jean-Christophe Billeter** and **Martine E. Maan**. I was very lucky to be the “scientific kid” of you two. Thank you so much for offering me the opportunity to do the PhD with you. It is such a pleasant and enjoyable journey for me to work with you two. One of the happiest moments of my PhD was to have meetings and retreats with you two to talk about science and a bit life. **Jean-Christophe**, thank you for being such a passionate supervisor for me. Your excitement of my PhD projects and positive attitude of all the small achievements of my experiments definitely motivated me to go further. You took your time to sit down with me many times to discuss my projects. You were there for me when I had problems. You offered me invaluable suggestions for my projects and writings. Your guidance and support will make me a better scientist. Thank you for organizing the wonderful parties and dinners at your place. I enjoyed the food you cook and felt so happy to be in such a wonderful team. Thank you for inviting me to visit Dijon. I had a great time for the short field collection, talking with the French scientists and conversations with you about my PhD projects on the train. Thank you for sharing your passion about science and some of your experiences about life. You helped me to grow up both in science and life. You encouraged me to be a fearless person and I hope I will always be. **Martine**, thank you for being such a calm and patient supervisor for me. I very often came to our meetings with problems and worries. But you helped me find clear directions and possible solutions at the end of our meetings. Thank you for listening to all my problems and worries, continuously asking me why and guiding me to rethink. I benefited from your way of critical thinking. Thank you for detailed and valuable feedback on my writings, especially when you were very busy. They were very inspiring and helpful. I think I will still go back to your comments sometime to remind myself about the important tips you gave. Thank you for organizing lab retreats, team retreats and drinks. It was a pleasure to talk with you and other lab members about my PhD projects. You are always willing to offer help not only in science but also in life. Thank you for calling me and offering help when I was in a panic for the covid-related issue. You are an extraordinary scientist and supervisor I hope one day I can be. I wish you all the best for the coming next.

My PhD life would not be so wonderful without all my awesome colleagues. I would like to thank all the fly people: **Gerard Overkamp**, **Jenke Gorter**, **Andrea Soto Padilla**, **Tiphaine Bailly**, **Pinar Kohlmeier**, **Mário Santos Mira**, **Philip Kohlmeier**, **Thomas A. Verschut**, **Nicolas Doubovetzky**, **Sanne Lamers**, **Adithya Sarma**, **Marijke Versteven**. Thank you all for the discussions during our fly lab meeting, the relaxing chats, drinks and parties. **Gerard**, you are the super hero of our lab. Thank you for making the enormous amount of fly food

and solving all the technical problems I have. You are very practical and efficient in solving lab issues. I am amazed at the design you came up with for my tracking platform and the helpful tips you gave for my posters and my experiments. **Jenke**, thank you for the instructions on making crosses and the mating experiments at the beginning of my PhD and the insightful comments on my fly-yeast matrix paper at the end of my PhD. **Andrea**, talking with you is always enjoyable and hilarious. **Tiphaine** and **Pinar**, thank you for welcoming me and showing me around when I came to the lab. You were like the sunshine that lightened my life in the lab. I can always share my ups and downs with you. You made my PhD life much better and colourful. I enjoyed the wonderful dinners, dancing, the bubble football, the laser tag, the BBQ, the paintball and the board games we had together. I wish both of you all the best for the new life in USA. **Mário**, thank you for helping me with my statistics and sending your greetings when I was stuck in China during the covid time. I like your humour and it was really pleasant to talk with you. Good luck with your writing and hope you will get to the end soon. **Philip**, I assume you will never forget the chicken nuggets. It was so hilarious when you tell stories. Thank you for your insightful comments on my fly-yeast matrix paper and my review. I wish you all the best for your new life in USA. **Thomas**, thank you for writing a review with me and sharing some of your experiences for building experimental box. I am amazed that you can be so organized. **Nicolas**, I am very glad that you took the invitation to be my paranymph. Thank you for helping me organize the coming big day and good luck with the rest of your PhD. **Sanne, Adi and Marijke**, you definitely brought new energy into our fly lab. I am glad to share the lab with you at the end of my PhD and thank you for the laughs you brought to the lab.

