Chapter 5

Role of eelgrass on bed-load transport and sediment resuspension under oscillatory flow

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Role of eelgrass on topsoil erosion

Abstract

Coastal vegetation is widely attributed to stabilize sediment. While most studies focused on how canopy causes flow reduction and thereby affect sediment dynamics, the role of roots and rhizomes on stabilizing the surface sediment has been less well studied. This study aims to quantify interactions between above and belowground biomass of eelgrass (i.e. living *Zostera marina* plants and mimics) with surface sediment erosion (i.e. bedload and suspended load), under different hydrodynamic forcing that was created using a wave flume. Belowground biomass played an important role preventing bedload erosion, by roughly halving the amount of sediment transported after being exposed to maximal orbital velocities of 27 cm s\(^{-1}\), with and without canopy. Surprisingly, for suspended sediment transport, we found opposite effects. In the presence of eelgrass, the critical erosion threshold started at lower velocities than on bare sediment, including sand and mud treatments. Moreover, in muddy systems, such resuspension reduced the light level below the minimum requirement of *Z. marina*. This surprising result for sediment resuspension was ascribed to a too small eelgrass patch for reducing waves but rather showing enhanced turbulence and scouring at meadow edges. Overall, we conclude that the conservation of the existent eelgrass meadows with developed roots and rhizomes is important for the sediment stabilization and the meadow scale should be taken into account to decrease sediment resuspension.
Introduction

Coastal vegetation provide a broad range of ecosystem services such as nutrient cycling, support for global fisheries, improvement of the water quality and carbon sequestration (Orth et al. 2006; Gedan et al. 2009; McLeod et al. 2011). In the face of global change, there is a growing interest in the role of mangroves, salt marshes and seagrass meadows in coastal protection (Temmerman et al. 2013; Bouma et al. 2014; Narayan et al. 2016). Coastal protection by vegetation is provided either by the standing biomass and/or by a reduction of the sediment erosion leading to an enhancement of higher foreshores, both related to wave attenuation (Bouma et al. 2014; Möller et al. 2014; Gracia et al. 2018). In this context, an in depth mechanistic understanding of the role of coastal vegetation on reducing sediment erosion is pivotal for making predictions in the future, where the frequency and magnitude of extreme sea levels is predicted to increase (Menéndez and Woodworth 2010; Vousdoukas et al. 2018).

Sediment erosion from the surface layer can be caused by the initiation of horizontal sediment transport (i.e., bedload) or by sediment resuspension (i.e., suspended load) (Einstein et al. 1940; Brown et al. 1995). Bedload occurs when sediment particles move along the bottom horizontally by rolling whereas sediment resuspension occurs when the sediment particles are lifted vertically into the water column creating turbidity and reducing the light (Einstein et al. 1940; Brown et al. 1995). Erosion of sediment with grain size smaller than 62.5 $\mu$m (mud) can be quantified as turbidity because these sediment particles are carried directly to the suspended load (Aberle et al. 2004). In contrast, sediment with particles larger than 62.5 $\mu$m (sand) will have bedload phase and should not be quantified only as turbidity (Aberle et al. 2004).

Several studies indicate that coastal vegetation may reduce erosion both on bedload form and resuspension (Ward et al. 1984; Christianen et al. 2013; Spencer et al. 2016), which is normally attributed to a reduction of the hydrodynamics within the canopy (Bos et al. 2007; Infantes et al. 2012; Möller et al. 2014). The reduction of the sediment erosion is important to maintain the water clarity, necessary for seagrass development (Dennison 1987; Duarte 1991), and to retain the sediment in coastal areas (Christianen et al. 2013; Ganthy et al. 2015; Spencer et al. 2016). Whereas a lot of work has focussed on the aboveground plant-flow interactions, the effect of belowground biomass (rhizomes and roots) on the sediment stabilization is still relatively poorly studied. The latter is especially true for seagrasses, despite of them being present in many coastal systems.

