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Deciphering cellular heterogeneity of the brain

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Chapter 7

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References

- Abbott, N. J., Rönnebeck, L., & Hansson, E. (2006). Astrocyte–endothelial interactions at the blood–brain barrier. *Nature Reviews Neuroscience*, 7(1), 41–53. <https://doi.org/10.1038/nrn1824>
- Abrams, D., Kumar, P., Karuturi, R. K. M., & George, J. (2019). A computational method to aid the design and analysis of single cell RNA-seq experiments for cell type identification. *BMC Bioinformatics*, 20(S11), 275. <https://doi.org/10.1186/s12859-019-2817-2>
- Abramzon, Y. A., Fratta, P., Traynor, B. J., & Chia, R. (2020). The overlapping genetics of amyotrophic lateral sclerosis and frontotemporal dementia. *Frontiers in Neuroscience*, 14, 42. <https://doi.org/10.3389/fnins.2020.00042>
- Abreu, S. L. (1982). Suppression of experimental allergic encephalomyelitis by interferon. *Immunological Communications*, 11(1), 1–7. <https://doi.org/10.3109/08820138209050718>
- Agarwal, D., Sandor, C., Volpato, V., Caffrey, T. M., Monzón-Sandoval, J., Bowden, R., ... Webber, C. (2020). A single-cell atlas of the human substantia nigra reveals cell-specific pathways associated with neurological disorders. *Nature Communications*, 11(1), 4183. <https://doi.org/10.1038/s41467-020-17876-0>
- Alata, W., Ye, Y., St-Amour, I., Vandal, M., & Calon, F. (2015). Human apolipoprotein E4 expression impairs cerebral vascularization and blood-brain barrier function in mice. *Journal of Cerebral Blood Flow and Metabolism*, 35(1), 86–94. <https://doi.org/10.1038/jcbfm.2014.172>
- Allen, I. V., McQuaid, S., Mirakhur, M., & Nevin, G. (2001). Pathological abnormalities in the normal-appearing white matter in multiple sclerosis. *Neurological Sciences*, 22(2), 141–144. <https://doi.org/10.1007/s100720170012>
- Alsema, A. M., Jiang, Q., Kracht, L., Gerrits, E., Dubbelaar, M. L., Miedema, A., ... Eggen, B. J. L. (2020). Profiling microglia from Alzheimer’s disease donors and non-demented elderly in acute human postmortem cortical tissue. *Frontiers in Molecular Neuroscience*, 13, 134. <https://doi.org/10.3389/fnmol.2020.00134>
- Anders, S., Pyl, P. T., & Huber, W. (2015). HTSeq—a Python framework to work with high-throughput sequencing data. *Bioinformatics*, 31(2), 166–169. <https://doi.org/10.1093/bioinformatics/btu638>
- André, S., Kojima, S., Yamazaki, N., Fink, C., Kaltner, H., Kayser, K., & Gabius, H.-J. (1999). Galectins-1 and -3 and their ligands in tumor biology. *Journal of Cancer Research and Clinical Oncology*, 125(8–9), 461–474. <https://doi.org/10.1007/s004320050303>
- Andreone, B. J., Chow, B. W., Tata, A., Lacoste, B., Ben-Zvi, A., Bullock, K., ... Gu, C. (2017). Blood-brain barrier permeability is regulated by lipid transport-dependent suppression of caveolae-mediated transcytosis. *Neuron*, 94(3), 581–594. <https://doi.org/10.1016/j.neuron.2017.03.043>
- Araki, K., Meguro, H., Kushiya, E., Takayama, C., Inoue, Y., & Mishina, M. (1993). Selective expression of the glutamate receptor channel $\delta 2$ subunit in cerebellar Purkinje cells. *Biochemical and Biophysical Research Communications*, 197(3), 1267–1276. <https://doi.org/10.1006/BBRC.1993.2614>
- Armulik, A., Genové, G., & Betsholtz, C. (2011). Pericytes: developmental, physiological, and pathological perspectives, problems, and promises. *Developmental Cell*, 21(2), 193–215.

<https://doi.org/10.1016/j.devcel.2011.07.001>

Armulik, A., Genové, G., Mäe, M., Nisancioglu, M. H., Wallgard, E., Niaudet, C., ... Betsholtz, C. (2010). Pericytes regulate the blood-brain barrier. *Nature*, 468(7323), 557–561. <https://doi.org/10.1038/nature09522>

Ascherio, A. (2013). Environmental factors in multiple sclerosis. *Expert Review of Neurotherapeutics*, 13(12 Suppl), 3–9. <https://doi.org/10.1586/14737175.2013.865866>

Ayata, P., Badimon, A., Strasburger, H. J., Duff, M. K., Montgomery, S. E., Loh, Y.-H. E., ... Schaefer, A. (2018). Epigenetic regulation of brain region-specific microglia clearance activity. *Nature Neuroscience*, 21(8), 1049–1060. <https://doi.org/10.1038/s41593-018-0192-3>

Bacher, R., Chu, L.-F., Leng, N., Gasch, A. P., Thomson, J. A., Stewart, R. M., ... Kendzioriski, C. (2017). SCnorm: robust normalization of single-cell RNA-seq data. *Nature Methods*, 14(6), 584–586. <https://doi.org/10.1038/nmeth.4263>

Baker, M., Mackenzie, I. R., Pickering-Brown, S. M., Gass, J., Rademakers, R., Lindholm, C., ... Hutton, M. (2006). Mutations in progranulin cause tau-negative frontotemporal dementia linked to chromosome 17. *Nature*, 442(7105), 916–919. <https://doi.org/10.1038/nature05016>

Bakken, T. E., Hodge, R. D., Miller, J. A., Yao, Z., Nguyen, T. N., Aevermann, B., ... Tasic, B. (2018). Single-nucleus and single-cell transcriptomes compared in matched cortical cell types. *PLOS ONE*, 13(12), e0209648. <https://doi.org/10.1371/journal.pone.0209648>

Balabanov, R., Washington, R., Wagnerova, J., & Dore-Duffy, P. (1996). CNS microvascular pericytes express macrophage-like function, cell surface integrin alpha M, and macrophage marker ED-2. *Microvascular Research*, 52(2), 127–142. <https://doi.org/10.1006/mvre.1996.0049>

Bandura, D. R., Baranov, V. I., Ornatsky, O. I., Antonov, A., Kinach, R., Lou, X., ... Tanner, S. D. (2009). Mass cytometry: technique for real time single cell multitarget immunoassay based on inductively coupled plasma time-of-flight mass spectrometry. *Analytical Chemistry*, 81(16), 6813–6822. <https://doi.org/10.1021/ac901049w>

Bankhead, P., Loughrey, M. B., Fernández, J. A., Dombrowski, Y., McArt, D. G., Dunne, P. D., ... Hamilton, P. W. (2017). QuPath: open source software for digital pathology image analysis. *Scientific Reports*, 7(1), 16878. <https://doi.org/10.1038/s41598-017-17204-5>

Baracska, K. L., Duchala, C. S., Miller, R. H., Macklin, W. B., & Trapp, B. D. (2002). Oligodendrogenesis is differentially regulated in gray and white matter of jimpy mice. *Journal of Neuroscience Research*, 70(5), 645–654. <https://doi.org/10.1002/jnr.10418>

Barnabé-Heider, F., Göritz, C., Sabelström, H., Takebayashi, H., Pfrieder, F. W., Meletis, K., & Frisén, J. (2010). Origin of new glial cells in intact and injured adult spinal cord. *Cell Stem Cell*, 7(4), 470–482. <https://doi.org/10.1016/j.stem.2010.07.014>

Batiuk, M. Y., Martirosyan, A., Wahis, J., de Vin, F., Marneffe, C., Kusserow, C., ... Holt, M. G. (2020). Identification of region-specific astrocyte subtypes at single cell resolution. *Nature Communications*, 11(1), 1220. <https://doi.org/10.1038/s41467-019-14198-8>

Beach, T. G., & McGeer, E. G. (1988). Lamina-specific arrangement of astrocytic gliosis and senile plaques in Alzheimer's disease visual cortex. *Brain Research*, 463, 357–361. [https://doi.org/10.1016/0006-8993\(88\)90410-6](https://doi.org/10.1016/0006-8993(88)90410-6)

Beauquis, J., Pavía, P., Pomilio, C., Vinuesa, A., Podlutskaya, N., Galvan, V., & Saravia, F. (2013).

Environmental enrichment prevents astroglial pathological changes in the hippocampus of APP transgenic mice, model of Alzheimer's disease. *Experimental Neurology*, 239, 28–37. <https://doi.org/10.1016/j.expneurol.2012.09.009>

Bechler, M. E., Byrne, L., & Ffrench-Constant, C. (2015). CNS myelin sheath lengths are an intrinsic property of oligodendrocytes. *Current Biology*, 25(18), 2411–2416. <https://doi.org/10.1016/j.cub.2015.07.056>

Bellik, L., Vinci, M. C., Filippi, S., Ledda, F., & Parenti, A. (2005). Intracellular pathways triggered by the selective FLT-1-agonist placental growth factor in vascular smooth muscle cells exposed to hypoxia. *British Journal of Pharmacology*, 146(4), 568–575. <https://doi.org/10.1038/sj.bjp.0706347>

Belloy, M. E., Napolioni, V., & Greicius, M. D. (2019). A quarter century of APOE and Alzheimer's disease: progress to date and the path forward. *Neuron*, 101(5), 820–838. <https://doi.org/10.1016/j.neuron.2019.01.056>

Ben-Zvi, A., Lacoste, B., Kur, E., Andreone, B. J., Maysnar, Y., Yan, H., & Gu, C. (2014). Mfsd2a is critical for the formation and function of the blood-brain barrier. *Nature*, 509(7501), 507–511. <https://doi.org/10.1038/nature13324>

Bennett, F. C., Bennett, M. L., Yaqoob, F., Mulinyawe, S. B., Grant, G. A., Hayden Gephart, M., ... Barres, B. A. (2018). A combination of ontogeny and CNS environment establishes microglial identity. *Neuron*, 98(6), 1170–1183. <https://doi.org/10.1016/j.neuron.2018.05.014>

Bennett, M. L., Bennett, F. C., Liddelow, S. A., Ajami, B., Zamanian, J. L., Fernhoff, N. B., ... Barres, B. A. (2016). New tools for studying microglia in the mouse and human CNS. *Proceedings of the National Academy of Sciences*, 113(12), E1738–46. <https://doi.org/10.1073/pnas.1525528113>

Benussi, L., Binetti, G., & Ghidoni, R. (2017). Loss of neuroprotective factors in neurodegenerative dementias: the end or the starting point? *Frontiers in Neuroscience*, 11(672). <https://doi.org/10.3389/FNINS.2017.00672>

Berghoff, S. A., Düking, T., Spieth, L., Winchenbach, J., Stumpf, S. K., Gerndt, N., ... Saher, G. (2017). Blood-brain barrier hyperpermeability precedes demyelination in the cuprizone model. *Acta Neuropathologica Communications*, 5(1), 94. <https://doi.org/10.1186/s40478-017-0497-6>

Bloom, G. S. (2014). Amyloid- β and tau. *JAMA Neurology*, 71(4), 505. <https://doi.org/10.1001/jamaneurol.2013.5847>

Borchelt, D. R., Ratovitski, T., van Lare, J., Lee, M. K., Gonzales, V., Jenkins, N. A., ... Sisodia, S. S. (1997). Accelerated amyloid deposition in the brains of transgenic mice coexpressing mutant presenilin 1 and amyloid precursor proteins. *Neuron*, 19(4), 939–945. [https://doi.org/10.1016/s0896-6273\(00\)80974-5](https://doi.org/10.1016/s0896-6273(00)80974-5)

Borggrewe, M., Grit, C., Vainchtein, I. D., Brouwer, N., Wesseling, E. M., Laman, J. D., ... Boddeke, E. W. G. M. (2021). Regionally diverse astrocyte subtypes and their heterogeneous response to EAE. *Glia*, 69(5), 1140–1154. <https://doi.org/10.1002/glia.23954>

Bossù, P., Salani, F., Alberici, A., Archetti, S., Bellelli, G., Galimberti, D., ... Borroni, B. (2011). Loss of function mutations in the progranulin gene are related to pro-inflammatory cytokine dysregulation in frontotemporal lobar degeneration patients. *Journal of Neuroinflammation*,

8, 65. <https://doi.org/10.1186/1742-2094-8-65>

Braak, H., & Braak, E. (1991). Neuropathological staging of Alzheimer-related changes. *Acta Neuropathologica*, 82(4), 239–259. <https://doi.org/10.1007/bf00308809>

Braak, Heiko, Alafuzoff, I., Arzberger, T., Kretschmar, H., & Del Tredici, K. (2006). Staging of Alzheimer disease-associated neurofibrillary pathology using paraffin sections and immunocytochemistry. *Acta Neuropathologica*, 112(4), 389–404. <https://doi.org/10.1007/s00401-006-0127-z>

Bribián, A., Medina-Rodríguez, E. M., Josa-Prado, F., García-Álvarez, I., Machín-Díaz, I., Esteban, P. F., ... de Castro, F. (2020). Functional heterogeneity of mouse and human brain OPCs: relevance for preclinical studies in multiple sclerosis. *Journal of Clinical Medicine*, 9(6). <https://doi.org/10.3390/jcm9061681>

Bright, F., Werry, E. L., Dobson-Stone, C., Piguet, O., Ittner, L. M., Halliday, G. M., ... Kril, J. J. (2019). Neuroinflammation in frontotemporal dementia. *Nature Reviews Neurology*, 15. <https://doi.org/10.1038/s41582-019-0231-z>

Bruinsma, I. B., Wilhelmus, M. M. M., Kox, M., Veerhuis, R., De Waal, R. M. W., & Verbeek, M. M. (2009). Apolipoprotein E protects cultured pericytes and astrocytes from D-A β 1-40-mediated cell death. *Brain Research*, 1315, 169–180. <https://doi.org/10.1016/j.brainres.2009.12.039>

Bundesen, L. Q., Scheel, T. A., Bregman, B. S., & Kromer, L. F. (2003). Ephrin-B2 and EphB2 regulation of astrocyte-meningeal fibroblast interactions in response to spinal cord lesions in adult rats. *The Journal of Neuroscience*, 23(21), 7789–7800. <https://doi.org/10.1523/JNEUROSCI.23-21-07789.2003>

Buniello, A., MacArthur, J. A. L., Cerezo, M., Harris, L. W., Hayhurst, J., Malangone, C., ... Parkinson, H. (2019). The NHGRI-EBI GWAS Catalog of published genome-wide association studies, targeted arrays and summary statistics 2019. *Nucleic Acids Research*, 47(Database issue), D1005. <https://doi.org/10.1093/NAR/GKY1120>

Burrell, J. R., Kiernan, M. C., Vucic, S., & Hodges, J. R. (2011). Motor neuron dysfunction in frontotemporal dementia. *Brain*, 134(Pt 9), 2582–2594. <https://doi.org/10.1093/brain/awr195>

Bushong, E. A., Martone, M. E., Jones, Y. Z., & Ellisman, M. H. (2002). Protoplasmic astrocytes in CA1 stratum radiatum occupy separate anatomical domains. *The Journal of Neuroscience*, 22(1), 183–192. <https://doi.org/10.1523/JNEUROSCI.22-01-00183.2002>

Bussolati, B., Grange, C., Bruno, S., Buttiglieri, S., Deregibus, M. C., Tei, L., ... Camussi, G. (2006). Neural-cell adhesion molecule (NCAM) expression by immature and tumor-derived endothelial cells favors cell organization into capillary-like structures. *Experimental Cell Research*, 312(6), 913–924. <https://doi.org/10.1016/j.yexcr.2005.12.004>

Butovsky, O., Jedrychowski, M. P., Moore, C. S., Cialic, R., Lanser, A. J., Gabriely, G., ... Weiner, H. L. (2014). Identification of a unique TGF- β -dependent molecular and functional signature in microglia. *Nature Neuroscience*, 17(1), 131–143. <https://doi.org/10.1038/nn.3599>

Butovsky, O., & Weiner, H. L. (2018). Microglial signatures and their role in health and disease. *Nature Reviews Neuroscience*, 19(10), 622–635. <https://doi.org/10.1038/s41583-018-0057-5>

Bystron, I., Blakemore, C., & Rakic, P. (2008). Development of the human cerebral cortex: Boulder Committee revisited. *Nature Reviews Neuroscience*, 9(2), 110–122. <https://doi.org/10.1038/nrn2208>

org/10.1038/nrn2252

Cahoy, J. D., Emery, B., Kaushal, A., Foo, L. C., Zamanian, J. L., Christopherson, K. S., ... Barres, B. A. (2008). A transcriptome database for astrocytes, neurons, and oligodendrocytes: a new resource for understanding brain development and function. *The Journal of Neuroscience*, 28(1), 264–278. <https://doi.org/10.1523/JNEUROSCI.4178-07.2008>

Calabresi, P. A., Fields, N. S., Maloni, H. W., Hanham, A., Carlino, J., Moore, J., ... Racke, M. K. (1998). Phase 1 trial of transforming growth factor beta 2 in chronic progressive MS. *Neurology*, 51(1), 289–292. <https://doi.org/10.1212/wnl.51.1.289>

Calzà, L., Giardino, L., Giuliani, A., Aloe, L., & Levi-Montalcini, R. (2000). Nerve growth factor control of neuronal expression of angiogenetic and vasoactive factors. *Proceedings of the National Academy of Sciences*, 98(7), 4160–4165. Retrieved from www.pnas.org. [orgcgidoi10.1073/pnas.051626998](https://doi.org/10.1073/pnas.051626998)

Cantarella, G., Lempereur, L., Presta, M., Ribatti, D., Lombardo, G., Lazarovici, P., ... Bernardini, R. (2002). Nerve growth factor-endothelial cell interaction leads to angiogenesis in vitro and in vivo. *FASEB Journal*, 16(10), 1307–1309. <https://doi.org/10.1096/fj.01-1000fje>

Cerbai, F., Lana, D., Nosi, D., Petkova-Kirova, P., & Zecchi, S. (2012). The neuron-astrocyte-microglia triad in normal brain ageing and in a model of neuroinflammation in the rat hippocampus. *PLOS ONE*, 7(9), 45250. <https://doi.org/10.1371/journal.pone.0045250>

Chakraborty, A., Kamermans, A., van het Hof, B., Castricum, K., Aanhane, E., van Horssen, J., ... de Vries, H. E. (2018). Angiopoietin like-4 as a novel vascular mediator in capillary cerebral amyloid angiopathy. *Brain*, 141(12), 3377–3388. <https://doi.org/10.1093/brain/awy274>

Chappell, J. C., Mouillesseaux, K. P., & Bautch, V. L. (2013). Flt-1 (vascular endothelial growth factor receptor-1) is essential for the vascular endothelial growth factor-Notch feedback loop during angiogenesis. *Arteriosclerosis, Thrombosis and Vascular Biology*, 33(8), 1952–1959. <https://doi.org/10.1161/ATVBAHA.113.301805>

Chen, A., Liao, S., Cheng, M., Ma, K., Wu, L., Lai, Y., ... Xu, X. (2021). Large field of view-spatially resolved transcriptomics at nanoscale resolution. *BioRxiv*, 2021.01.17.427004. <https://doi.org/10.1101/2021.01.17.427004>

Chen, P., Cai, W., Wang, L., & Deng, Q. (2008). A morphological and electrophysiological study on the postnatal development of oligodendrocyte precursor cells in the rat brain. *Brain Research*, 1243, 27–37. <https://doi.org/10.1016/J.BRAINRES.2008.09.029>

Chen, R., Wu, X., Jiang, L., & Zhang, Y. (2017). Single-cell RNA-seq reveals hypothalamic cell diversity. *Cell Reports*, 18(13), 3227–3241. <https://doi.org/10.1016/j.celrep.2017.03.004>

Chen, W.-T., Lu, A., Craessaerts, K., Pavie, B., Sala Frigerio, C., Corthout, N., ... De Strooper, B. (2020). Spatial Transcriptomics and In Situ Sequencing to Study Alzheimer's Disease. *Cell*, 182(4), 976–991. <https://doi.org/10.1016/j.cell.2020.06.038>

Chen, Y., Wu, H., Wang, S., Koito, H., Li, J., Ye, F., ... Lu, Q. R. (2009). The oligodendrocyte-specific G protein-coupled receptor GPR17 is a cell-intrinsic timer of myelination. *Nature Neuroscience*, 12(11), 1398–1406. <https://doi.org/10.1038/nn.2410>