I also would like to thank all the fish people, my office mates and the first-floor people: **Shane Wright, Elodie Wilwert, Tiziana Gobbin, Gerrit Potkam, Aude Giraud, Joana Sabino Pinto, Neeraj Kumar, Asmoro Lelono, Yoran Gerritsma, Merijn Driessen, Tom Sarraude, Christina Bauch, Yuqi Wang, Flavia Berlingheri, Paolo Panizzon and the ones I may have forgotten to mention.** Thank you all for the help, chats, the drinks and the fun times. Special thanks to **Shane** for insightful comments on my fly-yeast matrix paper and my review and **Gerrit** for helping me translate my thesis summary to Dutch. Thanks to **EGDB expertise group** for all lunch talks, the discussions and feedbacks. To my two PhD coordinators: **Corine Eising** and **Diana Koopmans**, thank you for sending the emails to inform me about the courses, events, workshops and conferences.

To my Bachelor students **Koen Freerks** and **Maxim Juistenga**: thank you for joining me for a research project and giving me chance to learn how to teach and supervise students. I enjoyed being part of your research experience and felt so proud to see you two presenting your research project. **Koen**, thank you for choosing to join me for your master project as well. I was very lucky to have you as a master student at the end of my PhD. Thank you for your enthusiasm for this tracking project and the data you have collected. I am looking forward to reading your report in the near future.

Acknowledgements

To the reading committee for my PhD thesis: **Paul Becher** (Swedish University of Agricultural Sciences), **Bregje Wertheim** (University of Groningen) and **Martijn Egas** (University of Amsterdam), thanks to each of you for reading and evaluating my thesis. **Bregje**, it was so nice to know you and talk with you. Thank you for the tips you gave for my postdoc application when you came to the fly lab for your fly stock.

I gratefully acknowledge the **Chinese Scholarship Council** (CSC) and the **University of Groningen** for the joint scholarship to me. I cannot pursue my study and undertake my PhD project in the Netherlands without the financial support.

I sincerely thank my master supervisors **Jingfeng Chen** and **Yezhong Tang**. Thank you for the greetings and support during my PhD.

Massive thanks to the Chinese friends I met in Groningen: **Tian Xie & Mengfei Cai, Siwei Chen, Qing Chen, Minpeng Liang & Yuru Liu, Bohuan Lin, Weijia Yao & Sha Luo, Xiaobo Tian, Yanfei Li, Guangcai Xu, Feng Yan, Qi Chen, Huatang Cao, Shuai Feng, Zhiwen Wang, Yanyan Liu, Yanfang Wang, Fangfang Liu, Lisheng Zhang, Yinyu Xiang, Wenjian Li, Haibin Wen, Fan Yang, Yanmei Liu, Wei Jiang, Ming Shi, Jing Wu, Xinyu Zhou, Yafeng Song, Yifei Mao, Chongnan Ye & Yuhan Luo**. Thank you all for being part of my PhD.