Reduction of erosion by belowground biomass has been previously assessed in terrestrial and saltmarsh plants (Baets et al. 2007; Feagin et al. 2009; Wang et al. 2017). For example, two types of erosion are common in coastal ecosystems: i) lateral cliff-erosion, which occur at the front of the (cliffed) saltmarsh, or in the edges of vegetation patches, and leads to narrowing of the marsh (Bouma et al. 2009b; Lo et al. 2017; Wang et al. 2017), and ii) horizontal surface-erosion, which is the gradual erosion of sediment particles from the surface layer in
between the plants, caused by the bed shear stress (Brown et al. 1995; Ganthy et al. 2015). Lateral cliff-erosion of saltmarshes seems to be controlled by the sediment type and root biomass at small scale (Feagin et al. 2009; Wang et al. 2017), and is outside the scope of the present study, as we focus on horizontal surface-erosion. In the case of seagrass, rhizomes and roots seem to play a major role in sediment stability (Christianen et al. 2013), but to the best of our knowledge, there are no mechanistic studies that directly quantify the effect of belowground biomass on surface-erosion. Similar to saltmarsh vegetation, we expect the effect of the seagrass belowground biomass on sediment stabilization to interact with other factors as the sediment properties and wave energy (Widdows et al. 2008; Feagin et al. 2009; Wang et al. 2017).

This study aims to i) quantify how much surface-erosion in the form of both suspended load and bedload transport are affected by the presence of eelgrass and ii) quantify the relative effect of aboveground and belowground biomass, sediment type and wave conditions on surface-erosion. To answer these questions, we carried out a flume experiment in which we used both artificial and natural eelgrass on muddy and sandy sediment, by applying a range of wave orbital velocities.

**Methods**

Eelgrass and sediment collection

Eelgrass samples with intact sediment were collected from the field to keep the sediment properties, above-ground and below-ground biomass undisturbed. Samples were collected in Bokevik bay in the Gullmars Fjord, Sweden (58°14’N, 11°26’E). Sediment with eelgrass was collected between 1.5 – 10 m depth to cover a range of sediment types, eelgrass densities and morphologies. Samples from 3-10 m depth were taken with a 0.35 x 0.35 m box-core from a vessel and placed into custom-made trays of 0.35 x 0.35 m (Dahl et al. 2018). The trays with the sediment were transported in PVC boxes to protect the sediment from tilting. Samples from shallow sites (1.5 m) were taken with cores of 12 cm diameter using scuba or snorkelling because the vessel with the box-core could not reach the shallow waters. The sediment thickness collected varied from 5 to 10 cm depending on the sediment compactness. To maintain the plants in optimal conditions until the hydrodynamic experiments, the sediment trays and cores were stored in shallow-water flow-through 1500 L outdoor tanks at the Sven Lovén Centre for Marine Infrastructure, Kristineberg.

Wave flume

The study was conducted in a hydraulic wave flume developed and constructed at the Netherlands Institute for Sea Research (NIOZ) and located at Kristineberg Marine Research Station (Fig. 1a). The wave flume was 3.5 m long, 0.6 m wide and 0.8 m high (Fig. 1b). Waves were generated with a pneumatic piston and damped with a wave absorber made of synthetic
fibre with a slope of 20º. The test section was composed of a PVC box of 0.35 x 0.35 x 0.15 m (length x width x height). The sediment trays were carefully inserted in the test section and adjusted in order to be at the same level as the flume bottom (Fig. 1a). In the case of the samples from shallow sites, 5 cores were carefully inserted together to fill the test section and one additional core was sliced to fill any remaining gap, producing a continuous bed of minimal disturbed sediment. The flume was filled with seawater until 25 cm depth. For each sediment sample, orbital flow velocities were increased stepwise from 2 to 27 cm s⁻¹, with each time step maintained during 8 min (56 minutes of wave exposure in total) (Table S1). These conditions represent similar wave exposures in shallow bays where eelgrass is present in the Swedish west coast (Infantes, unpublished data). Waves were measured next to the sample box with a pressure sensor (Druck, PT1830) and sampling rate of 25 Hz. Wave height ($H$) was calculated from the pressure data as,

$$H = 2 \cdot \sqrt{\frac{\sum_{i=0}^{n_i} P_i^2}{n}} \cdot \sqrt{2}$$  \hspace{1cm} [1]

where $P$ is the pressure data.

Flow velocities were measured with an acoustic Doppler velocimeter, ADV (Nortek, Vectrino) at 10 cm above the bottom and 5 cm in front of the test section to not interfere with the canopy. The sampling rate was 25 Hz, sampling volume of 7 mm, and velocity range of 0.3 m s⁻¹. Mean orbital velocities ($U_{rms}$, cm s⁻¹) were calculated as,

$$U_{rms} = \sqrt{\frac{1}{N} \sum_{i=1}^{n} (u_i^2)}$$  \hspace{1cm} [2]

where $u$ is the horizontal flow velocity during $n$ measurement points.