Chew, J., Gendron, T. F., Prudencio, M., Sasaguri, H., Zhang, Y.-J., Castanedes-Casey, M., ... Petrucelli, L. (2015). C9ORF72 repeat expansions in mice cause TDP-43 pathology, neuronal loss, and behavioral deficits. *Science*, 348(6239), 1151–1154. <https://doi.org/10.1126/science>

aaa9344

- Chitramuthu, B. P., Bennett, H. P. J., & Bateman, A. (2017). Progranulin: a new avenue towards the understanding and treatment of neurodegenerative disease. *Brain*, 140(12), 3081–3104. <https://doi.org/10.1093/brain/awx198>
- Chittajallu, R., Aguirre, A., & Gallo, V. (2004). NG2-positive cells in the mouse white and grey matter display distinct physiological properties. *The Journal of Physiology*, 561(1), 109–122. <https://doi.org/10.1113/jphysiol.2004.074252>
- Chiu, I. M., Morimoto, E. T. A., Goodarzi, H., Liao, J. T., O’Keeffe, S., Phatnani, H. P., ... Maniatis, T. (2013). A neurodegeneration-specific gene-expression signature of acutely isolated microglia from an amyotrophic lateral sclerosis mouse model. *Cell Reports*, 4(2), 385–401. <https://doi.org/10.1016/j.celrep.2013.06.018>
- Cirrito, J. R., Deane, R., Fagan, A. M., Spinner, M. L., Parsadanian, M., Finn, M. B., ... Holtzman, D. M. (2005). P-glycoprotein deficiency at the blood-brain barrier increases amyloid-beta deposition in an Alzheimer disease mouse model. *The Journal of Clinical Investigation*, 115(11), 3285–3290. <https://doi.org/10.1172/JCI25247>
- Compston, A., & Coles, A. (2008). Multiple sclerosis. *The Lancet*, 372(9648), 1502–1517. [https://doi.org/10.1016/S0140-6736\(08\)61620-7](https://doi.org/10.1016/S0140-6736(08)61620-7)
- Correale, J., Gaitán, M. I., Ysrraelit, M. C., & Fiol, M. P. (2017). Progressive multiple sclerosis: from pathogenic mechanisms to treatment. *Brain*, 140(3), 527–546. <https://doi.org/10.1093/brain/aww258>
- Crisan, M., Yap, S., Casteilla, L., Chen, C.-W., Corselli, M., Park, T. S., ... Péault, B. (2008). A perivascular origin for mesenchymal stem cells in multiple human organs. *Cell Stem Cell*, 3(3), 301–313. <https://doi.org/10.1016/j.stem.2008.07.003>
- Cruets, M., Gijssels, I., van der Zee, J., Engelborghs, S., Wils, H., Pirici, D., ... Van Broeckhoven, C. (2006). Null mutations in progranulin cause ubiquitin-positive frontotemporal dementia linked to chromosome 17q21. *Nature*, 442(7105), 920–924. <https://doi.org/10.1038/nature05017>
- Cummings, J. L., Morstorf, T., & Zhong, K. (2014). Alzheimer’s disease drug-development pipeline: few candidates, frequent failures. *Alzheimer’s Research & Therapy*, 6(4), 37. <https://doi.org/10.1186/alzrt269>
- Daneman, R., Zhou, L., Kebede, A. A., & Barres, B. A. (2010). Pericytes are required for blood-brain barrier integrity during embryogenesis. *Nature*, 468(7323), 562–566. <https://doi.org/10.1038/nature09513>
- Darmanis, S., Sloan, S. A., Zhang, Y., Enge, M., Caneda, C., Shuer, L. M., ... Quake, S. R. (2015). A survey of human brain transcriptome diversity at the single cell level. *Proceedings of the National Academy of Sciences*, 112(23), 7285–7290. <https://doi.org/10.1073/pnas.1507125112>
- Davis, A., Gao, R., & Navin, N. E. (2019). SCOPIT: sample size calculations for single-cell sequencing experiments. *BMC Bioinformatics*, 20(1), 566. <https://doi.org/10.1186/s12859-019-3167-9>
- De Biase, L. M., Schuebel, K. E., Fufeld, Z. H., Jair, K., Hawes, I. A., Cimbrow, R., ... Bonci, A. (2017). Local cues establish and maintain region-specific phenotypes of basal ganglia microglia. *Neuron*, 95(2), 341–356. <https://doi.org/10.1016/j.neuron.2017.06.020>

- De Carlos, J. A., & Borrell, J. (2007). A historical reflection of the contributions of Cajal and Golgi to the foundations of neuroscience. *Brain Research Reviews*, 55(1), 8–16. <https://doi.org/10.1016/j.brainresrev.2007.03.010>
- De Falco, S. (2012). The discovery of placenta growth factor and its biological activity. *Experimental and Molecular Medicine*, 44(1), 1–9. <https://doi.org/10.3858/emm.2012.44.1.025>
- De Strooper, B., & Karran, E. (2016). Leading edge: the cellular phase of Alzheimer's disease. *Cell*, 164(4), 603–615. <https://doi.org/10.1016/j.cell.2015.12.056>
- Derada Troletti, C., de Goede, P., Kamermans, A., & de Vries, H. E. (2016). Molecular alterations of the blood-brain barrier under inflammatory conditions: The role of endothelial to mesenchymal transition. *Biochimica et Biophysica Acta*, 1862(3), 452–460. <https://doi.org/10.1016/j.bbadis.2015.10.010>
- Derada Troletti, C., Fontijn, R. D., Gowing, E., Charabati, M., van Het Hof, B., Didouh, I., ... de Vries, H. E. (2019). Inflammation-induced endothelial to mesenchymal transition promotes brain endothelial cell dysfunction and occurs during multiple sclerosis pathophysiology. *Cell Death & Disease*, 10(2), 45. <https://doi.org/10.1038/s41419-018-1294-2>
- Dias, D. O., Kalkitsas, J., 1#, Y. K., Estrada, C. P., Tatarishvili, J., Ernst, A., ... Göritz, C. (2021). Pericyte-derived fibrotic scarring is conserved across diverse central nervous system lesions. *Nature Communications*, 12, 5501. <https://doi.org/10.1101/2020.04.30.068965>
- Diniz, D. G., Foro, C. A. R., Rego, C. M. D., Gloria, D. A., De Oliveira, F. R. R., Paes, J. M. P., ... Diniz, C. W. P. (2010). Environmental impoverishment and aging alter object recognition, spatial learning, and dentate gyrus astrocytes. *European Journal of Neuroscience*, 32(3), 509–519. <https://doi.org/10.1111/j.1460-9568.2010.07296.x>
- Djebali, S., Davis, C. A., Merkel, A., Dobin, A., Lassmann, T., Mortazavi, A., ... Gingeras, T. R. (2012). Landscape of transcription in human cells. *Nature*, 489(7414), 101–108. <https://doi.org/10.1038/nature11233>
- Dobin, A., & Gingeras, T. R. (2015). Mapping RNA-seq Reads with STAR. *Current Protocols in Bioinformatics*, 51, 11.14.1. <https://doi.org/10.1002/0471250953.BI1114S51>
- Dong, Y., & Benveniste, E. N. (2001). Immune function of astrocytes. *Glia*, 36(2), 180–190. <https://doi.org/10.1002/glia.1107>
- Dorrier, C. E., Aran, D., Haenelt, E. A., Sheehy, R. N., Hoi, K. K., Pintarić, L., ... Daneman, R. (2021). CNS fibroblasts form a fibrotic scar in response to immune cell infiltration. *Nature Neuroscience*, 24(2), 234–244. <https://doi.org/10.1038/s41593-020-00770-9>
- Dougherty, K. D., Dreyfus, C. F., & Black, I. B. (2000). Brain-derived neurotrophic factor in astrocytes, oligodendrocytes, and microglia/macrophages after spinal cord injury. *Neurobiology of Disease*, 7(6 Pt B), 574–585. <https://doi.org/10.1006/nbdi.2000.0318>
- Doyle, J. P., Dougherty, J. D., Heiman, M., Schmidt, E. F., Stevens, T. R., Ma, G., ... Heintz, N. (2008). Application of a translational profiling approach for the comparative analysis of CNS cell types. *Cell*, 135(4), 749–762. <https://doi.org/10.1016/j.cell.2008.10.029>
- Drejer, J., Larsson, O. M., & Schousboe, A. (1982). Characterization of L-glutamate uptake into and release from astrocytes and neurons cultured from different brain regions. *Experimental Brain Research*, 47(2), 259–269. <https://doi.org/10.1007/BF00239385>

- Du, Y., & Dreyfus, C. F. (2002). Oligodendrocytes as providers of growth factors. *Journal of Neuroscience Research*, 68(6), 647–654. <https://doi.org/10.1002/jnr.10245>
- Duffield, J. S., Forbes, S. J., Constandinou, C. M., Clay, S., Partolina, M., Vuthoori, S., ... Iredale, J. P. (2005). Selective depletion of macrophages reveals distinct, opposing roles during liver injury and repair. *The Journal of Clinical Investigation*, 115(1), 56–65. <https://doi.org/10.1172/JCI22675>
- Duncan, G. J., Manesh, S. B., Hilton, B. J., Assinck, P., Plemel, J. R., & Tetzlaff, W. (2020). The fate and function of oligodendrocyte progenitor cells after traumatic spinal cord injury. *Glia*, 68(2), 227–245. <https://doi.org/10.1002/glia.23706>
- Duò, A., Robinson, M. D., & Sonesson, C. (2018). A systematic performance evaluation of clustering methods for single-cell RNA-seq data. *F1000Research*, 7, 1141. <https://doi.org/10.12688/f1000research.15666.3>
- Eguchi, R., Nakano, T., & Wakabayashi, I. (2017). Progranulin and granulin-like protein as novel VEGF-independent angiogenic factors derived from human mesothelioma cells. *Oncogene*, 36(5), 714–722. <https://doi.org/10.1038/onc.2016.226>
- Eilken, H. M., Diéguez-Hurtado, R., Schmidt, I., Nakayama, M., Jeong, H.-W., Arf, H., ... Adams, R. H. (2017). Pericytes regulate VEGF-induced endothelial sprouting through VEGFR1. *Nature Communications*, 8(1), 1574. <https://doi.org/10.1038/s41467-017-01738-3>
- Ek Olofsson, H., & Englund, E. (2019). A cortical microvascular structure in vascular dementia, Alzheimer's disease, frontotemporal lobar degeneration and nondemented controls: a sign of angiogenesis due to brain ischaemia? *Neuropathology and Applied Neurobiology*, 45(6), 557–569. <https://doi.org/10.1111/nan.12552>
- Emrich, S. J., Barbazuk, W. B., Li, L., & Schnable, P. S. (2007). Gene discovery and annotation using LCM-454 transcriptome sequencing. *Genome Research*, 17(1), 69–73. <https://doi.org/10.1101/gr.5145806>
- Emsley, J. G., & Macklis, J. D. (2006). Astroglial heterogeneity closely reflects the neuronal-defined anatomy of the adult murine CNS. *Neuron Glia Biology*, 2(3), 175–186. <https://doi.org/10.1017/S1740925X06000202>
- Engelhardt, B., & Ransohoff, R. M. (2012). Capture, crawl, cross: the T cell code to breach the blood-brain barriers. *Trends in Immunology*, 33(12), 579–589. <https://doi.org/10.1016/j.it.2012.07.004>
- Erickson, C. A., & Barnes, C. A. (2003). The neurobiology of memory changes in normal aging. *Experimental Gerontology*, 38(1–2), 61–69. [https://doi.org/10.1016/s0531-5565\(02\)00160-2](https://doi.org/10.1016/s0531-5565(02)00160-2)
- Erickson, M. A., & Banks, W. A. (2013). Blood-brain barrier dysfunction as a cause and consequence of Alzheimer's disease. *Journal of Cerebral Blood Flow and Metabolism*, 33(10), 1500–1513. <https://doi.org/10.1038/jcbfm.2013.135>
- Eriksen, J. L., & Mackenzie, I. R. A. (2008). Progranulin: normal function and role in neurodegeneration. *Journal of Neurochemistry*, 104(2), 287–297. <https://doi.org/10.1111/j.1471-4159.2007.04968.x>
- Eriksson, A., Cao, R., Pawliuk, R., Berg, S. M., Tsang, M., Zhou, D., ... Cao, Y. (2002). Placenta growth factor-1 antagonizes VEGF-induced angiogenesis and tumor growth by the formation of functionally inactive PlGF-1/VEGF heterodimers. *Cancer Cell*, 1(1), 99–108. [https://doi.org/10.1016/S1535-6175\(02\)00002-0](https://doi.org/10.1016/S1535-6175(02)00002-0)

org/10.1016/s1535-6108(02)00028-4

Fabricius, K., Jacobsen, J. S., & Pakkenberg, B. (2013). Effect of age on neocortical brain cells in 90 year old human females—a cell counting study. *Neurobiology of Aging*, 34(1), 91–99. <https://doi.org/10.1016/j.neurobiolaging.2012.06.009>

Falcão, A. M., van Bruggen, D., Marques, S., Meijer, M., Jäkel, S., Agirre, E., ... Castelo-Branco, G. (2018). Disease-specific oligodendrocyte lineage cells arise in multiple sclerosis. *Nature Medicine*, 24(12), 1837–1844. <https://doi.org/10.1038/s41591-018-0236-y>

Fernández-Klett, F., Potas, J. R., Hilpert, D., Blazej, K., Radke, J., Huck, J., ... Priller, J. (2013). Early loss of pericytes and perivascular stromal cell-induced scar formation after stroke. *Journal of Cerebral Blood Flow and Metabolism*, 33(3), 428–439. <https://doi.org/10.1038/jcbfm.2012.187>

Fernández-Klett, F., & Priller, J. (2014). The fibrotic scar in neurological disorders. *Brain Pathology*, 24(4), 404–413. <https://doi.org/10.1111/bpa.12162>

Fernandez-Klett, F., Brandt, L., Fernández-Zapata, C., Abuelnor, B., Middeldorp, J., Sluijs, J. A., ... Priller, J. (2020). Denser brain capillary network with preserved pericytes in Alzheimer's disease. *Brain Pathology*, 30(6), 1071–1086. <https://doi.org/10.1111/bpa.12897>

Filiano, A. J., Martens, L. H., Young, A. H., Warmus, B. A., Zhou, P., Diaz-Ramirez, G., ... Roberson, E. D. (2013). Dissociation of frontotemporal dementia-related deficits and neuroinflammation in progranulin haploinsufficient mice. *The Journal of Neuroscience*, 33(12), 5352–5361. <https://doi.org/10.1523/JNEUROSCI.6103-11.2013>

Finak, G., McDavid, A., Yajima, M., Deng, J., Gersuk, V., Shalek, A. K., ... Gottardo, R. (2015). MAST: a flexible statistical framework for assessing transcriptional changes and characterizing heterogeneity in single-cell RNA sequencing data. *Genome Biology*, 16, 278. <https://doi.org/10.1186/S13059-015-0844-5>

Frade, J. M., & Barde, Y. A. (1998). Microglia-derived nerve growth factor causes cell death in the developing retina. *Neuron*, 20(1), 35–41. [https://doi.org/10.1016/s0896-6273\(00\)80432-8](https://doi.org/10.1016/s0896-6273(00)80432-8)

Franklin, R. J. M., & Ffrench-Constant, C. (2008). Remyelination in the CNS: from biology to therapy. *Nature Reviews Neuroscience*, 9(11), 839–855. <https://doi.org/10.1038/nrn2480>

Friedman, B. A., Srinivasan, K., Ayalon, G., Meilandt, W. J., Lin, H., Huntley, M. A., ... Hansen, D. V. (2018). Diverse brain myeloid expression profiles reveal distinct microglial activation states and aspects of Alzheimer's disease not evident in mouse models. *Cell Reports*, 22(3), 832–847. <https://doi.org/10.1016/j.celrep.2017.12.066>

Füchtbauer, L., Groth-Rasmussen, M., Holm, T. H., Løbner, M., Toft-Hansen, H., Khoroshii, R., & Owens, T. (2010). Angiotensin II Type 1 receptor (AT1) signaling in astrocytes regulates synaptic degeneration-induced leukocyte entry to the central nervous system. *Brain, Behavior, and Immunity*, 25(5), 897–904. <https://doi.org/10.1016/j.bbi.2010.09.015>

Gacche, R. N. (2015). Compensatory angiogenesis and tumor refractoriness. *Oncogenesis*, 4(6), e153–e153. <https://doi.org/10.1038/oncsis.2015.14>

Galatro, T. F., Holtman, I. R., Lerario, A. M., Vainchtein, I. D., Brouwer, N., Sola, P. R., ... Eggen, B. J. L. (2017a). Transcriptomic analysis of purified human cortical microglia reveals age-associated changes. *Nature Neuroscience*, 20(8), 1162–1171. <https://doi.org/10.1038/nn.4597>

- Galatro, T. F., Vainchtein, I. D., Brouwer, N., Boddeke, E. W. G. M., & Eggen, B. J. L. (2017b). Isolation of microglia and immune infiltrates from mouse and primate central nervous system. In *Methods in Molecular Biology* (Vol. 1559, pp. 333–342). https://doi.org/10.1007/978-1-4939-6786-5_23
- Galimberti, D., Bonsi, R., Fenoglio, C., Serpente, M., Cioffi, S. M. G., Fumagalli, G., ... Scarpini, E. (2015). Inflammatory molecules in Frontotemporal Dementia: cerebrospinal fluid signature of progranulin mutation carriers. *Brain, Behavior, and Immunity*, 49, 182–187. <https://doi.org/10.1016/j.bbi.2015.05.006>
- Gautier, E. L., Shay, T., Miller, J., Greter, M., Jakubzick, C., Ivanov, S., ... Randolph, G. J. (2012). Gene-expression profiles and transcriptional regulatory pathways that underlie the identity and diversity of mouse tissue macrophages. *Nature Immunology*, 13(11), 1118–1128. <https://doi.org/10.1038/ni.2419>
- Geinisman, Y., Ganeshina, O., Yoshida, R., Berry, R. W., Disterhoft, J. F., & Gallagher, M. (2004). Aging, spatial learning, and total synapse number in the rat CA1 stratum radiatum. *Neurobiology of Aging*, 25, 407–416. <https://doi.org/10.1016/j.neurobiolaging.2003.12.001>
- Geirsdottir, L., David, E., Keren-Shaul, H., Weiner, A., Bohlen, S. C., Neuber, J., ... Prinz, M. (2019). Cross-Species Single-Cell Analysis Reveals Divergence of the Primate Microglia Program. *Cell*, 179(7), 1609–1622.e16. <https://doi.org/10.1016/j.cell.2019.11.010>
- George, N. I., & Chang, C.-W. (2014). DAFS: a data-adaptive flag method for RNA-sequencing data to differentiate genes with low and high expression. *BMC Bioinformatics*, 15(1), 92. <https://doi.org/10.1186/1471-2105-15-92>
- Gerrits, E., Brouwer, N., Kooistra, S. M., Woodbury, M. E., Vermeiren, Y., Lambourne, M., ... Boddeke, E. W. G. M. (2021). Distinct amyloid- β and tau-associated microglia profiles in Alzheimer's disease. *Acta Neuropathologica*, 141(5), 1–16. <https://doi.org/10.1007/s00401-021-02263-w>
- Gerrits, E., Heng, Y., Boddeke, E. W. G. M., & Eggen, B. J. L. (2020). Transcriptional profiling of microglia; current state of the art and future perspectives. *Glia*, 68(4), 740–755. <https://doi.org/10.1002/glia.23767>
- Giannini, L. A. A., Xie, S. X., McMillan, C. T., Liang, M., Williams, A., Jester, C., ... Irwin, D. J. (2019a). Divergent patterns of TDP-43 and tau pathologies in primary progressive aphasia. *Annals of Neurology*, 85(5), 630–643. <https://doi.org/10.1002/ana.25465>
- Giannini, L. A. A., Xie, S. X., Peterson, C., Zhou, C., Lee, E. B., Wolk, D. A., ... Irwin, D. J. (2019b). Empiric methods to account for pre-analytical variability in digital histopathology in frontotemporal lobar degeneration. *Frontiers in Neuroscience*, 13, 682. <https://doi.org/10.3389/fnins.2019.00682>
- Ginhoux, F., Greter, M., Leboeuf, M., Nandi, S., See, P., Gokhan, S., ... Merad, M. (2010). Fate mapping analysis reveals that adult microglia derive from primitive macrophages. *Science*, 330(6005), 841. <https://doi.org/10.1126/SCIENCE.1194637>
- Göritz, C., Dias, D. O., Tomilin, N., Barbacid, M., Shupliakov, O., & Frisén, J. (2011). A pericyte origin of spinal cord scar tissue. *Science*, 333(6039), 238–242. <https://doi.org/10.1126/science.1203165>
- Gosselin, D., Link, V. M., Romanoski, C. E., Fonseca, G. J., Eichenfield, D. Z., Spann, N. J.,