Special thanks to the friends who give me enormous support throughout my PhD. **Tian**, you are the first person I know when I was preparing to come to Groningen. It was such wonderful “yuanfen” for us to know each other. You are such an enthusiastic friend who is always eager to learn more and try out different things. I feel so lucky to know you and learn from you. **Siwei**, you were the first roommate I had in Groningen. Thank you for sharing all the practical information. It was so helpful to have a roommate like you at the very beginning of my PhD. **Minpeng**, you were my neighbour both in Plutolaan and Plantenlaan. You are such a reliable and considerate friend. I can always get very useful suggestions and help from you. I wish you and **Yuru** all the best for your bright future. **Feng**, indeed, it is hard to find words to express how grateful I feel to have you as my friend. Thank you for the uncountable support and encouragement throughout my PhD. I will never forget how you stood by me through my dark time. I have no doubt that you will be successful whatever you choose to work on since you are so determined and concentrated. I wish you all the best for your new life in the UK. **Zhiwen**, you are a wonderful friend who I can always rely on. It is so lucky to have you as my neighbour. Thank you for sharing some nice food to me and giving a hand whenever I need it. Thank you for coming back for my defence. **Yanfei**, it is so comfortable to be together with you to talk about different things. Thank you for the amazing picnic we had and all the delicious food you made. Though it is a lot of work to take care of the baby, I think you will manage as you always do. **Yanmei**, you are the angel for me during the stressful period of my PhD. You remind me that I should thank all the people who come to my life so that I can move on in peace. Good luck with your final part of writing and looking forward to your defence. **Qi**, you can always come up with some hilarious ideas. I enjoyed the moment we

spent together for dinners, laser tag, bubble football, skating and running. All the best for finishing your writing and getting to the end of your PhD soon. **Bohuan**, I am amazed at your sincere curiosity for research and admire your deep thoughts about science. I enjoyed all the conversations we had. **Yafeng**, I am very happy to meet you again in Groningen. It was so nice that we had some fun time together in Beijing and in Groningen. Talking with you is so relaxing for me. I wish you all the best for your bright future.

Finally, to my family: my **Mum, Dad** and **Brother**, thank you for being there for me no matter whatever happens. Your unconditional support empowered me to pursue any goal I dreamed about. Your constant encouragement and positivity make me a free and fearless person. Mum and Dad, you are my energy station where I always get energy for being the person I want to be. Thank you for supporting all the decisions I made, no matter how far from home. I feel so lucky to be your daughter.

Last, but certainly not least, to **Xiufeng**, my best friend and my love, thank you for everything you did for me. Especially thank you for leaving everything behind and coming all the way from China to the Netherlands. You continuously surprise me with your wisdom, humour, bravery, flexibility, discipline and aspiration. Thank you for accompanying me for all the ups and downs, listening to all the complaints and keeping pushing me to go further.

The PhD journey will end soon, but the remarkable memories will stay forever. Thank you all.

Delft

Xiaocui Wang

October, 2022

Publications

Wang, X., Billeter, J. C., & Maan, M. E. (2022). Lack of alignment across yeast - dependent life - history traits may limit *Drosophila melanogaster* dietary specialization. *Journal of Evolutionary Biology*, 35(8), 1060-1071.

Wang, X., Verschut, T. A., Billeter, J. C., & Maan, M. E. (2021). Seven questions on the chemical ecology and neurogenetics of resource-mediated speciation. *Frontiers in Ecology and Evolution*, 9, 640486.

Wang, X., Zhao, Z. J., Cao, Y., Cui, J., Tang, Y., & Chen, J. (2019). Condition dependence of advertisement calls in male African clawed frogs. *Journal of Ethology*, 37(1), 75-81.

Cao, Y., Shen, J., **Wang, X.**, Tan, S., Li, P., Ran, J., ... & Chen, J. (2020). Effects of dietary protein variations at different life-stages on vocal dominance of the African clawed frogs. *Asian Herpetological Research*, 11(3), 249-257.

Chen, J., Qi, Y., Wu, Y., **Wang, X.**, & Tang, Y. (2019). Covariations between personality behaviors and metabolic/performance traits in an Asian agamid lizard (*Phrynocephalus vlangalii*). *PeerJ*, 7, e7205.

Manuscripts

Wang, X., Li, X., Maan, M. E., Billeter, J. C. A system for automated quantification of the foraging and sexual behavior of *Drosophila melanogaster* in heterogeneous environments. (In preparation)

Wang, X., Freerks, K., Maan, M. E., Billeter, J. C. Spatial coupling of food and mates in *Drosophila*. (In preparation)