Fig. 1. a) Diagram of the hydraulic wave flume and sensors. The wave flume is a further elaboration of the wave mesocosms used by La Nafie et al. (2012) and the wave tanks used by Wang et al. (2017) and (Lo et al. 2017), b) diagram of the top view of the wave flume. The total length of the flume is 3.5 m, 0.6 m wide and 0.8 m high.
Suspended load and critical erosion thresholds

Erosion as suspended load and the critical erosion threshold per treatment were assessed by measuring the turbidity and light reduction of the water column. Since sediment type and eelgrass densities varied widely in the field, a first trial with mimic plants aimed to assess the effect of both the absence/presence of shoots, and shoot density on sediment resuspension using the same sediment across treatments. These trials were performed both with muddy and sandy sediment and three treatments (Fig. 2a, b): 1) bare sediment, 2) 40 shoots of mimics in the test section, equivalent to 333 shoots m$^{-2}$, considered low density and 3) 90 shoots of mimics in the test section, equivalent to 750 shoots m$^{-2}$, considered high density (Lefebvre et al. 2010). Mimic shoots were made from four polyethylene blades of 25 cm length, 2 mm width and 1 mm of thickness attached to a wooden dowel with a 4 cm plastic straw of 0.4 cm of diameter (see González-Ortiz et al. 2014). Muddy and sandy sediment was prepared by homogeneously mixing sediment from the field that was previously sieved (2 mm) to remove shells and debris (Table 1).

To quantify the effect of above and belowground vegetation in sediment resuspension in different sediment types, three treatments were made with muddy and sandy intact sediment from the field: 1) eelgrass with both the aboveground and belowground biomass being present, 2) eelgrass from which only the rhizomes and roots were present, by cutting the aboveground leaves and 3) bare sediment without eelgrass (Fig. 2c, d). Treatment 2 was made only with sandy samples, since pilot trials showed similar results in muddy sediment with eelgrass and only-rhizomes. Three to seven replicates of each treatment were carried out. Hence, our treatments consisted of the test section filled with homogeneous sediment in which we also placed two densities of eelgrass mimics with a random distribution (333 vs. 750 shoots m$^{-2}$), and natural sediment with either intact eelgrass plants, with eelgrass roots and rhizomes only, and without any eelgrass (Fig. 2).

Water turbidity and the percentage of surface light reaching the bottom were measured during the last 2 min of each time step in all experiments. Water turbidity was measured with a turbidity meter (Campbell, OBS), located 10 cm after the sample box to be in line with the light sensors (Fig. 1a) and 5 cm above the bottom, at a sampling frequency of 25 Hz. At the moment of the measurements, the turbidity was homogeneous in the whole flume. Voltage data from the turbidity meter was calibrated to mg L$^{-1}$ of suspended particles by filtering 0.5 L of water at different concentrations on pre-weighed filters and calculating the weight difference after drying for 48 h at 60 $^\circ$C. Photosynthetic active radiation (PAR) was measured using two Apogee light meters separated 14 cm for later calculation of the light attenuation coefficients ($K_0$) and percentage of incident radiation at the bottom (Fig. 1a). Light was generated using two Sirio 2070, 500 W lamps placed 1.1 m above the water surface level. The height of the lamps was chosen in order to provide enough light to the light meter in the bottom. $K_0$ was calculated as,
\[ K_d = \frac{\ln \left( \frac{I_{Z2}}{I_{Z1}} \right)}{Z_2 - Z_1} \]  

where \( I_z \) is the PAR irradiance at depth \( Z \); and \( I_{Z2} \) at the deeper depth, \( Z_2 \) (Beer et al. 2014). The % of light at the bottom was calculated as (Beer et al. 2014).

\[ \% \text{ light at the bottom} = \frac{I_{Z2}}{I_{Z1}} \times 100 \]  

The critical erosion threshold was determined as a measurement of sediment stability. Two erosion phases can be distinguished accordingly to Amos et al. (1992, 1997): a first slow lineal increase of the resuspension with increasing orbital velocities (erosion Type I), and a second rapid increase of resuspension with increasing orbital velocities (erosion Type II). The first phase may correspond to the resuspension of the organic “fluff” layer (Amos et al. 1992, 1997; Bale et al. 2006). The critical erosions thresholds were determined as the start of the Type II erosion from scattegrams of turbidity plotted against the orbital velocities. The Type II erosion was fitted to an exponential regression where turbidity values below 9.5 mg L\(^{-1}\) (ambient concentration) were not accounted.