- ... Glass, C. K. (2014). Environment drives selection and function of enhancers controlling tissue-specific macrophage identities. *Cell*, 159(6), 1327–1340. <https://doi.org/10.1016/j.cell.2014.11.023>
- Gosselin, D., Skola, D., Coufal, N. G., Holtman, I. R., Schlachetzki, J. C. M., Sajti, E., ... Glass, C. K. (2017). An environment-dependent transcriptional network specifies human microglia identity. *Science*, 356(6344), eaal3222. <https://doi.org/10.1126/science.aal3222>
- Götz, J., Chen, F., Barmettler, R., & Nitsch, R. M. (2001). Tau filament formation in transgenic mice expressing P301L tau. *The Journal of Biological Chemistry*, 276(1), 529–534. <https://doi.org/10.1074/jbc.M006531200>
- Grabert, K., Michoel, T., Karavolos, M. H., Clohisey, S., Baillie, J. K., Stevens, M. P., ... McColl, B. W. (2016). Microglial brain region–dependent diversity and selective regional sensitivities to aging. *Nature Neuroscience*, 19(3), 504–516. <https://doi.org/10.1038/nn.4222>
- Graumann, U., Reynolds, R., Steck, A. J., & Schaeren-Wiemers, N. (2003). Molecular changes in normal appearing white matter in multiple sclerosis are characteristic of neuroprotective mechanisms against hypoxic insult. *Brain Pathology*, 13(4), 554–573. <https://doi.org/10.1111/j.1750-3639.2003.tb00485.x>
- Greaves, C. V., & Rohrer, J. D. (2019). An update on genetic frontotemporal dementia. *Journal of Neurology*, 266(8), 2075–2086. <https://doi.org/10.1007/s00415-019-09363-4>
- Greter, M., Lelios, I., & Croxford, A. L. (2015). Microglia versus myeloid cell nomenclature during brain inflammation. *Frontiers in Immunology*, 6, 249. <https://doi.org/10.3389/fimmu.2015.00249>
- Griffin, W. S., Stanley, L. C., Ling, C., White, L., MacLeod, V., Perrot, L. J., ... Araoz, C. (1989). Brain interleukin 1 and S-100 immunoreactivity are elevated in down syndrome and Alzheimer disease. *Proceedings of the National Academy of Sciences*, 86(19), 7611–7615. <https://doi.org/10.1073/pnas.86.19.7611>
- Grubman, A., Chew, G., Ouyang, J. F., Sun, G., Choo, X. Y., McLean, C., ... Polo, J. M. (2019). A single-cell atlas of entorhinal cortex from individuals with Alzheimer’s disease reveals cell-type-specific gene expression regulation. *Nature Neuroscience*, 22, 1–11. <https://doi.org/10.1038/s41593-019-0539-4>
- Guneykaya, D., Ivanov, A., Hernandez, D. P., Haage, V., Wojtas, B., Meyer, N., ... Wolf, S. A. (2018). Transcriptional and translational differences of microglia from male and female brains. *Cell Reports*, 24(10), 2773–2783. <https://doi.org/10.1016/j.celrep.2018.08.001>
- Guttmann, C. R., Rousset, M., Roch, J. A., Hannoun, S., Durand-Dubief, F., Belaroussi, B., ... Cotton, F. (2016). Multiple sclerosis lesion formation and early evolution revisited: a weekly high-resolution magnetic resonance imaging study. *Multiple Sclerosis*, 22(6), 761–769. <https://doi.org/10.1177/1352458515600247>
- Haarmann, A., Schuhmann, M. K., Silwedel, C., Monoranu, C.-M., Stoll, G., & Buttmann, M. (2019). Human brain endothelial CXCR2 is inflammation-inducible and mediates CXCL5- and CXCL8-triggered paraendothelial barrier breakdown. *International Journal of Molecular Sciences*, 20(3). <https://doi.org/10.3390/ijms20030602>
- Habib, N., Li, Y., Heidenreich, M., Swiech, L., Avraham-Davidi, I., Trombetta, J. J., ... Regev, A. (2016). Div-Seq: single-nucleus RNA-Seq reveals dynamics of rare adult newborn neurons.

- Science*, 353(6302), 925–928. <https://doi.org/10.1126/science.aad7038>
- Habib, N., McCabe, C., Medina, S., Varshavsky, M., Kitsberg, D., Dvir-Szternfeld, R., ... Schwartz, M. (2020). Disease-associated astrocytes in Alzheimer's disease and aging. *Nature Neuroscience*, 23(6), 701–706. <https://doi.org/10.1038/s41593-020-0624-8>
- Hafemeister, C. (2019). How Many Cells | Satija Lab. Retrieved September 22, 2021, from <https://satijalab.org/howmanycells/>
- Hafemeister, C., & Satija, R. (2019). Normalization and variance stabilization of single-cell RNA-seq data using regularized negative binomial regression. *Genome Biology*, 20(1), 296. <https://doi.org/10.1186/s13059-019-1874-1>
- Halliday, M. R., Rege, S. V., Ma, Q., Zhao, Z., Miller, C. A., Winkler, E. A., & Zlokovic, B. V. (2016). Accelerated pericyte degeneration and blood-brain barrier breakdown in apolipoprotein E4 carriers with Alzheimer's disease. *Journal of Cerebral Blood Flow and Metabolism*, 36(1), 216–227. <https://doi.org/10.1038/jcbfm.2015.44>
- Hallmann, A.-L., Araúzo-Bravo, M. J., Mavrommatis, L., Ehrlich, M., Röpke, A., Brockhaus, J., ... Hargus, G. (2017). Astrocyte pathology in a human neural stem cell model of frontotemporal dementia caused by mutant TAU protein. *Scientific Reports*, 7(1), 42991. <https://doi.org/10.1038/srep42991>
- Hammond, T. R., Dufort, C., Dissing-Olesen, L., Giera, S., Young, A., Wysoker, A., ... Stevens, B. (2019). Single-cell RNA sequencing of microglia throughout the mouse lifespan and in the injured brain reveals complex cell-state changes. *Immunity*, 50(1), 253–271. <https://doi.org/10.1016/j.immuni.2018.11.004>
- Hansson, O., Santillo, A. F., Meeter, L. H., Nilsson, K., Landqvist Waldö, M., Nilsson, C., ... Janelidze, S. (2019). CSF placental growth factor - a novel candidate biomarker of frontotemporal dementia. *Annals of Clinical and Translational Neurology*, 6(5), 863–872. <https://doi.org/10.1002/acn3.763>
- Hasel, P., Rose, I. V. L., Sadick, J. S., Kim, R. D., & Liddelow, S. A. (2021). Neuroinflammatory astrocyte subtypes in the mouse brain. *Nature Neuroscience*, 24(10), 1475–1487. <https://doi.org/10.1038/s41593-021-00905-6>
- Haug, H., & Eggers, R. (1991). Morphometry of the human cortex cerebri and corpus striatum during aging. *Neurobiology of Aging*, 12, 336–338. [https://doi.org/10.1016/0197-4580\(91\)90013-a](https://doi.org/10.1016/0197-4580(91)90013-a)
- Hawkes, C. A., Jayakody, N., Johnston, D. A., Bechmann, I., & Carare, R. O. (2014). Failure of perivascular drainage of β -amyloid in cerebral amyloid angiopathy. *Brain Pathology*, 24(4), 396–403. <https://doi.org/10.1111/bpa.12159>
- Hebenstreit, D. (2012). Methods, challenges and potentials of single cell RNA-seq. *Biology*, 1(3), 658–667. <https://doi.org/10.3390/biology1030658>
- Hecht, M., Krämer, L. M., von Arnim, C. A. F., Otto, M., & Thal, D. R. (2018). Capillary cerebral amyloid angiopathy in Alzheimer's disease: association with allocortical/hippocampal microinfarcts and cognitive decline. *Acta Neuropathologica*, 135(5), 681–694. <https://doi.org/10.1007/s00401-018-1834-y>
- Heimberg, G., Bhatnagar, R., El-Samad, H., & Thomson, M. (2016). Low dimensionality in gene expression data enables the accurate extraction of transcriptional programs from shallow

- sequencing. *Cell Systems*, 2(4), 239–250. <https://doi.org/10.1016/j.cels.2016.04.001>
- Heller, C., Foiani, M. S., Moore, K., Convery, R., Bocchetta, M., Neason, M., ... GENFI. (2020). Plasma glial fibrillary acidic protein is raised in progranulin-associated frontotemporal dementia. *Journal of Neurology, Neurosurgery, and Psychiatry*, 91(3), 263–270. <https://doi.org/10.1136/jnnp-2019-321954>
- Heneka, M. T., Sastre, M., Dumitrescu-Ozimek, L., Dewachter, I., Walter, J., Klockgether, T., & Van Leuven, F. (2005). Focal glial activation coincides with increased BACE1 activation and precedes amyloid plaque deposition in APP[V717I] transgenic mice. *Journal of Neuroinflammation*, 2(1), 22. <https://doi.org/10.1186/1742-2094-2-22>
- Heng, Y., Dubbelaar, M. L., Marie, S. K. N., Boddeke, E. W. G. M., & Eggen, B. J. L. (2021). The effects of postmortem delay on mouse and human microglia gene expression. *Glia*, 69(4), 1053–1060. <https://doi.org/10.1002/glia.23948>
- Hickman, S. E., Kingery, N. D., Ohsumi, T. K., Borowsky, M. L., Wang, L., Means, T. K., & El Khoury, J. (2013). The microglial sensome revealed by direct RNA sequencing. *Nature Neuroscience*, 16(12), 1896–1905. <https://doi.org/10.1038/nn.3554>
- Hill, R. A., Tong, L., Yuan, P., Murikinati, S., Gupta, S., & Grutzendler, J. (2015). Regional blood flow in the normal and ischemic brain is controlled by arteriolar smooth muscle cell contractility and not by capillary pericytes. *Neuron*, 87(1), 95–110. <https://doi.org/10.1016/j.neuron.2015.06.001>
- Hladky, S. B., & Barrand, M. A. (2014). Mechanisms of fluid movement into, through and out of the brain: evaluation of the evidence. *Fluids and Barriers of the CNS*, 11(1), 26. <https://doi.org/10.1186/2045-8118-11-26>
- Hodge, R. D., Bakken, T. E., Miller, J. A., Smith, K. A., Barkan, E. R., Graybuck, L. T., ... Lein, E. S. (2019). Conserved cell types with divergent features in human versus mouse cortex. *Nature*, 573(7772), 61–68. <https://doi.org/10.1038/s41586-019-1506-7>
- Hofman, M. A. (2014). Evolution of the human brain: when bigger is better. *Frontiers in Neuroanatomy*, 8, 15. <https://doi.org/10.3389/fnana.2014.00015>
- Hol, E. M., & Pekny, M. (2015). Glial fibrillary acidic protein (GFAP) and the astrocyte intermediate filament system in diseases of the central nervous system. *Current Opinion in Cell Biology*, 32, 121–130. <https://doi.org/10.1016/j.CEB.2015.02.004>
- Holtman, I. R., Raj, D. D., Miller, J. A., Schaafsma, W., Yin, Z., Brouwer, N., ... Eggen, B. J. L. (2015). Induction of a common microglia gene expression signature by aging and neurodegenerative conditions: a co-expression meta-analysis. *Acta Neuropathologica Communications*, 3(1), 31. <https://doi.org/10.1186/s40478-015-0203-5>
- Holtzman, D. M., Bales, K. R., Tenkova, T., Fagan, A. M., Parsadanian, M., Sartorius, L. J., ... Paul, S. M. (2000). Apolipoprotein E isoform-dependent amyloid deposition and neuritic degeneration in a mouse model of Alzheimer's disease. *Proceedings of the National Academy of Sciences*, 97(6), 2892–2897. <https://doi.org/10.1073/pnas.050004797>
- Hong, S., Beja-Glasser, V. F., Nfonoyim, B. M., Frouin, A., Li, S., Ramakrishnan, S., ... Stevens, B. (2016). Complement and microglia mediate early synapse loss in Alzheimer mouse models. *Science*, 352(6286), 712–716. <https://doi.org/10.1126/science.aad8373>

- Hong, Y., Yan, W., Chen, S., Sun, C., & Zhang, J. (2010). The role of Nrf2 signaling in the regulation of antioxidants and detoxifying enzymes after traumatic brain injury in rats and mice. *Acta Pharmacologica Sinica*, 31(11), 1421–1430. <https://doi.org/10.1038/aps.2010.101>
- Hou, S. T., Nilchi, L., Li, X., Gangaraju, S., Jiang, S. X., Aylsworth, A., ... Slinn, J. (2015). Semaphorin3A elevates vascular permeability and contributes to cerebral ischemia-induced brain damage. *Scientific Reports*, 5(1), 7890. <https://doi.org/10.1038/srep07890>
- Hu, P., Fabyanic, E., Kwon, D. Y., Tang, S., Zhou, Z., & Wu, H. (2017). Dissecting cell-type composition and activity-dependent transcriptional state in mammalian brains by massively parallel single-nucleus RNA-seq. *Molecular Cell*, 68(5), 1006–1015. <https://doi.org/10.1016/j.MOLCEL.2017.11.017>
- Hughes, E. G., Orthmann-Murphy, J. L., Langseth, A. J., & Bergles, D. E. (2018). Myelin remodeling through experience-dependent oligodendrogenesis in the adult somatosensory cortex. *Nature Neuroscience*, 21(5), 696–706. <https://doi.org/10.1038/s41593-018-0121-5>
- Hyman, S. E. (2005). Neurotransmitters. *Current Biology*, 15(5), R154-8. <https://doi.org/10.1016/j.cub.2005.02.037>
- Iliff, J. J., Wang, M., Liao, Y., Plogg, B. A., Peng, W., Gundersen, G. A., ... Nedergaard, M. (2012). A paravascular pathway facilitates CSF flow through the brain parenchyma and the clearance of interstitial solutes, including amyloid β . *Science Translational Medicine*, 4(147), 147ra111. <https://doi.org/10.1126/scitranslmed.3003748>
- International Multiple Sclerosis Genetics Consortium, I. M. S. G. (2019). Multiple sclerosis genomic map implicates peripheral immune cells and microglia in susceptibility. *Science*, 365(6460). <https://doi.org/10.1126/science.aav7188>
- Irwin, D. J., Byrne, M. D., McMillan, C. T., Cooper, F., Arnold, S. E., Lee, E. B., ... Trojanowski, J. Q. (2016). Semi-automated digital image analysis of Pick's disease and TDP-43 proteinopathy. *The Journal of Histochemistry and Cytochemistry*, 64(1), 54–66. <https://doi.org/10.1369/0022155415614303>
- Irwin, D. J., McMillan, C. T., Xie, S. X., Rascovsky, K., Van Deerlin, V. M., Coslett, H. B., ... Grossman, M. (2018). Asymmetry of post-mortem neuropathology in behavioural-variant frontotemporal dementia. *Brain*, 141(1), 288–301. <https://doi.org/10.1093/brain/awx319>
- Ishii, Y., Oya, T., Zheng, L., Gao, Z., Kawaguchi, M., Sabit, H., ... Sasahara, M. (2006). Mouse brains deficient in neuronal PDGF receptor-beta develop normally but are vulnerable to injury. *Journal of Neurochemistry*, 98(2), 588–600. <https://doi.org/10.1111/j.1471-4159.2006.03922.x>
- Jäkel, S., Agirre, E., Mendanha Falcão, A., van Bruggen, D., Lee, K. W., Knuesel, I., ... Castelo-Branco, G. (2019). Altered human oligodendrocyte heterogeneity in multiple sclerosis. *Nature*, 566(7745), 543–547. <https://doi.org/10.1038/s41586-019-0903-2>
- Jakovcevski, I., Miljkovic, D., Schachner, M., & Andjus, P. R. (2013). Tenascins and inflammation in disorders of the nervous system. *Amino Acids*, 44(4), 1115–1127. <https://doi.org/10.1007/s00726-012-1446-0>
- Jansen, I. E., Savage, J. E., Watanabe, K., Bryois, J., Williams, D. M., Steinberg, S., ... Posthuma, D. (2019). Genome-wide meta-analysis identifies new loci and functional pathways influencing Alzheimer's disease risk. *Nature Genetics*, 12(9), 43. <https://doi.org/10.1038/s41588-018-0311-9>

- Jellinger, K. A., & Bancher, C. (1998). Neuropathology of Alzheimer's disease: a critical update. In *Journal of Neural Transmission* (Vol. 54, pp. 77–95). https://doi.org/10.1007/978-3-7091-7508-8_8
- Jessen, N. A., Munk, A. S. F., Lundgaard, I., & Nedergaard, M. (2015). The glymphatic system: a beginner's guide. *Neurochemical Research*, 40(12), 2583–2599. <https://doi.org/10.1007/s11064-015-1581-6>
- Jiang, L., Wang, M., Lin, S., Jian, R., Li, X., Chan, J., ... Snyder, M. P. (2020). A Quantitative Proteome Map of the Human Body. *Cell*, 183(1), 269–283. <https://doi.org/10.1016/j.cell.2020.08.036>
- Jin, S., Guerrero-Juarez, C. F., Zhang, L., Chang, I., Ramos, R., Kuan, C.-H., ... Nie, Q. (2021). Inference and analysis of cell-cell communication using CellChat. *Nature Communications*, 12, 1088. <https://doi.org/10.1038/s41467-021-21246-9>
- Jordão, M. J. C., Sankowski, R., Brendecke, S. M., Sagar, Locatelli, G., Tai, Y.-H., ... Prinz, M. (2019). Single-cell profiling identifies myeloid cell subsets with distinct fates during neuroinflammation. *Science*, 363(6425), eaat7554. <https://doi.org/10.1126/science.aat7554>
- Kaltner, H., Seyrek, K., Heck, A., Sinowatz, F., & Gabius, H.-J. (2002). Galectin-1 and galectin-3 in fetal development of bovine respiratory and digestive tracts. *Cell and Tissue Research*, 307(1), 35–46. <https://doi.org/10.1007/s004410100457>
- Kalucka, J., de Rooij, L. P. M. H., Goveia, J., Rohlenova, K., Dumas, S. J., Meta, E., ... Carmeliet, P. (2020). Single-cell transcriptome atlas of murine endothelial cells. *Cell*, 180(4), 764–779. <https://doi.org/10.1016/j.cell.2020.01.015>
- Kamme, F., Salunga, R., Yu, J., Tran, D.-T., Zhu, J., Luo, L., ... Erlander, M. (2003). Single-cell microarray analysis in hippocampus CA1: demonstration and validation of cellular heterogeneity. *The Journal of Neuroscience*, 23(9), 3607–3615. <https://doi.org/10.1523/JNEUROSCI.23-09-03607.2003>
- Kampf, C., Olsson, I., Ryberg, U., Sjöstedt, E., & Pontén, F. (2012). Production of tissue microarrays, immunohistochemistry staining and digitalization within the human protein atlas. *Journal of Visualized Experiments*, (63), 6320. <https://doi.org/10.3791/3620>
- Kanazawa, M., Kawamura, K., Takahashi, T., Miura, M., Tanaka, Y., Koyama, M., ... Shimohata, T. (2015). Multiple therapeutic effects of progranulin on experimental acute ischaemic stroke. *Brain*, 138(Pt 7), 1932–1948. <https://doi.org/10.1093/brain/awv079>
- Kao, A. W., McKay, A., Singh, P. P., Brunet, A., & Huang, E. J. (2017). Progranulin, lysosomal regulation and neurodegenerative disease. *Nature Reviews Neuroscience*, 18(6), 325–333. <https://doi.org/10.1038/nrn.2017.36>
- Kealy, J., Greene, C., & Campbell, M. (2020). Blood-brain barrier regulation in psychiatric disorders. *Neuroscience Letters*, 726, 133664. <https://doi.org/10.1016/j.neulet.2018.06.033>
- Keirstead, H. S., & Blakemore, W. F. (1997). Identification of post-mitotic oligodendrocytes incapable of remyelination within the demyelinated adult spinal cord. *Journal of Neuropathology and Experimental Neurology*, 56(11), 1191–1201. <https://doi.org/10.1097/00005072-199711000-00003>
- Keller, D., Erö, C., & Markram, H. (2018). Cell densities in the mouse brain: a systematic review. *Frontiers in Neuroanatomy*, 12, 83. <https://doi.org/10.3389/fnana.2018.00083>

- Keren-Shaul, H., Spinrad, A., Weiner, A., Matcovitch-Natan, O., Dvir-Szternfeld, R., Ulland, T. K., ... Amit, I. (2017). A unique microglia type associated with restricting development of Alzheimer's disease. *Cell*, 169(7), 1276–1290. <https://doi.org/10.1016/J.CELL.2017.05.018>
- Kierdorf, K., Masuda, T., Jordão, M. J. C., & Prinz, M. (2019). Macrophages at CNS interfaces: ontogeny and function in health and disease. *Nature Reviews Neuroscience*, 20(9), 547–562. <https://doi.org/10.1038/s41583-019-0201-x>
- Kim, D., Langmead, B., & Salzberg, S. L. (2015). HISAT: a fast spliced aligner with low memory requirements. *Nature Methods*, 12(4), 357. <https://doi.org/10.1038/NMETH.3317>
- Kim, R. Y., Hoffman, A. S., Itoh, N., Ao, Y., Spence, R., Sofroniew, M. V., & Voskuhl, R. R. (2014). Astrocyte CCL2 sustains immune cell infiltration in chronic experimental autoimmune encephalomyelitis. *Journal of Neuroimmunology*, 274(1–2), 53–61. <https://doi.org/10.1016/j.jneuroim.2014.06.009>
- Kitazawa, M., Medeiros, R., & Laferla, F. M. (2012). Transgenic mouse models of Alzheimer disease: developing a better model as a tool for therapeutic interventions. *Current Pharmaceutical Design*, 18(8), 1131–1147. <https://doi.org/10.2174/138161212799315786>
- Kivioja, T., Vähäräutio, A., Karlsson, K., Bonke, M., Enge, M., Linnarsson, S., & Taipale, J. (2011). Counting absolute numbers of molecules using unique molecular identifiers. *Nature Methods*, 9(1), 72–74. <https://doi.org/10.1038/nmeth.1778>
- Kleshchevnikov, V., Shmatko, A., Dann, E., Aivazidis, A., King, H. W., Li, T., ... Bayraktar, O. A. (2020). Comprehensive mapping of tissue cell architecture via integrated single cell and spatial transcriptomics. *BioRxiv*, 2020.11.15.378125. <https://doi.org/10.1101/2020.11.15.378125>
- Kniewallner, K. M., Grimm, N., & Humpel, C. (2014). Platelet-derived nerve growth factor supports the survival of cholinergic neurons in organotypic rat brain slices. *Neuroscience Letters*, 574, 64–69. <https://doi.org/10.1016/J.NEULET.2014.05.033>
- Koch, C. M., Chiu, S. F., Akbarpour, M., Bharat, A., Ridge, K. M., Bartom, E. T., & Winter, D. R. (2018). A Beginner's Guide to Analysis of RNA Sequencing Data. *American Journal of Respiratory Cell and Molecular Biology*, 59(2), 145–157. <https://doi.org/10.1165/rcmb.2017-0430TR>
- Koppers, M., Blokhuis, A. M., Westeneng, H., Terpstra, M. L., Zundel, C. A. C., Vieira de Sá, R., ... Pasterkamp, R. J. (2015). C9orf72 ablation in mice does not cause motor neuron degeneration or motor deficits. *Annals of Neurology*, 78(3), 426–438. <https://doi.org/10.1002/ana.24453>
- Kotter, M. R., Setzu, A., Sim, F. J., Van Rooijen, N., & Franklin, R. J. (2001). Macrophage depletion impairs oligodendrocyte remyelination following lysolecithin-induced demyelination. *Glia*, 35(3), 204–212. <https://doi.org/10.1002/glia.1085>
- Kotter, M. R., Li, W.-W., Zhao, C., & Franklin, R. J. M. (2006). Myelin impairs CNS remyelination by inhibiting oligodendrocyte precursor cell differentiation. *The Journal of Neuroscience*, 26(1), 328–332. <https://doi.org/10.1523/JNEUROSCI.2615-05.2006>
- Kracht, L., Borggrewe, M., Eskandar, S., Brouwer, N., Lopes, S. M. C. de S., Laman, J. D., ... Eggen, B. J. L. (2020). Human fetal microglia acquire homeostatic immune-sensing properties early in development. *Science*, 369(6503), 530–537. <https://doi.org/10.1126/SCIENCE.ABA5906>

- Krasemann, S., Madore, C., Cialic, R., Baufeld, C., Calcagno, N., El Fatimy, R., ... Butovsky, O. (2017). The TREM2-APOE pathway drives the transcriptional phenotype of dysfunctional microglia in neurodegenerative diseases. *Immunity*, 47(3), 566–581. <https://doi.org/10.1016/j.immuni.2017.08.008>
- Krishnaswami, S. R., Grindberg, R. V, Novotny, M., Venepally, P., Lacar, B., Bhutani, K., ... Lasken, R. S. (2016). Using single nuclei for RNA-seq to capture the transcriptome of postmortem neurons. *Nature Protocols*, 11(3), 499–524. <https://doi.org/10.1038/nprot.2016.015>
- Kuhlmann, T, Miron, V., Cui, Q., Cuo, Q., Wegner, C., Antel, J., & Brück, W. (2008). Differentiation block of oligodendroglial progenitor cells as a cause for remyelination failure in chronic multiple sclerosis. *Brain*, 131(7), 1749–1758. <https://doi.org/10.1093/brain/awn096>
- Kuhlmann, T., Ludwin, S., Prat, A., Antel, J., Brück, W., & Lassmann, H. (2017). An updated histological classification system for multiple sclerosis lesions. *Acta Neuropathologica*, 133(1), 13–24. <https://doi.org/10.1007/s00401-016-1653-y>
- Kumar, P., Ning, Y., Polverini, P. J., & Kumar, P. (2008). Endothelial cells expressing Bcl-2 promote tumor metastasis by enhancing tumor angiogenesis, blood vessel leakiness and tumor invasion. *Laboratory Investigation*, 88, 740–749. <https://doi.org/10.1038/labinvest.2008.46>
- L. Lun, A. T., Bach, K., & Marioni, J. C. (2016). Pooling across cells to normalize single-cell RNA sequencing data with many zero counts. *Genome Biology*, 17(1), 75. <https://doi.org/10.1186/s13059-016-0947-7>
- Lake, B. B., Ai, R., Kaeser, G. E., Salathia, N. S., Yung, Y. C., Liu, R., ... Zhang, K. (2016). Neuronal subtypes and diversity revealed by single-nucleus RNA sequencing of the human brain. *Science*, 352(6293), 1586–1590. <https://doi.org/10.1126/science.aaf1204>
- Lake, B. B., Chen, S., Sos, B. C., Fan, J., Kaeser, G. E., Yung, Y. C., ... Zhang, K. (2018). Integrative single-cell analysis of transcriptional and epigenetic states in the human adult brain. *Nature Biotechnology*, 36(1), 70–80. <https://doi.org/10.1038/nbt.4038>
- Lake, B. B., Codeluppi, S., Yung, Y. C., Gao, D., Chun, J., Kharchenko, P. V, ... Zhang, K. (2017). A comparative strategy for single-nucleus and single-cell transcriptomes confirms accuracy in predicted cell-type expression from nuclear RNA. *Scientific Reports*, 7(1), 6031. <https://doi.org/10.1038/s41598-017-04426-w>
- Lall, D., Lorenzini, I., Mota, T. A., Holtzman, D. M., Sattler, R., Correspondence, R. H. B., ... Baloh, R. H. (2021). C9orf72 deficiency promotes microglial-mediated synaptic loss in aging and amyloid accumulation. *Neuron*, 109, 1–17. <https://doi.org/10.1016/j.neuron.2021.05.020>
- Langfelder, P., & Horvath, S. (2008). WGCNA: an R package for weighted correlation network analysis. *BMC Bioinformatics*, 9(1), 559. <https://doi.org/10.1186/1471-2105-9-559>
- Langfelder, P., & Horvath, S. (2012). Fast R functions for robust correlations and hierarchical clustering. *Journal of Statistical Software*, 46(11), 1–17. <https://doi.org/10.18637/jss.v046.i11>
- Lavin, Y., Winter, D., Blecher-Gonen, R., David, E., Keren-Shaul, H., Merad, M., ... Amit, I. (2014). Tissue-resident macrophage enhancer landscapes are shaped by the local microenvironment. *Cell*, 159(6), 1312–1326. <https://doi.org/10.1016/j.cell.2014.11.018>
- Lawson, L. J., Perry, V. H., Dri, P., & Gordon, S. (1990). Heterogeneity in the distribution and morphology of microglia in the normal adult mouse brain. *Neuroscience*, 39(1), 151–170.

[https://doi.org/10.1016/0306-4522\(90\)90229-W](https://doi.org/10.1016/0306-4522(90)90229-W)

Lee, S. H., Kim, W. T., Cornell-Bell, A. H., & Sontheimer, H. (1994). Astrocytes exhibit regional specificity in gap-junction coupling. *Glia*, 11(4), 315–325. <https://doi.org/10.1002/glia.440110404>

Lendahl, U., Nilsson, P., & Betsholtz, C. (2019). Emerging links between cerebrovascular and neurodegenerative diseases—a special role for pericytes. *EMBO Reports*, 20(11), e48070. <https://doi.org/10.15252/embr.201948070>

Lentferink, D. H., Jongsma, J. M., Werkman, I., & Baron, W. (2018). Grey matter OPCs are less mature and less sensitive to IFN γ than white matter OPCs: consequences for remyelination. *Scientific Reports*, 8(1), 2113. <https://doi.org/10.1038/s41598-018-19934-6>

Leroy, K., Ando, K., Laporte, V., Dedecker, R., Suain, V., Authélet, M., ... Brion, J.-P. (2012). Lack of tau proteins rescues neuronal cell death and decreases amyloidogenic processing of APP in APP/PS1 mice. *The American Journal of Pathology*, 181(6), 1928–1940. <https://doi.org/10.1016/j.ajpath.2012.08.012>

Levin, M. H., Bennett, J. L., & Verkman, A. S. (2013). Optic neuritis in neuromyelitis optica. *Progress in Retinal and Eye Research*, 36, 159. <https://doi.org/10.1016/j.PRETEYERES.2013.03.001>

Lewis, J., McGowan, E., Rockwood, J., Melrose, H., Nacharaju, P., Van Slegtenhorst, M., ... Hutton, M. (2000). Neurofibrillary tangles, amyotrophy and progressive motor disturbance in mice expressing mutant (P301L) tau protein. *Nature Genetics*, 25(4), 402–405. <https://doi.org/10.1038/78078>

Li, Q., Cheng, Z., Zhou, L., Darmanis, S., Neff, N. F., Okamoto, J., ... Barres, B. A. (2019). Developmental heterogeneity of microglia and brain myeloid cells revealed by deep single-cell RNA sequencing. *Neuron*, 101(2), 207–223. <https://doi.org/10.1016/j.neuron.2018.12.006>

Li, W., Notani, D., & Rosenfeld, M. G. (2016). Enhancers as non-coding RNA transcription units: recent insights and future perspectives. *Nature Reviews Genetics*, 17(4), 207–223. <https://doi.org/10.1038/nrg.2016.4>

Liddelw, S. A., Guttenplan, K. A., Clarke, L. E., Bennett, F. C., Bohlen, C. J., Schirmer, L., ... Barres, B. A. (2017). Neurotoxic reactive astrocytes are induced by activated microglia. *Nature*, 541(7638), 481–487. <https://doi.org/10.1038/nature21029>

Lippman, J. J., Lordkipanidze, T., Buell, M. E., Yoon, S. O., & Dunaevsky, A. (2008). Morphogenesis and regulation of Bergmann glial processes during Purkinje cell dendritic spine ensheathment and synaptogenesis. *Glia*, 56(13), 1463–1477. <https://doi.org/10.1002/glia.20712>

Liu, K. K. Y., & Dorovini-Zis, K. (2009). Regulation of CXCL12 and CXCR4 expression by human brain endothelial cells and their role in CD4+ and CD8+ T cell adhesion and transendothelial migration. *Journal of Neuroimmunology*, 215(1–2), 49–64. <https://doi.org/10.1016/j.jneuroim.2009.08.003>

Liu, X., Lu, Y., Zhang, Y., Li, Y., Zhou, J., Yuan, Y., ... He, C. (2012). Slit2 regulates the dispersal of oligodendrocyte precursor cells via Fyn/RhoA signaling. *The Journal of Biological Chemistry*, 287(21), 17503–17516. <https://doi.org/10.1074/jbc.M111.317610>

Lodish, H., Berk, A., Zipursky, S. L., Matsudaira, P., Baltimore, D., & Darnell, J. (2000).

- Overview of neuron structure and function. In *Molecular Cell Biology* (4th ed.). W. H. Freeman. Retrieved from <https://www.ncbi.nlm.nih.gov/books/NBK21535/>
- Love, M. I., Huber, W., & Anders, S. (2014). Moderated estimation of fold change and dispersion for RNA-seq data with DESeq2. *Genome Biology*, 15(12), 550. <https://doi.org/10.1186/s13059-014-0550-8>
- Lubetzki, C., & Stankoff, B. (2014). Demyelination in multiple sclerosis. *Handbook of Clinical Neurology*, 122, 89–99. <https://doi.org/10.1016/B978-0-444-52001-2.00004-2>
- Luchetti, S., Fransen, N. L., van Eden, C. G., Ramaglia, V., Mason, M., & Huitinga, I. (2018). Progressive multiple sclerosis patients show substantial lesion activity that correlates with clinical disease severity and sex: a retrospective autopsy cohort analysis. *Acta Neuropathologica*, 135(4), 511–528. <https://doi.org/10.1007/s00401-018-1818-y>
- Luchicchi, A., Hart, B., Frigerio, I., van Dam, A.-M., Perna, L., Offerhaus, H. L., ... Geurts, J. J. G. (2021). Axon-myelin unit blistering as early event in MS normal appearing white matter. *Annals of Neurology*, 89(4), 711–725. <https://doi.org/10.1002/ana.26014>
- Ludwin, S. K., & Maitland, M. (1984). Long-term remyelination fails to reconstitute normal thickness of central myelin sheaths. *Journal of the Neurological Sciences*, 64(2), 193–198. [https://doi.org/10.1016/0022-510x\(84\)90037-6](https://doi.org/10.1016/0022-510x(84)90037-6)
- Lui, H., Zhang, J., Makinson, S. R., Cahill, M. K., Kelley, K. W., Huang, H.-Y., ... Huang, E. J. (2016). Progranulin deficiency promotes circuit-specific synaptic pruning by microglia via complement activation. *Cell*, 165(4), 921–935. <https://doi.org/10.1016/j.cell.2016.04.001>
- Lun, A. T. L., Riesenfeld, S., Andrews, T., Dao, T. P., Gomes, T., & Marioni, J. C. (2019). EmptyDrops: distinguishing cells from empty droplets in droplet-based single-cell RNA sequencing data. *Genome Biology*, 20(1), 63. <https://doi.org/10.1186/s13059-019-1662-y>
- Lundgaard, I., Osório, M. J., Kress, B. T., Sanggaard, S., & Nedergaard, M. (2014). White matter astrocytes in health and disease. *Neuroscience*, 276, 161–173. <https://doi.org/10.1016/j.neuroscience.2013.10.050>
- Lytal, N., Ran, D., & An, L. (2020). Normalization methods on single-cell RNA-seq data: an empirical survey. *Frontiers in Genetics*, 11, 41. <https://doi.org/10.3389/fgene.2020.00041>
- Ma, S.-C., Li, Q., Peng, J.-Y., Zhouwen, J.-L., Diao, J.-F., Niu, J.-X., ... Jiang, W.-G. (2017). Claudin-5 regulates blood-brain barrier permeability by modifying brain microvascular endothelial cell proliferation, migration, and adhesion to prevent lung cancer metastasis. *CNS Neuroscience & Therapeutics*, 23(12), 947–960. <https://doi.org/10.1111/cns.12764>
- Mackenzie, I. R. A., & Neumann, M. (2016). Molecular neuropathology of frontotemporal dementia: insights into disease mechanisms from postmortem studies. *Journal of Neurochemistry*, 138 Suppl, 54–70. <https://doi.org/10.1111/jnc.13588>
- Mäe, M. A., He, L., Nordling, S., Vazquez-Liebanas, E., Nahar, K., Jung, B., ... Betsholtz, C. (2021). Single-cell analysis of blood-brain barrier response to pericyte loss. *Circulation Research*, 128(4), e46–e62. <https://doi.org/10.1161/CIRCRESAHA.120.317473>
- Maes, C., Coenegrachts, L., Stockmans, I., Daci, E., Luttun, A., Petryk, A., ... Carmeliet, G. (2006). Placental growth factor mediates mesenchymal cell development, cartilage turnover, and bone remodeling during fracture repair. *The Journal of Clinical Investigation*, 116(5), 1230–1242. <https://doi.org/10.1172/JCI26772>

- Magnusson, J. P., Göritz, C., Tatarishvili, J., Dias, D. O., Smith, E. M. K., Lindvall, O., ... Frisé, J. (2014). A latent neurogenic program in astrocytes regulated by Notch signaling in the mouse. *Science*, 346(6206), 237–241. <https://doi.org/10.1126/science.346.6206.237>
- Månberg, A., Skene, N., Sanders, F., Trusohamn, M., Remnestrål, J., Szczepińska, A., ... Lewandowski, S. A. (2021). Altered perivascular fibroblast activity precedes ALS disease onset. *Nature Medicine*, 27(4), 640–646. <https://doi.org/10.1038/s41591-021-01295-9>
- Mandel, I., Paperna, T., Glass-Marmor, L., Volkowich, A., Badarny, S., Schwartz, I., ... Miller, A. (2012). Tight junction proteins expression and modulation in immune cells and multiple sclerosis. *Journal of Cellular and Molecular Medicine*, 16(4), 765–775. <https://doi.org/10.1111/j.1582-4934.2011.01380.x>
- Marques, S., van Bruggen, D., Vanichkina, D. P., Floriddia, E. M., Munguba, H., Våremo, L., ... Castelo-Branco, G. (2018). Transcriptional convergence of oligodendrocyte lineage progenitors during development. *Developmental Cell*, 46(4), 504–517. <https://doi.org/10.1016/j.devcel.2018.07.005>
- Marques, S., Zeisel, A., Codeluppi, S., van Bruggen, D., Mendanha Falcão, A., Xiao, L., ... Castelo-Branco, G. (2016). Oligodendrocyte heterogeneity in the mouse juvenile and adult central nervous system. *Science*, 352(6291), 1326–1329. <https://doi.org/10.1126/science.aaf6463>
- Marsh, S. E., Kamath, T., Walker, A. J., Dissing-Olesen, L., Hammond, T. R., Young, A. M. H., ... Stevens, B. (2020). Single cell sequencing reveals glial specific responses to tissue processing & enzymatic dissociation in mice and humans. *BioRxiv*, 2020.12.03.408542. <https://doi.org/10.1101/2020.12.03.408542>
- Martin, J. A., Craft, D. K., Su, J. H., Kim, R. C., & Cotman, C. W. (2001). Astrocytes degenerate in frontotemporal dementia: possible relation to hypoperfusion. *Neurobiology of Aging*, 22(2), 195–207. Retrieved from www.elsevier.com/locate/neuaging
- Masters, C. L., & Selkoe, D. J. (2012). Biochemistry of amyloid β -protein and amyloid deposits in Alzheimer disease. *Cold Spring Harbor Perspectives in Medicine*, 2(6), a006262. <https://doi.org/10.1101/cshperspect.a006262>
- Masuda, T., Sankowski, R., Staszewski, O., Böttcher, C., Amann, L., Sagar, ... Prinz, M. (2019). Spatial and temporal heterogeneity of mouse and human microglia at single-cell resolution. *Nature*, 566(7744), 388–392. <https://doi.org/10.1038/s41586-019-0924-x>
- Matcovitch-Natan, O., Winter, D. R., Giladi, A., Vargas Aguilar, S., Spinrad, A., Sarrazin, S., ... Amit, I. (2016). Microglia development follows a stepwise program to regulate brain homeostasis. *Science*, 353(6301), aad8670. <https://doi.org/10.1126/science.aad8670>
- Mathys, H., Adaikkan, C., Gao, F., Young, J. Z., Manet, E., Hemberg, M., ... Tsai, L.-H. (2017). Temporal tracking of microglia activation in neurodegeneration at single-cell resolution. *Cell Reports*, 21(2), 366–380. <https://doi.org/10.1016/j.celrep.2017.09.039>
- Mathys, H., Davila-Velderrain, J., Peng, Z., Gao, F., Mohammadi, S., Young, J. Z., ... Tsai, L.-H. (2019). Single-cell transcriptomic analysis of Alzheimer's disease. *Nature*, 570(7761), 332–337. <https://doi.org/10.1038/s41586-019-1195-2>
- Matias, I., Morgado, J., & Gomes, F. C. A. (2019). Astrocyte heterogeneity: impact to brain aging and disease. *Frontiers in Aging Neuroscience*, 11, 59. <https://doi.org/10.3389/fnagi.2019.00059>