Bedload erosion with eelgrass plants

To quantify the role of eelgrass on horizontal sandy sediment transport, bedload erosion was assessed in sandy samples after the exposition to the 7 different wave settings ranging from 0 to 27 cm s\(^{-1}\) (Table S1). This experiment was performed with the three sandy treatments (Fig. 2c): 1) eelgrass aboveground and belowground biomass, 2) rhizomes and roots only and 3) bare sediment without eelgrass. We defined bedload erosion as the sediment transported outside of the test-section box. After the trials, the water of the flume was emptied, while the sandy sediment remained on the flume bottom. Then, the sediment deposited on the flume bottom outside of the test-section was collected with a window wiper and a dustpan. The sediment was then dried at 60\(^{\circ}\)C for one week and weighed.
Fig. 2. Experimental design where it was quantified a) suspended load with mimic plants and sandy sediment b) suspended load with mimic plants and muddy sediment, c) suspended load and bedload with eelgrass and sandy sediment and d) suspended load with eelgrass and muddy sediment. Treatment “Rhizomes-only” include root and rhizomes biomass.

Sediment and vegetation properties

Sediment bulk density, organic content, water content and grain size of the 1-2 cm top layer were determined for each sample. Bulk density was calculated as sediment dry weight in a volume of 20 ml. Water content was calculated as the difference of wet and dry weight. Organic content was determined by loss on ignition (LoI) method after burning the sediment sample for 5h at 450°C. The sediment grain size was analyzed using a Malvern® Mastersizer 2000. Sediment samples with mean grain size above 62.5 µm were classified as sandy, while grain sizes below 62.5 µm were classified as muddy.

Plant morphologies were measured at the end of the experiment for each sample. The length and thickness of the leaves and rhizomes were measured. The total root length per sample was extrapolated from the total root biomass of the sample and the dry weight of three subsamples of 15 to 20 random roots selected from the sample, which were previously measured and then dried at 60°C for 48h. Then, the diameter of the roots was measured in the subsamples. The root length density was calculated using the total root length per volume of sediment (see Baets et al. 2007). Above-ground and below-ground biomass was calculated by drying separately the leaves, roots and rhizomes at 60°C for 48h.

Statistical analysis

One-way ANOVA’s were used to analyze significant differences in sediment properties, bedload erosion and turbidity between the treatments followed by a Tukey HSD post hoc test. Two-way ANOVA was used to analyze the effect of eelgrass presence and sediment type on turbidity. Differences at P values of 0.05 were considered significant. Turbidity values at 25 cm s⁻¹ for each sample were used for the analysis. Spearman correlation coefficients ($r_s$) and principal
component analysis (PCA) (Fig. S1) were done with all the treatments to assess possible correlations between i) turbidity and plant/sediment characteristics and ii) bedload erosion and plant/sediment characteristics. Data was standardized for the PCA.

**Results**

Suspended load and critical erosion thresholds

Sediment resuspension increased at higher densities of eelgrass mimics for both sandy and muddy sediments (Fig. 3a, b). The increase of resuspension was linear during the erosion Type I and exponential during the erosion Type II. After the exposure to all the wave settings, the highest mimic density (750 shoots m⁻²) reached a maximum turbidity of 58 mg L⁻¹ in sandy sediment, while in muddy sediment the turbidity was five times larger, 290 mg L⁻¹. The critical erosion threshold (increase of turbidity and reduction of $K_d$) started at orbital velocities above 12 cm s⁻¹ in sand with eelgrass mimics and at 13.7 cm s⁻¹ for bare sand (Fig. 3a). In contrast, the critical erosion threshold started with orbital velocities above 5 cm s⁻¹ with 750 shoots m⁻² and 9 cm s⁻¹ with 333 shoots m⁻² in mud with eelgrass mimics (Fig. 3b). The critical erosion threshold in bare mud was lower than with mimics (5 cm s⁻¹), and contrary to the treatments with mimics, turbidity increased linearly at orbital flow velocities above 9 cm s⁻¹ (Fig. 3b). The increase in turbidity showed a significant correlation with the light attenuation coefficient ($K_d$) $(R^2=0.97, p<0.001)$. The maximum $K_d$ ~ 3.8 m⁻¹ and ~ 25 m⁻¹ were obtained with sandy and muddy sediment respectively at orbital velocities between 21-29 cm s⁻¹.