- Mattei, D., Ivanov, A., van Oostrum, M., Pantelyushin, S., Richetto, J., Mueller, F., ... Meyer, U. (2020). Enzymatic dissociation induces transcriptional and proteotype bias in brain cell populations. *International Journal of Molecular Sciences*, 21(21). <https://doi.org/10.3390/ijms21217944>
- Maynard, K. R., Collado-Torres, L., Weber, L. M., Uyttingco, C., Barry, B. K., Williams, S. R., ... Jaffe, A. E. (2021). Transcriptome-scale spatial gene expression in the human dorsolateral prefrontal cortex. *Nature Neuroscience*, 24(3), 425–436. <https://doi.org/10.1038/s41593-020-00787-0>
- Meeter, L. H., Kaat, L. D., Rohrer, J. D., & van Swieten, J. C. (2017). Imaging and fluid biomarkers in frontotemporal dementia. *Nature Reviews Neurology*, 13(7), 406–419. <https://doi.org/10.1038/nrneurol.2017.75>
- Melief, J., Orre, M., Bossers, K., van Eden, C. G., Schuurman, K. G., Mason, M. R. J., ... Huitinga, I. (2019). Transcriptome analysis of normal-appearing white matter reveals cortisol- and disease-associated gene expression profiles in multiple sclerosis. *Acta Neuropathologica Communications*, 7(1), 60. <https://doi.org/10.1186/s40478-019-0705-7>
- Mendiola, A. S., Ryu, J. K., Bardehle, S., Meyer-Franke, A., Ang, K. K.-H., Wilson, C., ... Akassoglou, K. (2020). Transcriptional profiling and therapeutic targeting of oxidative stress in neuroinflammation. *Nature Immunology*, 21(5), 513–524. <https://doi.org/10.1038/s41590-020-0654-0>
- Menzel, L., Kleber, L., Friedrich, C., Hummel, R., Dangel, L., Winter, J., ... Schäfer, M. K. E. (2017). Progranulin protects against exaggerated axonal injury and astrogliosis following traumatic brain injury. *Glia*, 65(2), 278–292. <https://doi.org/10.1002/glia.23091>
- Mestre, H., Hablitz, L. M., Xavier, A. L., Feng, W., Zou, W., Pu, T., ... Nedergaard, M. (2018). Aquaporin-4-dependent glymphatic solute transport in the rodent brain. *eLife*, 7, e40070. <https://doi.org/10.7554/eLife.40070>
- Michael, B. D., Bricio-Moreno, L., Sorensen, E. W., Solomon, T., Kurt-Jones, E. A., Luster Correspondence, A. D., ... Luster, A. D. (2020). Astrocyte- and neuron-derived CXCL1 drives neutrophil transmigration and blood-brain barrier permeability in viral encephalitis. *Cell Reports*, 32(11), 108150. <https://doi.org/10.1016/j.celrep.2020.108150>
- Miller, D. H., Khan, O. A., Sheremata, W. A., Blumhardt, L. D., Rice, G. P. A., Libonati, M. A., ... International Natalizumab Multiple Sclerosis Trial Group. (2003). A controlled trial of natalizumab for relapsing multiple sclerosis. *The New England Journal of Medicine*, 348(1), 15–23. <https://doi.org/10.1056/NEJMoa020696>
- Mitew, S., Gobius, I., Fenlon, L. R., McDougall, S. J., Hawkes, D., Xing, Y. L., ... Emery, B. (2018). Pharmacogenetic stimulation of neuronal activity increases myelination in an axon-specific manner. *Nature Communications*, 9(1), 306. <https://doi.org/10.1038/s41467-017-02719-2>
- Moisse, K., Volkening, K., Leystra-Lantz, C., Welch, I., Hill, T., & Strong, M. J. (2009). Divergent patterns of cytosolic TDP-43 and neuronal progranulin expression following axotomy: implications for TDP-43 in the physiological response to neuronal injury. *Brain Research*, 1249, 202–211. <https://doi.org/10.1016/j.brainres.2008.10.021>
- Montagne, A., Barnes, S. R., Sweeney, M. D., Halliday, M. R., Sagare, A. P., Zhao, Z., ... Chair, S. Z. (2015). Blood-brain barrier breakdown in the aging human hippocampus. *Neuron*,

85(2), 296–302. <https://doi.org/10.1016/j.neuron.2014.12.032>

Montine, T. J., Phelps, C. H., Beach, T. G., Bigio, E. H., Cairns, N. J., Dickson, D. W., ... Alzheimer's Association. (2012). National Institute on Aging–Alzheimer's Association guidelines for the neuropathologic assessment of Alzheimer's disease: a practical approach. *Acta Neuropathologica*, 123(1), 1–11. <https://doi.org/10.1007/s00401-011-0910-3>

Morabito, S., Miyoshi, E., Michael, N., Shahin, S., Martini, A. C., Head, E., ... Swarup, V. (2021). Single-nucleus chromatin accessibility and transcriptomic characterization of Alzheimer's disease. *Nature Genetics*, 53(8), 1143–1155. <https://doi.org/10.1038/s41588-021-00894-z>

Mortazavi, A., Williams, B. A., McCue, K., Schaeffer, L., & Wold, B. (2008). Mapping and quantifying mammalian transcriptomes by RNA-Seq. *Nature Methods*, 5(7), 621–628. <https://doi.org/10.1038/nmeth.1226>

Mucke, L., Yu, G. Q., McConlogue, L., Rockenstein, E. M., Abraham, C. R., & Masliah, E. (2000). Astroglial expression of human alpha(1)-antichymotrypsin enhances alzheimer-like pathology in amyloid protein precursor transgenic mice. *The American Journal of Pathology*, 157(6), 2003–2010. [https://doi.org/10.1016/s0002-9440\(10\)64839-0](https://doi.org/10.1016/s0002-9440(10)64839-0)

Munji, R. N., Soung, A. L., Weiner, G. A., Sohet, F., Semple, B. D., Trivedi, A., ... Daneman, R. (2019). Profiling the mouse brain endothelial transcriptome in health and disease models reveals a core blood–brain barrier dysfunction module. *Nature Neuroscience*, 22(11), 1892–1902. <https://doi.org/10.1038/s41593-019-0497-x>

Muñoz, M. F., Puebla, M., & Figueroa, X. F. (2015). Control of the neurovascular coupling by nitric oxide-dependent regulation of astrocytic Ca²⁺ signaling. *Frontiers in Cellular Neuroscience*, 9, 59. <https://doi.org/10.3389/fncel.2015.00059>

Murtha, L. A., Yang, Q., Parsons, M. W., Levi, C. R., Beard, D. J., Spratt, N. J., & McLeod, D. D. (2014). Cerebrospinal fluid is drained primarily via the spinal canal and olfactory route in young and aged spontaneously hypertensive rats. *Fluids and Barriers of the CNS*, 11, 12. <https://doi.org/10.1186/2045-8118-11-12>

Nait-Oumesmar, B., Picard-Riera, N., Kerninon, C., Decker, L., Seilhean, D., Höglinger, G. U., ... Baron-Van Evercooren, A. (2007). Activation of the subventricular zone in multiple sclerosis: evidence for early glial progenitors. *Proceedings of the National Academy of Sciences*, 104(11), 4694–4699. <https://doi.org/10.1073/pnas.0606835104>

Nalivaeva, N. N., & Turner, A. J. (2019). Targeting amyloid clearance in Alzheimer's disease as a therapeutic strategy. *British Journal of Pharmacology*, 176(18), 3447–3463. <https://doi.org/10.1111/bph.14593>

Nation, D. A., Sweeney, M. D., Montagne, A., Sagare, A. P., D'Orazio, L. M., Pachicano, M., ... Zlokovic, B. V. (2019). Blood-brain barrier breakdown is an early biomarker of human cognitive dysfunction. *Nature Medicine*, 25(2), 270–276. <https://doi.org/10.1038/s41591-018-0297-y>

Nelson, P. T., Alafuzoff, I., Bigio, E. H., Bouras, C., Braak, H., Cairns, N. J., ... Beach, T. G. (2012). Correlation of Alzheimer disease neuropathologic changes with cognitive status: a review of the literature. *Journal of Neuropathology and Experimental Neurology*, 71(5), 362–381. <https://doi.org/10.1097/NEN.0b013e31825018f7>

Neu, K. E., Tang, Q., Wilson, P. C., & Khan, A. A. (2017). Single-cell genomics: approaches

and utility in immunology. *Trends in Immunology*, 38(2), 140–149. <https://doi.org/10.1016/j.it.2016.12.001>

Newman, A. M., Steen, C. B., Liu, C. L., Gentles, A. J., Chaudhuri, A. A., Scherer, F., ... Alizadeh, A. A. (2019). Determining cell type abundance and expression from bulk tissues with digital cytometry. *Nature Biotechnology*, 37(7), 773–782. <https://doi.org/10.1038/s41587-019-0114-2>

Nguyen, A. T., Wang, K., Hu, G., Wang, X., Miao, Z., Azevedo, J. A., ... Lee, E. B. (2020). APOE and TREM2 regulate amyloid-responsive microglia in Alzheimer's disease. *Acta Neuropathologica*, 1–17. <https://doi.org/10.1007/s00401-020-02200-3>

Nicholson, D. A., Yoshida, R., Berry, R. W., Gallagher, M., & Geinisman, Y. (2004). Reduction in size of perforated postsynaptic densities in hippocampal axospinous synapses and age-related spatial learning impairments. *The Journal of Neuroscience*, 24(35), 7648–7653. <https://doi.org/10.1523/JNEUROSCI.1725-04.2004>

Nitta, T., Hata, M., Gotoh, S., Seo, Y., Sasaki, H., Hashimoto, N., ... Tsukita, S. (2003). Size-selective loosening of the blood-brain barrier in claudin-5-deficient mice. *The Journal of Cell Biology*, 161(3), 653–660. <https://doi.org/10.1083/jcb.200302070>

Number of people with MS | Atlas of MS. (2021). Retrieved June 22, 2021, from <https://www.atlasofms.org/map/global/epidemiology/number-of-people-with-ms>

O’Brown, N. M., Pfau, S. J., & Gu, C. (2018). Bridging barriers: a comparative look at the blood-brain barrier across organisms. *Genes & Development*, 32(7–8), 466–478. <https://doi.org/10.1101/gad.309823.117>

Oberheim, N. A., Takano, T., Han, X., He, W., Lin, J. H. C., Wang, F., ... Nedergaard, M. (2009). Uniquely hominid features of adult human astrocytes. *The Journal of Neuroscience*, 29(10), 3276. <https://doi.org/10.1523/JNEUROSCI.4707-08.2009>

Oddo, S., Caccamo, A., Shepherd, J. D., Murphy, M. P., Golde, T. E., Kaye, R., ... LaFerla, F. M. (2003). Triple-transgenic model of Alzheimer's disease with plaques and tangles: intracellular Abeta and synaptic dysfunction. *Neuron*, 39(3), 409–421. [https://doi.org/10.1016/s0896-6273\(03\)00434-3](https://doi.org/10.1016/s0896-6273(03)00434-3)

Olabarria, M., Noristani, H. N., Verkhratsky, A., & Rodríguez, J. J. (2010). Concomitant astroglial atrophy and astrogliosis in a triple transgenic animal model of Alzheimer's disease. *Glia*, 58(7), 831–838. <https://doi.org/10.1002/glia.20967>

Olah, M., Menon, V., Habib, N., Taga, M. F., Ma, Y., Yung, C. J., ... Jager, P. L. De. (2020). Single cell RNA sequencing of human microglia uncovers a subset associated with Alzheimer's disease. *Nature Communications*, 11, 6129. <https://doi.org/10.1038/S41467-020-19737-2>

Olah, M., Menon, V., Habib, N., Taga, M., Yung, C., Cimpean, M., ... Jager, P. L. De. (2018). A single cell-based atlas of human microglial states reveals associations with neurological disorders and histopathological features of the aging brain. *BioRxiv*. <https://doi.org/10.1101/343780>

Olszewska, D. A., Lonergan, R., Fallon, E. M., & Lynch, T. (2016). Genetics of frontotemporal dementia. *Current Neurology and Neuroscience Reports*, 16(12), 107. <https://doi.org/10.1007/s11910-016-0707-9>

Onyike, C. U., & Diehl-Schmid, J. (2013). The epidemiology of frontotemporal dementia.

- International Review of Psychiatry*, 25(2), 130–137. <https://doi.org/10.3109/09540261.2013.776523>
- Özen, I., Deierborg, T., Miharada, K., Padel, T., Englund, E., Genové, G., & Paul, G. (2014). Brain pericytes acquire a microglial phenotype after stroke. *Acta Neuropathologica*, 128(3), 381–396. <https://doi.org/10.1007/s00401-014-1295-x>
- Padel, T., Roth, M., Gaceb, A., Li, J.-Y., Björkqvist, M., & Paul, G. (2018). Brain pericyte activation occurs early in Huntington's disease. *Experimental Neurology*, 305, 139–150. <https://doi.org/10.1016/j.expneurol.2018.03.015>
- Paolicelli, R. C., Bolasco, G., Pagani, F., Maggi, L., Scianni, M., Panzanelli, P., ... Gross, C. T. (2011). Synaptic pruning by microglia is necessary for normal brain development. *Science*, 333(6048), 1456–1458. <https://doi.org/10.1126/SCIENCE.1202529>
- Papadopoulos, M. C., & Verkman, A. S. (2013). Aquaporin water channels in the nervous system. *Nature Reviews Neuroscience*, 14(4), 265–277. <https://doi.org/10.1038/nrn3468>
- Pasterkamp, R. J., De Winter, F., Holtmaat, A. J., & Verhaagen, J. (1998). Evidence for a role of the chemorepellent semaphorin III and its receptor neuropilin-1 in the regeneration of primary olfactory axons. *The Journal of Neuroscience*, 18(23), 9962–9976. <https://doi.org/10.1523/JNEUROSCI.18-23-09962.1998>
- Patani, R., Balaratnam, M., Vora, A., & Reynolds, R. (2007). Remyelination can be extensive in multiple sclerosis despite a long disease course. *Neuropathology and Applied Neurobiology*, 33(3), 277–287. <https://doi.org/10.1111/j.1365-2990.2007.00805.x>
- Patrikios, P., Stadelmann, C., Kutzelnigg, A., Rauschka, H., Schmidbauer, M., Laursen, H., ... Lassmann, H. (2006). Remyelination is extensive in a subset of multiple sclerosis patients. *Brain*, 129(Pt 12), 3165–3172. <https://doi.org/10.1093/brain/awl217>
- Paty, D. W., & Li, D. K. (1993). Interferon beta-1b is effective in relapsing-remitting multiple sclerosis. *Neurology*, 43(4), 662–667. <https://doi.org/10.1212/wnl.43.4.662>
- Paul, G., Özen, I., Christophersen, N. S., Reinbothe, T., Bengzon, J., Visse, E., ... Brundin, P. (2012). The adult human brain harbors multipotent perivascular mesenchymal stem cells. *PLOS ONE*, 7(4), e35577. <https://doi.org/10.1371/journal.pone.0035577>
- Pekny, M., Wilhelmsson, U., & Pekna, M. (2014). The dual role of astrocyte activation and reactive gliosis. *Neuroscience Letters*, 565, 30–38. <https://doi.org/10.1016/j.neulet.2013.12.071>
- Pelvig, D. P., Pakkenberg, H., Stark, A. K., & Pakkenberg, B. (2008). Neocortical glial cell numbers in human brains. *Neurobiology of Aging*, 29, 1754–1762. <https://doi.org/10.1016/j.neurobiolaging.2007.04.013>
- Perez-Pinera, P., Chang, Y., & Deuel, T. F. (2007). Pleiotrophin, a multifunctional tumor promoter through induction of tumor angiogenesis, remodeling of the tumor microenvironment, and activation of stromal fibroblasts. *Cell Cycle*, 6(23), 2877–2883. <https://doi.org/10.4161/cc.6.23.5090>
- Perlman, K., Couturier, C. P., Yaqubi, M., Tanti, A., Cui, Q., Pernin, F., ... Antel, J. P. (2020). Developmental trajectory of oligodendrocyte progenitor cells in the human brain revealed by single cell RNA sequencing. *Glia*, 68(6), 1291–1303. <https://doi.org/10.1002/glia.23777>
- Picelli, S., Faridani, O. R., Björklund, Å. K., Winberg, G., Sagasser, S., & Sandberg, R. (2014).

- Full-length RNA-seq from single cells using Smart-seq2. *Nature Protocols*, 9, 171. Retrieved from <https://doi.org/10.1038/nprot.2014.006>
- Pierson, E., & Yau, C. (2015). ZIFA: Dimensionality reduction for zero-inflated single-cell gene expression analysis. *Genome Biology*, 16, 241. <https://doi.org/10.1186/s13059-015-0805-z>
- Piscopo, P., Grasso, M., Fontana, F., Crestini, A., Puopolo, M., Del Vescovo, V., ... Denti, M. A. (2016). Reduced miR-659-3p levels correlate with progranulin increase in hypoxic conditions: implications for frontotemporal dementia. *Frontiers in Molecular Neuroscience*, 9, 31. <https://doi.org/10.3389/fnmol.2016.00031>
- Plemel, J. R., Stratton, J. A., Michaels, N. J., Rawji, K. S., Zhang, E., Sinha, S., ... Yong, V. W. (2020). Microglia response following acute demyelination is heterogeneous and limits infiltrating macrophage dispersion. *Science Advances*, 6(3), eaay6324. <https://doi.org/10.1126/sciadv.aay6324>
- Ponath, G., Park, C., & Pitt, D. (2018). The role of astrocytes in multiple sclerosis. *Frontiers in Immunology*, 9, 217. <https://doi.org/10.3389/fimmu.2018.00217>
- Ponath, G., Ramanan, S., Mubarak, M., Housley, W., Lee, S., Sahinkaya, F. R., ... Pitt, D. (2017). Myelin phagocytosis by astrocytes after myelin damage promotes lesion pathology. *Brain*, 140(2), 399–413. <https://doi.org/10.1093/brain/aww298>
- Prabhakaran, S., Azizi, E., Carr, A., & Pe'er, D. (2016). Dirichlet process mixture model for correcting technical variation in single-cell gene expression data. *JMLR Workshop and Conference Proceedings*, 48, 1070–1079. Retrieved from <http://www.ncbi.nlm.nih.gov/pubmed/29928470>
- Prinz, M., & Priller, J. (2014). Microglia and brain macrophages in the molecular age: from origin to neuropsychiatric disease. *Nature Reviews Neuroscience*, 15(5), 300–312. <https://doi.org/10.1038/nrn3722>
- Privratsky, J. R., & Newman, P. J. (2014). PECAM-1: regulator of endothelial junctional integrity. *Cell and Tissue Research*, 355(3), 607–619. <https://doi.org/10.1007/s00441-013-1779-3>
- Profaci, C. P., Munji, R. N., Pulido, R. S., & Daneman, R. (2020). The blood-brain barrier in health and disease: Important unanswered questions. *The Journal of Experimental Medicine*, 217(4), e20190062. <https://doi.org/10.1084/jem.20190062>
- Propson, N. E., Roy, E. R., Litvinchuk, A., Köhl, J., & Zheng, H. (2021). Endothelial C3a receptor mediates vascular inflammation and blood-brain barrier permeability during aging. *The Journal of Clinical Investigation*, 131(1), e140966. <https://doi.org/10.1172/JCI140966>
- Qiu, X., Mao, Q., Tang, Y., Wang, L., Chawla, R., Pliner, H. A., & Trapnell, C. (2017). Reversed graph embedding resolves complex single-cell trajectories. *Nature Methods*, 14(10), 979–982. <https://doi.org/10.1038/nmeth.4402>
- Rakic, P. (2009). Evolution of the neocortex: a perspective from developmental biology. *Nature Reviews Neuroscience*, 10(10), 724–735. <https://doi.org/10.1038/nrn2719>
- Rasia-Filho, A. A., Guerra, K. T. K., Vásquez, C. E., Dall'Oglio, A., Reberger, R., Jung, C. R., & Calcagnotto, M. E. (2021). The subcortical-allocortical- neocortical continuum for the emergence and morphological heterogeneity of pyramidal neurons in the human brain. *Frontiers in Synaptic Neuroscience*, 13, 7. <https://doi.org/10.3389/fnsyn.2021.616607>