Sediment resuspension in natural samples followed a linear increase during the erosion Type I and an exponential increase during the erosion Type II in all the treatments (Fig. 3c, d). Critical erosion threshold started at orbital flow velocities around 9.4 cm s⁻¹ in sandy sediment with eelgrass, 10.5 cm s⁻¹ with rhizomes-only and at 14.4 cm s⁻¹ in bare sediment (Fig. 3c). In contrast, the critical erosion threshold in muddy sediment started at velocities of 5.1 cm s⁻¹ in eelgrass samples and 7.6 cm s⁻¹ in bare sediment (Fig. 3d). Turbidity and $K_d$ were lower in sandy sediments than in muddy (Fig. 3). $K_d$ reached maximum values of ~ 3.6 m⁻¹ and ~ 21.4 m⁻¹ in sand and mud respectively at orbital velocities between 22-29 cm s⁻¹. In the muddy trials, rhizomes and roots started to be uprooted around 15 cm s⁻¹, which was not the case in the sandy trials. In addition, the turbulent kinetic energy (TKE) was calculated from the ADV data as a measurement of turbulence, which was correlated to the $U_{rms}$. Calculations of the TKE and plots of the turbidity with TKE can be found in the supplementary information Fig. S2.
Fig. 3. Orbital flow velocities ($U_{rms}$, cm s$^{-1}$) and turbidity (mg L$^{-1}$) for a) mimic eelgrass with sandy sediment, b) mimic eelgrass with muddy sediment, c) natural eelgrass with sandy sediment and d) natural eelgrass with muddy sediment. The dashed horizontal line represents the ambient concentration (9.5 mg L$^{-1}$). The dashed vertical arrows represent the observed critical erosion threshold for eelgrass with sand (12 cm s$^{-1}$), rhizomes-only with sand (14 cm s$^{-1}$), bare sand (17.5 cm s$^{-1}$), eelgrass with mud (4 cm s$^{-1}$) and bare mud (11 cm s$^{-1}$) respectively. Each point represents a single measurement. Turbidity against the TKE can be found in Fig. S2.

Eelgrass presence was correlated with higher turbidity in both sandy ($r_s = 0.61, p<0.01$) and muddy sediment ($r_s = 0.81, p<0.01$). ANOVA and Tukey HSD tested for the turbidity reached at 21 and 25 cm s$^{-1}$, showed significant differences between all the treatments except between rhizomes-only and eelgrass with sand (One-way ANOVA: 21 cm s$^{-1}$: $F_{2,13} = 14.91, p<0.001$; 25 cm s$^{-1}$: $F_{2,13} = 11.81, p<0.001$). Given a flume water depth of 25 cm, only the muddy trials reduced the light below the minimum light-requirement for Z. marina (20% of surface light) for both mimics and natural eelgrass (Fig. 4). In contrast, none of the sandy trials with or without eelgrass reached the minimum light-requirement threshold. In the eelgrass mimics experiment, the light threshold was reached at 10 cm s$^{-1}$ and 15 cm s$^{-1}$ with 750 shoots m$^{-2}$ and 333 shoots m$^{-2}$ respectively. In natural eelgrass with muddy sediment,
the light threshold was reached at orbital velocities of 15 cm s\(^{-1}\) with eelgrass and 25 cm s\(^{-1}\) with bare sediment (Fig. 4).

Fig. 4. Percentage of surface light reaching the bottom with increasing flow orbital velocities (\(U_{rms}, \text{cm s}^{-1}\)) for a) mimic eelgrass and b) natural eelgrass. The dot line indicates the minimum light requirement for eelgrass growth (20 %).
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Bedload erosion

Sediment erosion in the form of bedload transport was significantly lower in the presence of eelgrass and rhizomes-only, when compared to bare sediment (One-way ANOVA: $F_{2,7} = 24.21$, $p<0.05$) (Fig. 5). The total erosion after one hour of wave exposure was similar for vegetated sandy sediment, vegetated muddy sediment and bare muddy sediment, although erosion in sandy trials was mainly as bedload while in muddy trials was mainly as suspended load (Fig. 5). In contrast, bare sandy sediment had a predominance of bedload transport compared to the other treatments (Fig. 5).