- Rensink, A. A. M., Verbeek, M. M., Otte-Höller, I., ten Donkelaar, H. T., de Waal, R. M. W., & Kremer, B. (2002). Inhibition of amyloid-beta-induced cell death in human brain pericytes in vitro. *Brain Research*, 952(1), 111–121. [https://doi.org/10.1016/s0006-8993\(02\)03218-3](https://doi.org/10.1016/s0006-8993(02)03218-3)
- Riku, Y., Duyckaerts, C., Boluda, S., Plu, I., Le Ber, I., Millecamps, S., ... Seilhean, D. (2019). Increased prevalence of granulovacuolar degeneration in C9orf72 mutation. *Acta Neuropathologica*, 138(5), 783–793. <https://doi.org/10.1007/s00401-019-02028-6>
- Rivers, L. E., Young, K. M., Rizzi, M., Jamen, F., Psachoulia, K., Wade, A., ... Richardson, W. D. (2008). PDGFRA/NG2 glia generate myelinating oligodendrocytes and piriform projection neurons in adult mice. *Nature Neuroscience*, 11(12), 1392–1401. <https://doi.org/10.1038/nn.2220>
- Robinson, A. P., Harp, C. T., Noronha, A., & Miller, S. D. (2014). The experimental autoimmune encephalomyelitis (EAE) model of MS: utility for understanding disease pathophysiology and treatment. *Handbook of Clinical Neurology*, 122, 173–189. <https://doi.org/10.1016/B978-0-444-52001-2.00008-X>
- Robinson, M. D., McCarthy, D. J., & Smyth, G. K. (2010). edgeR: a Bioconductor package for differential expression analysis of digital gene expression data. *Bioinformatics*, 26(1), 139–140. <https://doi.org/10.1093/bioinformatics/btp616>
- Rodriguez-Arellano, J. J., Parpura, V., Zorec, R., & Verkhratsky, A. A. (2016). Astrocytes in physiological aging and alzheimer's disease. *Neuroscience*, 323, 170–182. <https://doi.org/10.1016/j.neuroscience.2015.01.007>
- Rohrer, J. D. (2012). Structural brain imaging in frontotemporal dementia. *Biochimica et Biophysica Acta*, 1822(3), 325–332. <https://doi.org/10.1016/J.BBADIS.2011.07.014>
- Rosenstein, J. M., Krum, J. M., & Ruhrberg, C. (2010). VEGF in the nervous system. *Organogenesis*, 6(2), 107–114. <https://doi.org/10.4161/org.6.2.11687>
- Rosenthal, J. F., Hoffman, B. M., & Tyor, W. R. (2020). CNS inflammatory demyelinating disorders: MS, NMOSD and MOG antibody associated disease. *Journal of Investigative Medicine*, 68(2), 321–330. <https://doi.org/10.1136/jim-2019-001126>
- Rott, O., Fleischer, B., & Cash, E. (1994). Interleukin-10 prevents experimental allergic encephalomyelitis in rats. *European Journal of Immunology*, 24(6), 1434–1440. <https://doi.org/10.1002/eji.1830240629>
- Roussotte, F. F., Gutman, B. A., Madsen, S. K., Colby, J. B., Thompson, P. M., & Alzheimer's Disease Neuroimaging Initiative. (2014). Combined effects of Alzheimer risk variants in the CLU and ApoE genes on ventricular expansion patterns in the elderly. *The Journal of Neuroscience*, 34(19), 6537–6545. <https://doi.org/10.1523/JNEUROSCI.5236-13.2014>
- Ryu, J. K., & McLarnon, J. G. (2009). A leaky blood-brain barrier, fibrinogen infiltration and microglial reactivity in inflamed Alzheimer's disease brain. *Journal of Cellular and Molecular Medicine*, 13(9A), 2911–2925. <https://doi.org/10.1111/j.1582-4934.2008.00434.x>
- Sabelström, H., Stenudd, M., Réu, P., Dias, D. O., Elfineh, M., Zdunek, S., ... Frisén, J. (2013). Resident neural stem cells restrict tissue damage and neuronal loss after spinal cord injury in mice. *Science*, 342(6158), 637–640. <https://doi.org/10.1126/SCIENCE.1242576>
- Safaiyan, S., Besson-Girard, S., Gberk Kaya, T., Brendel, M., Gokce, O., & Simons Correspondence, M. (2021). White matter aging drives microglial diversity. *Neuron*, 109(7),

100–1117. <https://doi.org/10.1016/j.neuron.2021.01.027>

Saliba, A.-E., Westermann, A. J., Gorski, S. A., & Vogel, J. (2014). Single-cell RNA-seq: advances and future challenges. *Nucleic Acids Research*, 42(14), 8845–8860. <https://doi.org/10.1093/nar/gku555>

Salvadores, N., Gerónimo-Olvera, C., & Court, F. A. (2020). Axonal degeneration in AD: the contribution of A β and tau. *Frontiers in Aging Neuroscience*, 12, 319. <https://doi.org/10.3389/fnagi.2020.581767>

Sanz, E., Bean, J. C., Carey, D. P., Quintana, A., & McKnight, G. S. (2019). RiboTag: ribosomal tagging strategy to analyze cell-type-specific mRNA expression in vivo. *Current Protocols in Neuroscience*, 88(1), e77. <https://doi.org/10.1002/cpns.77>

Saunders, A., Macosko, E. Z., Wysoker, A., Goldman, M., Krienen, F. M., de Rivera, H., ... McCarroll, S. A. (2018). Molecular diversity and specializations among the cells of the adult mouse brain. *Cell*, 174(4), 1015–1030. <https://doi.org/10.1016/j.cell.2018.07.028>

Schafer, D. P., Lehrman, E. K., Kautzman, A. G., Koyama, R., Mardinly, A. R., Yamasaki, R., ... Stevens, B. (2012). Microglia sculpt postnatal neural circuits in an activity and complement-dependent manner. *Neuron*, 74(4), 691. <https://doi.org/10.1016/J.NEURON.2012.03.026>

Schirmer, L., Velmeshev, D., Holmqvist, S., Kaufmann, M., Werneburg, S., Jung, D., ... Rowitch, D. H. (2019). Neuronal vulnerability and multilineage diversity in multiple sclerosis. *Nature*, 573(7772), 75–82. <https://doi.org/10.1038/s41586-019-1404-z>

Schläger, C., Körner, H., Krueger, M., Vidoli, S., Haberl, M., Mielke, D., ... Flügel, A. (2016). Effector T-cell trafficking between the leptomeninges and the cerebrospinal fluid. *Nature*, 530(7590), 349–353. <https://doi.org/10.1038/nature16939>

Schmid, K. T., Cruceanu, C., Böttcher, A., Lickert, H., Binder, E. B., Theis, F. J., & Heinig, M. (2020). Design and power analysis for multi-sample single cell genomics experiments. *BioRxiv*. <https://doi.org/10.1101/2020.04.01.019851>

Schorlemmer, H. U., & Seiler, F. R. (1991). 15-Deoxyspergualin (15-DOS) for therapy in an animal model of multiple sclerosis (MS): disease modifying activity on acute and chronic relapsing experimental allergic encephalomyelitis (EAE). *Agents and Actions*, 34(1–2), 156–160. <https://doi.org/10.1007/BF01993265>

Seelaar, H., Rohrer, J. D., Pijnenburg, Y. A. L., Fox, N. C., & van Swieten, J. C. (2011). Clinical, genetic and pathological heterogeneity of frontotemporal dementia: a review. *Journal of Neurology, Neurosurgery, and Psychiatry*, 82(5), 476–486. <https://doi.org/10.1136/jnnp.2010.212225>

Seilhean, D., Le Ber, I., Sarazin, M., Lacomblez, L., Millicamps, S., Salachas, F., ... Duyckaerts, C. (2011). Fronto-temporal lobar degeneration: neuropathology in 60 cases. *Journal of Neural Transmission*, 118(5), 753–764. <https://doi.org/10.1007/s00702-011-0649-y>

Selewa, A., Dohn, R., Eckart, H., Lozano, S., Xie, B., Gauchat, E., ... Basu, A. (2020). Systematic comparison of high-throughput single-cell and single-nucleus transcriptomes during cardiomyocyte differentiation. *Scientific Reports*, 10(1), 1535. <https://doi.org/10.1038/s41598-020-58327-6>

Sengillo, J. D., Winkler, E. A., Walker, C. T., Sullivan, J. S., Johnson, M., & Zlokovic, B. V. (2013). Deficiency in mural vascular cells coincides with blood-brain barrier disruption in

- Alzheimer's disease. *Brain Pathology*, 23(3), 303–310. <https://doi.org/10.1111/bpa.12004>
- Shibata, M., Yamada, S., Kumar, S. R., Calero, M., Bading, J., Frangione, B., ... Zlokovic, B. V. (2000). Clearance of Alzheimer's amyloid-ss(1-40) peptide from brain by LDL receptor-related protein-1 at the blood-brain barrier. *The Journal of Clinical Investigation*, 106(12), 1489–1499. <https://doi.org/10.1172/JCI10498>
- Sierksma, A., Lu, A., Mancuso, R., Fattorelli, N., Thrupp, N., Salta, E., ... Fiers, M. (2020). Novel Alzheimer risk genes determine the microglia response to amyloid- β but not to TAU pathology. *EMBO Molecular Medicine*, 12(3), e10606. <https://doi.org/10.15252/emmm.201910606>
- Sild, M., & Ruthazer, E. S. (2011). Radial glia: progenitor, pathway, and partner. *The Neuroscientist*, 17(3), 288–302. <https://doi.org/10.1177/1073858410385870>
- Simpson, J. E., Ince, P. G., Lace, G., Forster, G., Shaw, P. J., Matthews, F., ... Wharton, S. B. (2010). Astrocyte phenotype in relation to Alzheimer-type pathology in the ageing brain. *Neurobiology of Aging*, 31, 578–590. <https://doi.org/10.1016/j.neurobiolaging.2008.05.015>
- Singh, V. P., Pratap, K., Sinha, J., Desiraju, K., Bahal, D., & Kukreti, R. (2016). Critical evaluation of challenges and future use of animals in experimentation for biomedical research. *International Journal of Immunopathology and Pharmacology*, 29(4), 551–561. <https://doi.org/10.1177/0394632016671728>
- Sjöstedt, E., Zhong, W., Fagerberg, L., Karlsson, M., Mitsios, N., Adori, C., ... Mulder, J. (2020). An atlas of the protein-coding genes in the human, pig, and mouse brain. *Science*, 367(6482). <https://doi.org/10.1126/science.aay5947>
- Skene, N. G., & Grant, S. G. N. (2016). Identification of vulnerable cell types in major brain disorders using single cell transcriptomes and expression weighted cell type enrichment. *Frontiers in Neuroscience*, 10, 16. <https://doi.org/10.3389/fnins.2016.00016>
- Smith, K. R., Damiano, J., Franceschetti, S., Carpenter, S., Canafoglia, L., Morbin, M., ... Berkovic, S. F. (2012). Strikingly different clinicopathological phenotypes determined by progranulin-mutation dosage. *American Journal of Human Genetics*, 90(6), 1102–1107. <https://doi.org/10.1016/j.ajhg.2012.04.021>
- Sobue, A., Komine, O., Hara, Y., Endo, F., Mizoguchi, H., Watanabe, S., ... Yamanaka, K. (2021). Microglial gene signature reveals loss of homeostatic microglia associated with neurodegeneration of Alzheimer's disease. *Acta Neuropathologica Communications*, 9(1), 1. <https://doi.org/10.1186/s40478-020-01099-x>
- Somjen, G. G. (1988). Nervenkit: notes on the history of the concept of neuroglia. *Glia*, 1(1), 2–9. <https://doi.org/10.1002/glia.440010103>
- Song, H. W., Foreman, K. L., Gastfriend, B. D., Kuo, J. S., Palecek, S. P., & Shusta, E. V. (2020). Transcriptomic comparison of human and mouse brain microvessels. *Scientific Reports*, 10(1), 12358. <https://doi.org/10.1038/s41598-020-69096-7>
- Sørensen, T. L., Tani, M., Jensen, J., Pierce, V., Lucchinetti, C., Folcik, V. A., ... Ransohoff, R. M. (1999). Expression of specific chemokines and chemokine receptors in the central nervous system of multiple sclerosis patients. *The Journal of Clinical Investigation*, 103(6), 807–815. <https://doi.org/10.1172/JCI15150>
- Sousa, C., Golebiewska, A., Poovathingal, S. K., Kaoma, T., Pires-Afonso, Y., Martina,

- S., ... Michelucci, A. (2018). Single-cell transcriptomics reveals distinct inflammation-induced microglia signatures. *EMBO Reports*, 19(11), e46171. <https://doi.org/10.15252/embr.201846171>
- Srinivasan, K., Friedman, B. A., Larson, J. L., Lauffer, B. E., Goldstein, L. D., Appling, L. L., ... Hansen, D. V. (2016). Untangling the brain's neuroinflammatory and neurodegenerative transcriptional responses. *Nature Communications*, 7, 11295. <https://doi.org/10.1038/ncomms11295>
- Sriram, S., & Steiner, I. (2005). Experimental allergic encephalomyelitis: a misleading model of multiple sclerosis. *Annals of Neurology*, 58(6), 939–945. <https://doi.org/10.1002/ana.20743>
- Ståhl, P. L., Salmén, F., Vickovic, S., Lundmark, A., Navarro, J. F., Magnusson, J., ... Frisén, J. (2016). Visualization and analysis of gene expression in tissue sections by spatial transcriptomics. *Science*, 353(6294), 78–82. <https://doi.org/10.1126/science.aaf2403>
- Stark, R., Grzelak, M., & Hadfield, J. (2019). RNA sequencing: the teenage years. *Nature Reviews Genetics*, 20(11), 631–656. <https://doi.org/10.1038/s41576-019-0150-2>
- Stoeckius, M., Hafemeister, C., Stephenson, W., Houck-Loomis, B., Chattopadhyay, P. K., Swerdlow, H., ... Smibert, P. (2017). Simultaneous epitope and transcriptome measurement in single cells. *Nature Methods*, 14(9), 865–868. <https://doi.org/10.1038/nmeth.4380>
- Stoffels, J. M. J., de Jonge, J. C., Stancic, M., Nomden, A., van Strien, M. E., Ma, D., ... Baron, W. (2013). Fibronectin aggregation in multiple sclerosis lesions impairs remyelination. *Brain*, 136(1), 116–131. <https://doi.org/10.1093/brain/aws313>
- Storck, S. E., Meister, S., Nahrath, J., Meißner, J. N., Schubert, N., Di Spiezio, A., ... Pietrzik, C. U. (2016). Endothelial LRP1 transports amyloid- β (1-42) across the blood-brain barrier. *The Journal of Clinical Investigation*, 126(1), 123–136. <https://doi.org/10.1172/JCI81108>
- Stuart, T., Butler, A., Hoffman, P., Stoeckius, M., Smibert, P., Satija, R., ... Hao, Y. (2019). Comprehensive integration of single-cell data. *Cell*, 177, 1888–1902. <https://doi.org/10.1016/j.cell.2019.05.031>
- Südhof, T. C. (2017). Synaptic neurexin complexes: a molecular code for the logic of neural circuits. *Cell*, 171(4), 745–769. <https://doi.org/10.1016/j.cell.2017.10.024>
- Tanaka, Y., Matsuwaki, T., Yamanouchi, K., & Nishihara, M. (2013). Exacerbated inflammatory responses related to activated microglia after traumatic brain injury in progranulin-deficient mice. *Neuroscience*, 231, 49–60. <https://doi.org/10.1016/j.neuroscience.2012.11.032>
- Tang, F., Barbacioru, C., Wang, Y., Nordman, E., Lee, C., Xu, N., ... Surani, M. A. (2009a). mRNA-Seq whole-transcriptome analysis of a single cell. *Nature Methods*, 6, 377. <https://doi.org/10.1038/nmeth.1315>
- Tang, F., Barbacioru, C., Wang, Y., Nordman, E., Lee, C., Xu, N., ... Surani, M. A. (2009b). mRNA-Seq whole-transcriptome analysis of a single cell. *Nature Methods*, 6(5), 377–382. <https://doi.org/10.1038/nmeth.1315>
- Tangkeangsirisin, W., & Serrero, G. (2004). PC cell-derived growth factor (PCDGF/GP88, progranulin) stimulates migration, invasiveness and VEGF expression in breast cancer cells. *Carcinogenesis*, 25(9), 1587–1592. <https://doi.org/10.1093/carcin/bgh171>
- Tann, J. Y., Wong, L., Sajikumar, S., & Ibáñez, C. F. (2019). Abnormal TDP-43 function impairs

- activity-dependent BDNF secretion, synaptic plasticity, and cognitive behavior through altered Sortilin splicing. *The EMBO Journal*, 38(5). <https://doi.org/10.15252/embj.2018100989>
- Tasic, B., Yao, Z., Graybiel, L. T., Smith, K. A., Nguyen, T. N., Bertagnoli, D., ... Zeng, H. (2018). Shared and distinct transcriptomic cell types across neocortical areas. *Nature*, 563(7729), 72–78. <https://doi.org/10.1038/s41586-018-0654-5>
- Tekman, M., Batut, B., Ostrovsky, A., Antoniewski, C., Clements, D., Ramirez, F., ... Grüning, B. (2020). A single-cell RNA-seq training and analysis suite using the Galaxy framework. *BioRxiv*, 2020.06.06.137570. <https://doi.org/10.1101/2020.06.06.137570>
- Török, O., Schreiner, B., Schaffenrath, J., Tsai, H.-C., Maheshwari, U., Stifter, S. A., ... Keller, A. (2021). Pericytes regulate vascular immune homeostasis in the CNS. *Proceedings of the National Academy of Sciences*, 118(10), e2016587118. <https://doi.org/10.1073/pnas.2016587118>
- Traugott, U. (1987). Multiple sclerosis: relevance of class I and class II MHC-expressing cells to lesion development. *Journal of Neuroimmunology*, 16(2), 283–302. [https://doi.org/10.1016/0165-5728\(87\)90082-8](https://doi.org/10.1016/0165-5728(87)90082-8)
- Trotter, J. H., Dargaei, Z., Wöhr, M., Liakath-Ali, K., Raju, K., Essayan-Perez, S., ... Südhof, T. C. (2020). Astrocytic neuroligin-1 orchestrates functional synapse assembly. *BioRxiv*, 2020.08.21.262097. <https://doi.org/10.1101/2020.08.21.262097>
- Umoh, M. E., Dammer, E. B., Dai, J., Duong, D. M., Lah, J. J., Levey, A. I., ... Seyfried, N. T. (2018). A proteomic network approach across the ALS-FTD disease spectrum resolves clinical phenotypes and genetic vulnerability in human brain. *EMBO Molecular Medicine*, 10(1), 48–62. <https://doi.org/10.15252/emmm.201708202>
- Ulvestad, E., Williams, K., Vedeler, C., Antel, J., Nyland, H., Mørk, S., & Matre, R. (1994). Reactive microglia in multiple sclerosis lesions have an increased expression of receptors for the Fc part of IgG. *Journal of the Neurological Sciences*, 121(2), 125–131. [https://doi.org/10.1016/0022-510x\(94\)90340-9](https://doi.org/10.1016/0022-510x(94)90340-9)
- Vågberg, M., Norgren, N., Dring, A., Lindqvist, T., Birgander, R., Zetterberg, H., & Svenningsson, A. (2015). Levels and age dependency of neurofilament light and glial fibrillary acidic protein in healthy individuals and their relation to the brain parenchymal fraction. *PLOS ONE*, 10(8), e0135886. <https://doi.org/10.1371/journal.pone.0135886>
- Van den Berge, K., Perraudeau, F., Soneson, C., Love, M. I., Risso, D., Vert, J.-P., ... Clement, L. (2018). Observation weights unlock bulk RNA-seq tools for zero inflation and single-cell applications. *Genome Biology*, 19(1), 24. <https://doi.org/10.1186/s13059-018-1406-4>
- van den Bos, H., Spierings, D. C. J., Taudt, A., Bakker, B., Porubský, D., Falconer, E., ... Lansdon, P. M. (2016). Single-cell whole genome sequencing reveals no evidence for common aneuploidy in normal and Alzheimer's disease neurons. *Genome Biology*, 17(1), 116. <https://doi.org/10.1186/s13059-016-0976-2>
- van der Poel, M., Ulas, T., Mizze, M. R., Hsiao, C.-C., Miedema, S. S. M., Adelia, ... Huitinga, I. (2019). Transcriptional profiling of human microglia reveals grey-white matter heterogeneity and multiple sclerosis-associated changes. *Nature Communications*, 10(1), 1139. <https://doi.org/10.1038/s41467-019-08976-7>
- van Swieten, J. C., & Heutink, P. (2008). Mutations in progranulin (GRN) within the spectrum of clinical and pathological phenotypes of frontotemporal dementia. *The Lancet Neurology*,