![Fig. 5. Total erosion (g) in the form of bedload and suspended load for sandy and muddy trials after 56 min of wave exposure (Mean ± SD). Significant differences in the bedload of sandy treatments are indicated by upper case letters and differences in the suspended load between all the treatments are indicated by lower case letters (Tukey HSD, $p<0.05$). Bedload erosion in muddy sediment is negligible. Total erosion as suspended load (g) was extrapolated from the turbidity (mg L$^{-1}$) at the end of the trials to the volume of water contained in the flume, assuming that the turbidity was homogeneous in the whole flume.](#)

Sediment and vegetation properties

Sediment and vegetation properties are summarized in Table 1. The sediment water content and organic content were significantly higher in the muddy sediment, whereas the bulk density was significantly higher in sandy sediment (One-way ANOVA: SWC: $F_{4,19}=164.6$, $p<0.001$, OC: $F_{4,19}=46.8$, $p<0.001$, Bulk density: $F_{4,19}=82.09$, $p<0.001$). No significant differences were found in bulk density, water content or organic content comparing within the sandy or muddy trials separately.
Eelgrass morphology varied between the samples present in sand and mud (Table 1). Leaves were shorter (< 30 cm) and thinner (0.3 cm), rhizomes were thinner (0.25 cm) and root diameter was thinner (0.05 cm) in sandy samples. In contrast, leaves were longer (> 30 cm) and wider (0.7 cm), rhizomes were thicker (0.5 cm) and root diameter was larger (0.07 cm) in muddy samples. In contrast to muddy samples, eelgrass in sandy sediment formed a dense network of roots and rhizomes which aggregated and retained the sediment.

Table 1. Vegetation and sediment properties. Mean (Std. Err).

<table>
<thead>
<tr>
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<th>Mimic eelgrass</th>
<th>Natural eelgrass</th>
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<tr>
<td></td>
<td>Sand</td>
<td>Mud</td>
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<td><strong>Seagrass morphology</strong></td>
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<tr>
<td>nº of shoots</td>
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<td>Root diameter (cm)</td>
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<td>Total rhiz length (m)</td>
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<td>Total root length (m)</td>
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<td>Root length density (cm cm⁻³)</td>
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<td>DW leaves (g)</td>
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<tr>
<td>DW Rhiz+roots (g)</td>
<td>-</td>
<td>-</td>
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<tr>
<td><strong>Biomass</strong></td>
<td></td>
<td></td>
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<tr>
<td><strong>Water content (%)</strong></td>
<td>19.9</td>
<td>59.6</td>
</tr>
<tr>
<td><strong>Organic content (%)</strong></td>
<td>0.4</td>
<td>5.5</td>
</tr>
<tr>
<td><strong>Bulk density (g cm⁻³)</strong></td>
<td>1.7</td>
<td>0.6</td>
</tr>
<tr>
<td><strong>Sand &gt;62.5 µm (%)</strong></td>
<td>86.2</td>
<td>3.7</td>
</tr>
<tr>
<td><strong>Mud &lt;62.5 µm (silt + clay) (%)</strong></td>
<td>14.1</td>
<td>96.4</td>
</tr>
<tr>
<td><strong>SD50 (µm)</strong></td>
<td>256.1</td>
<td>42.1</td>
</tr>
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</table>

*DW = dry weight

Water content, organic content and the percentage of mud were significantly correlated to higher turbidity, comparing all the samples (mud and sand) (Fig. 6). Bulk density, the percentage of sand and the median grain size (SD50) were correlated to lower turbidity (Fig. 6). No significant correlations with plant morphology and turbidity were found. There was a significant interaction between sediment type (mud or sand) and eelgrass presence (Two-way ANOVA: F₁,₁₉=173, p<0.001), which led to the maximum rates of turbidity and lower critical erosion thresholds in the mud with eelgrass treatments (Fig. 3).
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Fig. 6. Matrix plot of the variables significantly correlated with the turbidity. All the Spearman correlation coefficients ($r_s$) are significant ($p < 0.001$). SWC = sediment water content (%), OC = organic content (%), Bulk = bulk density (g cm$^{-3}$), Sand (%), Mud (%), SD50 = median grain size (µm), Turb25 = turbidity (mg L$^{-1}$) reached at orbital velocities around 25 cm s$^{-1}$.

Belowground biomass was the only variable significantly correlated with less bedload erosion in sandy sediment (Spearman, $r_s=-0.68$, $p<0.001$) (Fig. 7a). None of the morphologic traits were significantly correlated with bedload erosion. Eelgrass with lower percentage of mud seems to reduce less the bedload erosion compared to eelgrass with higher percentage of mud, although the correlation was not significant (Fig. 7b).

Fig. 7. a) Correlation between belowground biomass (g) and bedload erosion (g). Spearman correlation= -0.68, $p<0.001$. b) Scattergram between mud content and bedload erosion (g). Eelgrass and only-rhizome trials, even with variable mud content, had lower bedload erosion than bare sediment trials.
**DISCUSSION**

This study showed that the bedload erosion was reduced in sandy trials with eelgrass (either with leaves or only rhizomes and roots) compared with bare sediment, even at the highest wave velocities (~27 cm s\(^{-1}\)). Nevertheless, turbidity increased in the presence of natural eelgrass or mimics compared to bare sediment at orbital velocities above 5 and 10 cm s\(^{-1}\) in both muddy and sandy sediments respectively. Sediment properties showed key differences in the maximum turbidity reached with mud (~272 mg L\(^{-1}\)) and with sand (~74 mg L\(^{-1}\)) and the type of erosion (i.e., suspended load with mud vs. bedload with sand).

**Eelgrass presence on sediment resuspension**

Previous studies show that seagrass can reduce sediment resuspension by a reduction of the water flow and the shear stress compared to the unvegetated areas, either with currents (Gambi et al. 1990; Widdows et al. 2008) or waves (Ward et al. 1984; Hansen and Reidenbach 2012; Infantes et al. 2012; Ros et al. 2014). Nevertheless, the results of this study showed that eelgrass did enhance the resuspension of the sediment and lowered the critical erosion threshold both in sandy and muddy trials. Hydrodynamics inside the eelgrass patch could not be measured due to the small size (0.12 m\(^2\), 35 x 35 cm). Other experiments with patches ranging from 2.2 m to 0.3 m width have shown to increase the sediment dynamics created by the turbulence generated by the shoots or by the meadow edges under both currents (Bouma et al. 2007, Fonseca and Koehl 2006, Chen et al. 2012) and waves (Granata et al. 2001). Low plant densities could also increase the turbulence and scouring around shoots as flow moves through the sparse canopy (Bouma et al. 2009; Lefebvre et al. 2010). This increase in turbulence and scouring by the eelgrass presence suggests the observed increase in sediment resuspension and turbidity in all trials.

**Sediment characteristics on turbidity and light attenuation**

The comparison between muddy and sandy trials indicate that the critical erosion threshold and the subsequent increase in turbidity and light attenuation (\(K_d\)) are dependent on the sediment properties, in accordance with Bale et al. (2006), together with the interaction with eelgrass presence. In our experimental set up with a water depth of 25 cm, only the muddy sediment reduced the light below the minimum 20%, assessed for *Z. marina* (Dennison et al. 1993). Sandy treatments led to a major part of erosion in the form of bedload (from 2567 to 918 g) and a small part as suspended load (from 154 to 329 g), not causing high turbidity nor light reduction, in agreement with Houwing (1999). In contrast, muddy sediment with mean grain size smaller than 62.5 \(\mu\)m, passed directly to suspended load with negligible bedload phase, causing higher turbidity and light reduction (Widdows et al. 2008; Grabowski et al. 2011). The response was similar for mimic treatments, although mimics needed less wave velocities to reduce the percentage of light than natural eelgrass (~10 cm s\(^{-1}\) and 15 cm s\(^{-1}\) respectively).
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Mimic shoots were slightly stiffer than natural eelgrass, which is related with more scouring and turbulences around the shoots (Bouma et al. 2009a; Ros et al. 2014).