7(10), 965–974. [https://doi.org/10.1016/S1474-4422\(08\)70194-7](https://doi.org/10.1016/S1474-4422(08)70194-7)

van Wageningen, T. A., Vlaar, E., Kooij, G., Jongenelen, C. A. M., Geurts, J. J. G., & van Dam, A.-M. (2019). Regulation of microglial TMEM119 and P2RY12 immunoreactivity in multiple sclerosis white and grey matter lesions is dependent on their inflammatory environment. *Acta Neuropathologica Communications*, 7(1), 206. <https://doi.org/10.1186/s40478-019-0850-z>

Vanlandewijck, M., He, L., Mäe, M. A., Andrae, J., Ando, K., Del Gaudio, F., ... Betsholtz, C. (2018). A molecular atlas of cell types and zonation in the brain vasculature. *Nature*, 554(7693), 475–480. <https://doi.org/10.1038/nature25739>

Velmeshev, D., Schirmer, L., Jung, D., Haeussler, M., Perez, Y., Mayer, S., ... Kriegstein, A. R. (2019). Single-cell genomics identifies cell type-specific molecular changes in autism. *Science*, 364(6441), 685–689. <https://doi.org/10.1126/science.aav8130>

Vickers, J. C., Dickson, T. C., Adlard, P. A., Saunders, H. L., King, C. E., & McCormack, G. (2000). The cause of neuronal degeneration in Alzheimer's disease. *Progress in Neurobiology*, 60(2), 139–165. [https://doi.org/10.1016/s0301-0082\(99\)00023-4](https://doi.org/10.1016/s0301-0082(99)00023-4)

Vieth, B., Ziegenhain, C., Parekh, S., Enard, W., & Hellmann, I. (2017). powsimR: power analysis for bulk and single cell RNA-seq experiments. *Bioinformatics*, 33(21), 3486–3488. <https://doi.org/10.1093/bioinformatics/btx435>

Wageningen, T. A., Gerrits, E., Palacin i Bonson, S., Huitinga, I., Eggen, B. J. L., & Dam, A. (2021). Exploring reported genes of microglia RNA -sequencing data: Uses and considerations. *Glia*, 69(12), 2933–2946. <https://doi.org/10.1002/glia.24078>

Waller, R., Woodroffe, M. N., Wharton, S. B., Ince, P. G., Francese, S., Heath, P. R., ... Simpson, J. E. (2016). Gene expression profiling of the astrocyte transcriptome in multiple sclerosis normal appearing white matter reveals a neuroprotective role. *Journal of Neuroimmunology*, 299, 139–146. <https://doi.org/10.1016/j.jneuroim.2016.09.010>

Wang, C., Sun, B., Zhou, Y., Grubb, A., & Gan, L. (2012). Cathepsin B degrades amyloid- β in mice expressing wild-type human amyloid precursor protein. *The Journal of Biological Chemistry*, 287(47), 39834–39841. <https://doi.org/10.1074/jbc.M112.371641>

Wang, E. T., Sandberg, R., Luo, S., Khrebtkova, I., Zhang, L., Mayr, C., ... Burge, C. B. (2008). Alternative isoform regulation in human tissue transcriptomes. *Nature*, 456(7221), 470–476. <https://doi.org/10.1038/nature07509>

Ward, M. E., Chen, R., Huang, H.-Y., Ludwig, C., Telpoukhovskaia, M., Taubes, A., ... Green, A. J. (2017). Individuals with progranulin haploinsufficiency exhibit features of neuronal ceroid lipofuscinosis. *Science Translational Medicine*, 9(385). <https://doi.org/10.1126/scitranslmed.aah5642>

Watanabe, M., Toyama, Y., & Nishiyama, A. (2002). Differentiation of proliferated NG2-positive glial progenitor cells in a remyelinating lesion. *Journal of Neuroscience Research*, 69(6), 826–836. <https://doi.org/10.1002/jnr.10338>

Watkins, T. A., Emery, B., Mulinyawe, S., & Barres, B. A. (2008). Distinct stages of myelination regulated by gamma-secretase and astrocytes in a rapidly myelinating CNS coculture system. *Neuron*, 60(4), 555–569. <https://doi.org/10.1016/j.neuron.2008.09.011>

Weidner, L. D., Kannan, P., Mitsios, N., Kang, S. J., Hall, M. D., Theodore, W. H., ... Mulder, J. (2018). The expression of inflammatory markers and their potential influence on efflux

- transporters in drug-resistant mesial temporal lobe epilepsy tissue. *Epilepsia*, 59(8), 1507–1517. <https://doi.org/10.1111/epi.14505>
- Werkman, I. L., Lentferink, D. H., & Baron, W. (2021). Macroglial diversity: white and grey areas and relevance to remyelination. *Cellular and Molecular Life Sciences*, 78(1), 143–171. <https://doi.org/10.1007/s00018-020-03586-9>
- Wharton, S. B., O’Callaghan, J. P., Savva, G. M., Nicoll, J. A. R., Matthews, F., Simpson, J. E., ... MRC Cognitive Function and Ageing Neuropathology Study Group. (2009). Population variation in glial fibrillary acidic protein levels in brain ageing: relationship to Alzheimer-type pathology and dementia. *Dementia and Geriatric Cognitive Disorders*, 27(5), 465–473. <https://doi.org/10.1159/000217729>
- Wilkins, A., Majed, H., Layfield, R., Compston, A., & Chandran, S. (2003). Oligodendrocytes promote neuronal survival and axonal length by distinct intracellular mechanisms: a novel role for oligodendrocyte-derived glial cell line-derived neurotrophic factor. *The Journal of Neuroscience*, 23(12), 4967–4974. <https://doi.org/10.1523/JNEUROSCI.23-12-04967.2003>
- Winkler, E. A., Sengillo, J. D., Sullivan, J. S., Henkel, J. S., Appel, S. H., & Zlokovic, B. V. (2013). Blood-spinal cord barrier breakdown and pericyte reductions in amyotrophic lateral sclerosis. *Acta Neuropathologica*, 125(1), 111–120. <https://doi.org/10.1007/s00401-012-1039-8>
- Wisniewski, H. M., Vorbrod, A. W., & Wegiel, J. (1997). Amyloid angiopathy and blood-brain barrier changes in Alzheimer’s disease. *Annals of the New York Academy of Sciences*, 826, 161–172. <https://doi.org/10.1111/j.1749-6632.1997.tb48468.x>
- Witcher, M. R., Park, Y. D., Lee, M. R., Sharma, S., Harris, K. M., & Kirov, S. A. (2010). Three-dimensional relationships between perisynaptic astroglia and human hippocampal synapses. *Glia*, 58(5), 572–587. <https://doi.org/10.1002/glia.20946>
- Wlodarczyk, A., Holtman, I. R., Krueger, M., Yogev, N., Bruttger, J., Khoroshi, R., ... Owens, T. (2017). A novel microglial subset plays a key role in myelinogenesis in developing brain. *The EMBO Journal*, 36(22), 3292–3308. <https://doi.org/10.15252/embj.201696056>
- Wolf, Y., Shemer, A., Levy-Efrati, L., Gross, M., Kim, J.-S., Engel, A., ... Jung, S. (2018). Microglial MHC class II is dispensable for experimental autoimmune encephalomyelitis and cuprizone-induced demyelination. *European Journal of Immunology*, 48(8), 1308–1318. <https://doi.org/10.1002/eji.201847540>
- Wolock, S. L., Lopez, R., & Klein, A. M. (2019). Scrublet: computational identification of cell doublets in single-Cell transcriptomic data. *Cell Systems*, 8(4), 281–291.e9. <https://doi.org/10.1016/J.CELS.2018.11.005>
- Woollacott, I. O. C., & Rohrer, J. D. (2016). The clinical spectrum of sporadic and familial forms of frontotemporal dementia. *Journal of Neurochemistry*, 138, 6–31. <https://doi.org/10.1111/jnc.13654>
- Xi, S., Gibilisco, L., Kummer, M., Biber, K., Wachter, A., & Woodbury, M. (2020). ABACUS: A flexible UMI counter that leverages intronic reads for single-nucleus RNAseq analysis. *BioRxiv*, 2020.11.13.381624. <https://doi.org/10.1101/2020.11.13.381624>
- Yamazaki, Y., Shinohara, M., Yamazaki, A., Murray, M. E., Liesinger, A. M., ... Bu, G. (2019). Selective loss of cortical endothelial tight junction proteins during Alzheimer’s disease progression. *Brain*, 142(4), 1077–1092. <https://doi.org/10.1093/brain/awz011>

- Yang, T.-T., Lin, C., Hsu, C.-T., Wang, T.-F., Ke, F.-Y., & Kuo, Y.-M. (2013). Differential distribution and activation of microglia in the brain of male C57BL/6J mice. *Brain Structure and Function*, 218(4), 1051–1060. <https://doi.org/10.1007/s00429-012-0446-x>
- Yao, Z., van Velthoven, C. T. J., Nguyen, T. N., Goldy, J., Sedeno-Cortes, A. E., Baftizadeh, F., ... Zeng, H. (2021). A taxonomy of transcriptomic cell types across the isocortex and hippocampal formation. *Cell*, 184(12), 3222–3241.e26. <https://doi.org/10.1016/j.cell.2021.04.021>
- Yednock, T. A., Cannon, C., Fritz, L. C., Sanchez-Madrid, F., Steinman, L., & Karin, N. (1992). Prevention of experimental autoimmune encephalomyelitis by antibodies against alpha 4 beta 1 integrin. *Nature*, 356(6364), 63–66. <https://doi.org/10.1038/356063a0>
- Yeh, C.-Y., Vadhvana, B., Verkhatsky, A., & Rodríguez, J. J. (2011). Early astrocytic atrophy in the entorhinal cortex of a triple transgenic animal model of Alzheimer's disease. *ASN Neuro*, 3(5), 271–279. <https://doi.org/10.1042/AN20110025>
- Yeh, T.-H., Lee, D. Y., Gianino, S. M., & Gutmann, D. H. (2009). Microarray analyses reveal regional astrocyte heterogeneity with implications for neurofibromatosis type 1 (NF1)-regulated glial proliferation. *Glia*, 57(11), 1239–1249. <https://doi.org/10.1002/glia.20845>
- Yeung, M. S. Y., Zdunek, S., Bergmann, O., Bernard, S., Salehpour, M., Alkass, K., ... Frisén, J. (2014). Dynamics of oligodendrocyte generation and myelination in the human brain. *Cell*, 159(4), 766–774. <https://doi.org/10.1016/j.cell.2014.10.011>
- Yin, F., Banerjee, R., Thomas, B., Zhou, P., Qian, L., Jia, T., ... Ding, A. (2010). Exaggerated inflammation, impaired host defense, and neuropathology in progranulin-deficient mice. *The Journal of Experimental Medicine*, 207(1), 117–128. <https://doi.org/10.1084/jem.20091568>
- Yona, S., Kim, K.-W., Wolf, Y., Mildner, A., Varol, D., Breker, M., ... Jung, S. (2013). Fate mapping reveals origins and dynamics of monocytes and tissue macrophages under homeostasis. *Immunity*, 38(1), 79–91. <https://doi.org/10.1016/j.immuni.2012.12.001>
- Yu, C.-H., Yhee, J.-Y., Kim, J.-H., Im, K.-S., Kim, N.-H., Kwon, S.-Y., ... Sur, J.-H. (2012). Increased expression of vascular endothelial growth factor in neo-vascularized canine brain tissue. *Canadian Journal of Veterinary Research*, 76(1), 62–68. Retrieved from <http://www.ncbi.nlm.nih.gov/pubmed/22754097>
- Yuste, R., Hawrylycz, M., Aalling, N., Aguilar-Valles, A., Arendt, D., Armañanzas, R., ... Lein, E. (2020). A community-based transcriptomics classification and nomenclature of neocortical cell types. *Nature Neuroscience*, 23(12), 1456–1468. <https://doi.org/10.1038/s41593-020-0685-8>
- Zamboni, M., Llorens-Bobadilla, E., Magnusson, J. P., & Frisén, J. (2020). A widespread neurogenic potential of neocortical astrocytes is induced by injury. *Cell Stem Cell*, 27(4), 605–617. <https://doi.org/10.1016/j.stem.2020.07.006>
- Zeinstra, E., Wilczak, N., & De Keyser, J. (2003). Reactive astrocytes in chronic active lesions of multiple sclerosis express co-stimulatory molecules B7-1 and B7-2. *Journal of Neuroimmunology*, 135(1–2), 166–171. [https://doi.org/10.1016/s0165-5728\(02\)00462-9](https://doi.org/10.1016/s0165-5728(02)00462-9)
- Zeis, T., Graumann, U., Reynolds, R., & Schaeren-Wiemers, N. (2008). Normal-appearing white matter in multiple sclerosis is in a subtle balance between inflammation and neuroprotection. *Brain*, 131(1), 288–303. <https://doi.org/10.1093/brain/awm291>
- Zeisel, A., Hochgerner, H., Lönnerberg, P., Johnsson, A., Memic, F., van der Zwan, J., ...

- Linnarsson, S. (2018a). Molecular architecture of the mouse nervous system. *Cell*, 174(4), 999–1014. <https://doi.org/10.1016/j.cell.2018.06.021>
- Zeisel, A., Hochgerner, H., Lönnerberg, P., Johnsson, A., Memić, F., van der Zwan, J., ... Linnarsson, S. (2018b). Molecular architecture of the mouse nervous system. *Cell*, 174(4), 999–1014. <https://doi.org/10.1016/j.cell.2018.06.021>
- Zeisel, A., Muñoz-Manchado, A. B., Codeluppi, S., Lönnerberg, P., La Manno, G., Jureus, A., ... Linnarsson, S. (2015). Brain structure. Cell types in the mouse cortex and hippocampus revealed by single-cell RNA-seq. *Science*, 347(6226), 1138–1142. <https://doi.org/10.1126/science.aaa1934>
- Zeng, H., & Sanes, J. R. (2017). Neuronal cell-type classification: challenges, opportunities and the path forward. *Nature Reviews Neuroscience*, 18(9), 530–546. <https://doi.org/10.1038/nrn.2017.85>
- Zhan, J., Mann, T., Joost, S., Behrangi, N., Frank, M., & Kipp, M. (2020). The cuprizone model: dos and do dots. *Cells*, 9(4). <https://doi.org/10.3390/cells9040843>
- Zhang, J., Velmeshv, D., Hashimoto, K., Huang, Y.-H., Hofmann, J. W., Shi, X., ... Huang, E. J. (2020). Neurotoxic microglia promote TDP-43 proteinopathy in progranulin deficiency. *Nature*, 588, 459–465. <https://doi.org/10.1038/s41586-020-2709-7>
- Zhang, L., & Dimberg, A. (2016). Pleiotrophin is a driver of vascular abnormalization in glioblastoma. *Molecular & Cellular Oncology*, 3(6), e1141087. <https://doi.org/10.1080/23723556.2016.1141087>
- Zhang, Xiaochuan, Yin, X., Zhang, J., Li, A., Gong, H., Luo, Q., ... Jiang, H. (2019). High-resolution mapping of brain vasculature and its impairment in the hippocampus of Alzheimer's disease mice. *National Science Review*, 6(6), 1223–1238. <https://doi.org/10.1093/nsr/nwz124>
- Zhang, Xiaoming, Heng, Y., Kooistra, S. M., Weering, H. R. J., Brummer, M. L., Gerrits, E., ... Eggen, B. J. L. (2021). Intrinsic DNA damage repair deficiency results in progressive microglia loss and replacement. *Glia*, 69(3), 729–745. <https://doi.org/10.1002/glia.23925>
- Zhang, Ye, & Barres, B. A. (2010). Astrocyte heterogeneity: an underappreciated topic in neurobiology. *Current Opinion in Neurobiology*, 20(5), 588–594. <https://doi.org/10.1016/j.conb.2010.06.005>
- Zhang, Yuqing, Parmigiani, G., & Johnson, W. E. (2020). ComBat-seq: batch effect adjustment for RNA-seq count data. *NAR Genomics and Bioinformatics*, 2(3). <https://doi.org/10.1093/nargab/lqaa078>
- Zhao, Z., Sagare, A. P., Ma, Q., Halliday, M. R., Kong, P., Kisler, K., ... Zlokovic, B. V. (2015). Central role for PICALM in amyloid- β blood-brain barrier transcytosis and clearance. *Nature Neuroscience*, 18(7), 978–987. <https://doi.org/10.1038/nn.4025>
- Zhou, M., Tang, W., Fu, Y., Xu, X., Wang, Z., Lu, Y., ... Yi, F. (2015). Progranulin protects against renal ischemia/reperfusion injury in mice. *Kidney International*, 87(5), 918–929. <https://doi.org/10.1038/ki.2014.403>
- Zhou, Yingyao, Zhou, B., Pache, L., Chang, M., Khodabakhshi, A. H., Tanaseichuk, O., ... Chanda, S. K. (2019). Metascape provides a biologist-oriented resource for the analysis of systems-level datasets. *Nature Communications*, 10(1), 1523. <https://doi.org/10.1038/s41467-019-09234-6>

Zhou, Yingyue, Song, W. M., Andhey, P. S., Swain, A., Levy, T., Miller, K. R., ... Colonna, M. (2020). Human and mouse single-nucleus transcriptomics reveal TREM2-dependent and TREM2-independent cellular responses in Alzheimer's disease. *Nature Medicine*, 26(1), 131–142. <https://doi.org/10.1038/s41591-019-0695-9>

Abbreviations

AD	Alzheimer's disease
ALS	Amyotrophic lateral sclerosis
APOE	Apolipoprotein E
APP	Amyloid- β precursor protein
APPtg	APP transgenic
ARM	Activated response microglia
ATM	Axonal tract-associated microglia
ATP	Adenosine triphosphate
BBB	Blood brain barrier
BCSFB	Blood cerebrospinal fluid barrier
BDNF	Brain derived neurotrophic factor
BLMB	Blood leptomeningeal barrier
BSA	Bovine serum albumin
C9orf72	Chromosome 9 open reading frame 72
CA	Cornu ammonis
CAA	Cerebral amyloid angiopathy
CAM	CNS-associated macrophage
CLDN5	Claudin-5
CNS	Central nervous system
COP	Committed oligodendrocyte precursor
CPM	Counts per million
CSF	Cerebrospinal fluid
CTR	Control
CWM	Control white matter
DAM	Disease-associated microglia
DE	Differential expression
DEG	Differentially expressed gene
DG	Dentate gyrus
EAE	Experimental autoimmune encephalomyelitis
EC	Entorhinal cortex
EGLN1	Egl-9 family hypoxia inducible factor 1
EOAD	Early-onset Alzheimer's disease

EWCE	Expression weighted cell type enrichment
FACS	Fluorescence-activated cell sorting
FANS	Fluorescence-activated nuclei sorting
FC	Frontal cortex
FLT1	Fms related receptor tyrosine kinase 1
FPKM	Fragments per kilobase of transcript per million
FTD	Frontotemporal dementia
MS	Multiple sclerosis
FTD-GRN	Progranulin-associated frontotemporal dementia
GABA	Gamma-aminobutyric acid
GDNF	Glial derived neurotrophic factor
GM	Grey matter
GRN	Progranulin
GWAS	Genome wide association studies
HIF1	Hypoxia inducible factor 1
HLA-DR	Major histocompatibility complex class II, DR alpha
HSP	Heat shock protein
HVG	Highly variable gene
IEG	Immediate-early gene
IGF	Insulin-like growth factor
IHC	Immunohistochemistry
ISF	Interstitial fluid
ISS	In situ sequencing
LOAD	Late-onset Alzheimer's disease
LPS	Lipopolysaccharide
LRP1	LDL receptor related protein 1
MAPT	Microtubule associated protein tau
MFO	Myelin forming oligodendrocyte
MOG	Myelin oligodendrocyte glycoprotein
MOGAD	MOG-associated disease
MOL	Mature oligodendrocyte
MSC	Mesenchymal stem cell
NAWM	Normal appearing white matter

NBB	Netherlands brain bank
NCL	Neuronal ceroid lipofuscinosis
NFO	Newly formed oligodendrocyte
NMOSD	Neuromyelitis optica spectrum disorder
NS	Normal serum
OC	Occipital cortex
OPC	Oligodendrocyte progenitor cell
OTC	Occipitotemporal cortex
PCA	Principal component analysis
PIGF	Placental growth factor
PLP1	Proteolipid protein 1
PSEN	Presenilin-1
PSENtg	PSEN transgenic
PTN	Pleiotrophin
RNAseq	RNA-sequencing
scRNAseq	Single-cell RNA-sequencing
SMC	Smooth muscle cell
snRNAseq	Single-nucleus RNA-sequencing
ST	Spatial transcriptomics
TAUtg	Tau transgenic
TBI	Traumatic brain injury
TC	Temporal cortex
UMAP	Uniform manifold approximation and projection
UMI	Unique molecular identifier
WAM	White matter-associated microglia
WGCNA	Weighted gene coexpression network analysis
WM	White matter
WML	White matter lesion

English summary

The brain is a complex organ composed of a wide variety of cell types with intricate cellular organization, that is far from being completely understood. The human brain comprises over 100 billion neurons with an even larger number of supporting cells (glia) and a vascular architecture that is comprised of multiple distinct cell types. Understanding heterogeneity of the cells in the healthy brain and how these are affected by diseases will help reveal new targets for future therapies and biomarkers to monitor the disease. In this thesis, we studied cellular heterogeneity of the brain in relation to neurodegenerative diseases, in particular multiple sclerosis (MS), Alzheimer's disease (AD) and frontotemporal dementia (FTD).