Importance of the belowground biomass: Applications for conservation and restoration

Bedload erosion of sandy sediment was reduced in treatments with eelgrass compared to bare sediment. Furthermore, there were no differences between the treatments of full canopy eelgrass (above- and belowground biomass), and only roots and rhizomes (only belowground biomass), suggesting that the effect of sediment stabilization is mediated by the belowground biomass rather than the canopy. These results are important and in line with earlier findings from Christianen et al. (2013), which increases the still limited available literature on this topic. In this study, eelgrass present in sandy sediment had a dense network of roots and rhizomes with root length densities of 0.8 cm cm\(^{-3}\). In terrestrial plants, a high density of roots with less than 1 mm of diameter are related with less erosion (Baets et al. 2007). In contrast, eelgrass in muddy sediment with root length density of 0.003 cm cm\(^{-3}\) led to less aggregation and retention of the sediment, as found by Widdows et al. (2008). In addition, the sediment properties of sandy eelgrass samples with higher percentage mud (>20 %), might have increased the sediment cohesiveness reducing even more the erodibility (Brown et al. 1995; Gailani et al. 2001) (Fig. 7b). Nevertheless, eelgrass samples with less mud (< 5 %) still had less bedload erosion than the bare sediment (with higher % of mud) (Fig. 7b), suggesting again that belowground biomass may play a major role reducing bedload erosion.

This study underlines the importance of the conservation of the existent eelgrass meadows with developed belowground biomass to reduce the sediment erosion by bedload transport. The data obtained in this study confirms that an eelgrass patch of 0.12 m\(^2\) with developed belowground biomass has a stabilizing effect of the sediment, reducing bedload transport. On the other hand, such a small patch will not have any effect preventing resuspension of the sediment. This experiment provides more evidence that the fragmentation of the meadows due to anthropogenic causes could increase the turbidity by exposing more edges of the meadow to hydrodynamics which increases the sediment resuspension (Allaoui et al. 2016).

Sediment characteristics as bulk density and grain size, exposure to hydrodynamics and patch size are important factors to consider during eelgrass restoration. Our results suggest that in sites with sediment median grain size smaller than 75 µm and exposure to orbital velocities above 10 cm s\(^{-1}\), sediment resuspension may be a problem in the restorations (Van Der Heide et al. 2007; Moksnes et al. 2018), and larger patches than 0.12 m\(^2\) might be needed to reduce sediment resuspension (Silliman et al. 2015; van Katwijk et al. 2016). On the other hand, in sandy sediments with mean grain size larger than 130 µm and exposed to orbital velocities up to 30 cm s\(^{-1}\), sediment resuspension might not reduce the light below the 20% in shallow waters (Adams et al. 2016). Bedload erosion might however be the cause of restoration failure due to the lack of a developed network of roots and rhizomes and uprooting of the initial plant.
units. In this case, to start a self-reinforcing feedback with sediment stabilization and low bedload erosion, wave barriers (Maxwell et al. 2017) or biodegradable geotextiles (Zanuttigh et al. 2015) could be implemented to reduce hydrodynamics and stabilize the sediment at the restoration site until the seagrass patches develop a dense root and rhizome network.

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Table S1. Wave settings generated with the hydraulic flume.

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Fig. S1. PCA of the two principal component (PC) indicating the more explanatory variables related to the a) turbidity and b) bedload erosion, separated per treatments. The size of the colour circles is a rescale of the quantity of turbidity/bedload erosion in each sample. BG=belowground biomass; DW= dry weight; OC=organic content; WC=Water content; Bulk= bulk density; S=sand; Rhiz=rhizomes.
Fig. S2. Turbulent kinetic energy (TKE, m$^2$s$^{-2}$) and turbidity (mg L$^{-1}$) for a) mimic eelgrass with sandy sediment, b) mimic eelgrass with muddy sediment, c) natural eelgrass with sandy sediment and d) natural eelgrass with muddy sediment. The dashed horizontal line represents the ambient concentration (9.5 mg L$^{-1}$). Each point represents a single measurement. TKE was calculated from the ADV data as 

\[ \frac{1}{2}(u'^2 + v'^2 + w'^2), \]

in m$^2$s$^{-2}$, as a measurement of turbulent fluctuation. $u$, $v$, and $w$ are the velocities in $x$, $y$ and $z$ direction respectively. $u'$, $v'$ and $z'$ are the components of turbulent velocity, calculated as:

\[
u'(t) = v(t) - \bar{v}.
\]

\[
u'(t) = v(t) - \bar{v}.
\]

\[
u'(t) = v(t) - \bar{v}.
\]