Chapter 1 provides an overview of cell types in the brain and how these are affected by neurodegenerative diseases. Largely, cell types in the healthy brain can be divided into neurons, oligodendrocytes, astrocytes, microglia and CNS-associated macrophages (CAMs), and distinct cell types that comprise the brain's vasculature, including endothelial cells, pericytes, smooth muscle cells and fibroblasts. Neurodegenerative diseases are characterized by neurodegeneration, however often the disease mechanisms initially affect other cell types, and neuronal damage occurs as a secondary effect. For example, in MS it is known that oligodendrocytes are severely affected, resulting in loss of myelin that covers and protects axons, making the axons prone to damage and degeneration. In MS and AD, a significant role for microglia has been implied by genetic studies and dysfunction of these cells may possibly underlie neurodegeneration in these diseases. In FTD, studies with mouse models point towards microglia activation underlying neurodegeneration. However, in this thesis we show that this is not the case in humans.

In **chapter 2**, we studied brain tissues from MS donors. MS is a neurodegenerative disease, characterized by demyelinated lesions in the CNS, ultimately resulting in damage to axons which leads to cognitive problems, blindness, impaired motor function or even paralysis. We performed RNAseq on three groups of tissues: 1) white matter from control donors (CWM); 2) normal-appearing white matter (NAWM) from MS donors (non-demyelinated); 3) white matter lesions (WML) from MS donors. Between NAWM and WML tissues, most differentially expressed genes associated with myelination and inflammation, likely caused by the extensive demyelination that is present in WMLs and infiltration of immune cells. When comparing NAWM with CWM tissues, inflammation and stress pathways were enriched that pointed towards microglia and CAMs, indicating that these cells show reactive changes independent of demyelination. To investigate how microglia/CAMs are affected in MS tissues, we performed scRNAseq on CD45^{pos}CD11B^{pos} cells isolated from brain tissues from five MS donors. Of each donor, three brain regions were studied: 1) normal-appearing cortical tissue (NACT) which consisted of both grey and white matter; 2) NAWM; 3) WML. We identified eight subtypes of microglia/CAMs. One of these clusters showed enriched expression of immediate-early genes and heat shock proteins, and was particularly abundant in NAWM samples. Another cluster, that was enriched in WMLs, showed a typical activated/phagocytic microglia profile, that was similar to those described during developmental myelinogenesis. To confirm that this microglia phenotype arose in response to demyelination, we performed scRNAseq of microglia isolated from brains from mice fed with cuprizone. Indeed, we observed that a similar activated/phagocytic profile arose after demyelination, and disappeared in remyelinated mouse brains. These data indicate that microglia respond

to demyelination by adopting an activated/phagocytic profile, possibly as an attempt to stimulate (re)myelination. Thus, we showed that brain macrophages adopt distinct profiles in myelinated (NAWM) and demyelinated (WML) MS brain tissues, possibly representing an early stress response to MS pathology in NAWM and an activated/phagocytic phenotype accompanying demyelination.

Despite the valuable findings in **chapter 2**, there were also some disadvantages to the study design. Due to the dependence on fresh brain tissues for the cell isolation, we were able to include only five donors for scRNAseq. As an alternative approach, in **chapter 3** we set up a protocol to isolate nuclei from frozen brain tissues followed by snRNAseq. We validated, for microglia, that the nuclear transcriptome is a good proxy for the cellular transcriptome. Studying nuclei instead of cells allows to use archived, well characterized frozen brain tissues. These tissues can be obtained from brain banks and studied in large numbers at a time, avoiding confounding batch effects.

As a first large-scale application of snRNAseq in the lab, in **chapter 4** we studied brain tissues from CTR and late onset AD (LOAD) donors. AD is the most common type of dementia, and is neuropathologically characterized by accumulation of extracellular amyloid- β and intracellular tau. From CTR and LOAD donors, two brain regions were studied, where we specifically selected the LOAD cohort such that the occipital cortex (OC) only contained amyloid- β pathology and the occipitotemporal cortex (OTC) both amyloid- β and tau pathology. We studied the snRNAseq profiles of seven distinct cell types, and only in microglia disease-associated changes were observed. 12 subtypes of microglia were identified, which largely could be grouped into six categories: 'homeostatic', 'AD1', 'AD2', 'pro-inflammatory', 'stress' and 'proliferation'. The abundance of AD1 microglia correlated significantly with amyloid- β load in the tissues, and these nuclei had an activated/phagocytic gene expression profile similar to microglia in amyloid mouse models. The abundance of AD2 microglia correlated significantly with tau load, and these nuclei had a gene expression profile that suggested that these cells were more neurotrophic. These data indicate that microglia adopt distinct transcriptomic profiles in response to amyloid- β and tau pathology in LOAD, which offers new targets for microglia-state-specific therapeutic strategies.

In **chapter 5** we studied brain tissues from FTD-GRN donors. FTD is the second most common form of dementia, and neurodegeneration is typically present in the frontal and temporal lobes, whilst the posterior part of the brain (such as the occipital cortex) is seemingly unaffected. GRN is a growth factor that has been implicated in angiogenesis, wound healing, inflammation, brain development and, more recently, lysosomal function, a property mainly attributed to microglia. Heterozygous mutations in the *GRN* gene cause haploinsufficiency of the resulting protein and account for approximately 30% of genetic FTD cases. Homozygous deletion of *GRN* in humans causes a different disease, namely neuronal ceroid lipofuscinosis. Regardless, *Grn*^{-/-} mice are frequently studied as a model for FTD-GRN, where it was shown that microglia adopt reactive profiles, and this co-occurs with neurodegeneration. To investigate microglia profiles in FTD-GRN, we performed snRNAseq on frontal, temporal and occipital cortices from FTD-GRN and CTR donors. We did not observe profound microglia activation in FTD-GRN, neither in the RNAseq data nor *in situ* using IHC. Instead, we observed that astrocytes, endothelial cells, pericytes and fibroblasts were severely affected. Astrocytes adopted multiple profiles in FTD-GRN tissues,

characterized by enriched expression of genes associated with astrogliosis, BBB dysfunction, interferon signaling, oxidative stress and neuronal support. In endothelial cells, a depleted expression of homeostatic marker genes, such as *CLDN5*, *MFSD2A*, and *VEGF* was observed, as well as enriched expression of complement signaling and genes associated with BBB dysfunction. Additionally, we observed increased T-cell infiltration in FTD-GRN brain tissues *in situ*, possibly resulting from a dysfunctional BBB. Pericytes lost expression of homeostatic marker genes, and communication with endothelial cells. Fibroblasts were more abundant in FTD-GRN brain tissues, indicating that the tissue has become fibrotic. Taken together, we showed that not microglia, but the brain vasculature is severely affected in FTD-GRN brain tissues. In *Grn*^{-/-} mice, we observed that similar vascular changes occurred in 2-months old mice, whereas microgliosis and neurodegeneration occurred after 12 months of age. This indicates that the neurovascular findings in FTD-GRN underlie neurodegeneration, and are an essential feature of FTD-GRN pathophysiology. Hence, neurovascular changes should be considered as a novel and prime target for therapeutic interventions of FTD-GRN.

Chapter 6 summarizes the contribution of the findings reported in this thesis to the understanding of the pathological mechanisms of neurodegenerative diseases, and discusses potential pitfalls and future perspectives. In this thesis we used postmortem human brain tissues to investigate how different cells are affected by the underlying neurodegenerative pathology. Studying postmortem human brain tissues has several potential pitfalls, including limited tissue availability, studying end-stage disease and inability to modulate the disease process. On the other hand, alternative approaches, such as mouse models, often do not fully recapitulate the human disease, and mouse and human brains are significantly different. Additionally, a wide variety of technical factors may affect experimental outcomes and comparison of reported differentially expressed genes between studies revealed low overlap. Hence, one should be aware of confounding factors when interpreting scientific data.

Molecular biological techniques are advancing rapidly and in particular scRNAseq and snRNAseq have greatly enabled our ability to study cellular heterogeneity of the brain. The downside of sc/snRNAseq is loss of spatial information and potentially biased cell/nucleus isolation. With the advent of spatial transcriptomics techniques, where RNA expression is quantified *in situ*, transcriptomic analyses can be performed without prior isolation of cells or nuclei whilst retaining spatial information. This opens up many new avenues, in particular when the technique can be applied on a subcellular level. Potential examples of the application of this technique are mapping of cellular subtypes identified with sc/snRNAseq onto the tissue in relation to pathology (e.g. AD1 microglia with amyloid- β plaques), or studying cell-cell interactions. Lastly, open questions remain how induction of cellular subtypes identified with sc/snRNAseq is regulated, and whether these subtypes translate into functional subtypes. Multimodal studies, where transcriptomics is combined with epigenomics or proteomics may help answer these questions.

Nederlandse samenvatting

De hersenen zijn een complex orgaan dat bestaat uit een grote verscheidenheid aan celtypen met een ingewikkelde cellulaire organisatie, die nog lang niet volledig wordt begrepen. Het menselijk brein bestaat uit meer dan 100 miljard neuronen met een nog groter aantal ondersteunende cellen (glia) en een vasculaire architectuur die bestaat uit meerdere verschillende celtypen. Inzicht in de heterogeniteit van de cellen in de gezonde hersenen en hoe deze worden beïnvloed door ziekten, zal helpen bij het identificeren van nieuwe aanknopingspunten voor toekomstige therapieën en bio-markers om de ziekte te volgen. In dit proefschrift hebben we de cellulaire heterogeniteit van de hersenen bestudeerd in relatie tot neurodegeneratieve ziekten, in het bijzonder multiple sclerose (MS), de ziekte van Alzheimer (AD) en fronto-temporale dementie (FTD).

Hoofdstuk 1 geeft een overzicht van de verschillende celtypen in de hersenen en hoe deze worden beïnvloed door neurodegeneratieve ziekten. Grotendeels kunnen cellen in de gezonde hersenen worden onderverdeeld in neuronen, oligodendrocyten, astrocyten, microglia en CNS-geassocieerde macrofagen (CAM), en verschillende celtypen die het vaatstelsel van de hersenen vormen, waaronder endotheelcellen, pericyten, glad spierweefsel en fibroblasten. Neurodegeneratieve ziekten worden gekenmerkt door neurodegeneratie, maar vaak richten de ziektemechanismen zich in eerste instantie op andere celtypen, en is neuronale schade een secundair effect. Bij MS is het bijvoorbeeld bekend dat oligodendrocyten ernstig worden aangetast, wat leidt tot verlies van myeline dat axonen beschermt, waardoor de axonen vatbaar zijn voor beschadiging en degeneratie. Bij MS en AD is een significante rol voor microglia geïmpliceerd door genetische onderzoeken. Een verstoorde functie van deze cellen kan mogelijk ten grondslag liggen aan neurodegeneratie bij deze ziekten. In FTD wijzen studies met muismodellen in de richting van microglia-activering die ten grondslag ligt aan neurodegeneratie. In dit proefschrift laten we echter zien dat dit niet het geval is bij mensen.

In **hoofdstuk 2** hebben we hersenweefsel van MS-donoren bestudeerd. MS is een neurodegeneratieve ziekte, gekenmerkt door gedemyeliniseerde laesies in het centrale zenuwstelsel. Uiteindelijk leidt deze demyelinisatie tot schade aan axonen met cognitieve problemen, blindheid, verminderde motorische functie of zelfs verlamming als gevolg. We hebben met behulp van RNAseq de gen activiteit van drie groepen weefsels bepaald: 1) witte stof van controle-donoren (CWM); 2) normaal lijkende witte stof (NAWM) van MS-donoren (niet-gedemyeliniseerd); 3) witte stof laesies (WML) van MS-donoren. De meeste differentieel tot expressie gebrachte genen tussen NAWM en WML waren geassocieerd met myelinisatie en ontsteking, waarschijnlijk veroorzaakt door de uitgebreide demyelinisatie die aanwezig is in WMLs en de infiltratie van immuun cellen. Bij het vergelijken van de genexpressie profielen van NAWM met CWM-weefsels, waren ontstekings- en stressroutes verrijkt die wezen op betrokkenheid van microglia en CAMs, wat aangeeft dat deze cellen reactieve veranderingen vertonen, onafhankelijk van demyelinisatie. Om te onderzoeken hoe microglia/CAMs worden beïnvloed in MS-weefsels, hebben we het genexpressie niveau in individuele cellen bepaald door middel van single cell RNA sequencing (scRNAseq) van CD45^{pos}CD11B^{pos}-cellen die we hebben geïsoleerd uit hersenweefsel van vijf MS-donoren. Van elke donor werden drie hersengebieden bestudeerd: 1) normaal lijkend corticaal weefsel (NACT) dat bestond uit zowel grijze als witte stof; 2) NAWM; 3) WML. We identificeerden acht subtypes van microglia en CAMs. Een van deze subtypes vertoonde een verrijkte

expressie van 'immediate-early' genen en genen voor 'heat shock'-eiwitten, en was vooral aanwezig in NAWM-weefsels. Een ander cluster dat was verrijkt in WMLs, vertoonde een typisch geactiveerd/fagocytisch microglia profiel, dat vergelijkbaar was met het profiel dat is beschreven bij myelinisatie tijdens de ontwikkeling. Om te bevestigen dat dit microglia-fenotype ontstond als reactie op demyelinisatie, voerden we scRNAseq uit van microglia geïsoleerd uit de hersenen van muizen aan wiens dieet cuprizone was toegevoegd, een stof die leidt tot verlies van myeline. Inderdaad zagen we dat een vergelijkbaar geactiveerd/fagocytisch profiel ontstond na demyelinisatie en dit profiel verdween in geremyeliniseerde muizenhersenen. Deze bevindingen geven aan dat microglia reageren op demyelinisatie door een geactiveerd/fagocytisch profiel aan te nemen, mogelijk als een poging om myeline herstel te stimuleren. In deze studie hebben we aangetoond dat hersenmacrofagen verschillende profielen aannemen in gemyeliniseerde (NAWM) en gedemyeliniseerde (WML) MS-hersenenweefsels, wat mogelijk duidt op een vroege stressrespons op MS-pathologie in NAWM en een geactiveerd/fagocytisch fenotype na demyelinisatie.

Ondanks de waardevolle bevindingen in **hoofdstuk 2** waren er ook enkele nadelen aan de onderzoeksopzet. Vanwege de afhankelijkheid van vers hersenweefsel voor de cel isolatie, waren we in staat om slechts vijf donoren voor scRNAseq te includeren. Als alternatieve benadering hebben we in **hoofdstuk 3** een protocol geïmplementeerd om kernen te isoleren uit bevroren hersenweefsel, gevolgd door single nucleus RNA sequencing (snRNAseq). We hebben voor microglia gevalideerd dat het nucleaire transcriptoom een goede benadering is van het cellulaire transcriptoom. Door kernen in plaats van cellen te bestuderen, kunnen gearchiveerde, goed gekarakteriseerde bevroren hersenweefsels worden gebruikt. Deze weefsels kunnen worden verkregen uit hersenbanken en in grote aantallen tegelijk worden bestudeerd, waardoor batch-effecten worden vermeden.

Als een eerste grootschalige toepassing van snRNAseq in het laboratorium, hebben we in **hoofdstuk 4** hersenweefsel van CTR- en late-onset AD (LOAD)-donoren bestudeerd. AD is de meest voorkomende vorm van dementie en wordt neuropathologisch gekenmerkt door accumulatie van extracellulair amyloïde- β en intracellulair tau. Van CTR- en LOAD-donoren werden twee hersenregio's bestudeerd, waarbij we het LOAD-cohort dusdanig selecteerden dat de occipitale cortex (OC) alleen amyloïde- β -pathologie bevatte en de occipitotemporale cortex (OTC) zowel amyloïde- β - als tau-pathologie. We bestudeerden de snRNAseq-profielen van zeven verschillende celtypen en alleen in microglia werden ziekte gerelateerde veranderingen waargenomen. Er werden 12 subtypes van microglia geïdentificeerd, die grotendeels konden worden onderverdeeld in zes categorieën: 'homeostatisch', 'AD1', 'AD2', 'pro-inflammatoir', 'stress' en 'proliferatie'. De hoeveelheid van AD1-microglia correleerde significant met de hoeveelheid amyloïde- β in de weefsels, en deze kernen hadden een geactiveerd/fagocytisch genexpressieprofiel, vergelijkbaar met microglia in amyloïde muismodellen. De hoeveelheid AD2-microglia correleerde significant met de hoeveelheid tau en deze kernen hadden een genexpressieprofiel dat suggereerde dat deze microglia ondersteunend waren voor de neuronen. Deze gegevens suggereren dat microglia verschillende genexpressie profielen aannemen als reactie op amyloïde- β en tau-pathologie in LOAD, wat nieuwe aanknopingspunten geeft voor microglia-subtype specifieke therapeutische strategieën.

In **hoofdstuk 5** hebben we hersenweefsel van FTD-GRN donoren bestudeerd. FTD is de op een na (na AD) meest voorkomende vorm van dementie, en neurodegeneratie is typisch

aanwezig in de frontale en temporale kwabben, terwijl het achterste deel van de hersenen (zoals de occipitale kwab) gespaard blijft. GRN is een groeifactor die is betrokken bij angiogenese, wondgenezing, ontsteking, hersenontwikkeling en meer recentelijk lysosomale functies, een rol die voornamelijk wordt toegeschreven aan microglia. Heterozygote mutaties van het *GRN*-gen veroorzaken haplo-insufficiëntie van het resulterende eiwit en ongeveer 30% van de genetische FTD-gevallen hebben een *GRN* mutatie. Homozygote deletie van *GRN* bij mensen veroorzaakt een andere ziekte, neuronale ceroïde lipofuscinose. Desondanks worden *Grn*^{-/-} muizen vaak bestudeerd als een model voor FTD-GRN, en hierin werd aangetoond dat microglia reactieve profielen aannemen wat tegelijk gebeurt met neurodegeneratie. Om microglia-profielen in FTD-GRN te onderzoeken hebben we snRNAseq uitgevoerd op frontale, temporale en occipitale kwabben van FTD-GRN- en CTR-donoren. Hierin hebben we geen microglia-activering waargenomen in FTD-GRN, noch in de RNAseq-gegevens noch *in situ* met behulp van IHC. In plaats daarvan zagen we dat astrocyten, endotheelcellen, pericyten en fibroblasten extreme veranderingen hadden ondergaan. Astrocyten namen meerdere profielen aan in FTD-GRN-weefsels, gekenmerkt door verrijkte expressie van genen geassocieerd met astrogliose, bloed-hersen-barrière (BHB-)disfunctie, interferon-signalering, oxidatieve stress en neuronale ondersteuning. In endotheelcellen werd een verminderde expressie van homeostatische marker genen, zoals *CLDN5*, *MFS2A* en *VEGF*, waargenomen, evenals een verrijkte expressie van complementensignalering en genen geassocieerd met BHB-disfunctie. Bovendien zagen we verhoogde T-celinfiltratie in FTD-GRN-hersenweefsels *in situ*, mogelijk als gevolg van een disfunctionele BHB. Pericyten verloren expressie van homeostatische marker genen en communicatie met endotheelcellen. Fibroblasten waren meer aanwezig in FTD-GRN-hersenweefsels, wat aangeeft dat het weefsel mogelijk fibrotischer is geworden. Samenvattend toonden we aan dat niet microglia, maar de hersenvasculatuur ernstig was aangetast in FTD-GRN-hersenweefsel. Bij *Grn*^{-/-} muizen zagen we dat vergelijkbare vasculaire veranderingen optraden bij muizen van 2 maanden oud, terwijl microgliose en neurodegeneratie optraden bij een leeftijd van 12 maanden. Dit geeft aan dat de neurovasculaire bevindingen in FTD-GRN mogelijk ten grondslag liggen aan neurodegeneratie en een essentieel kenmerk zijn van FTD-GRN pathofysiologie. Daarom moeten neurovasculaire veranderingen worden beschouwd als een nieuw en belangrijk doelwit voor therapeutische interventies van FTD-GRN.

Hoofdstuk 6 vat de bijdrage van de in dit proefschrift beschreven bevindingen aan het begrijpen van de pathologische mechanismen van neurodegeneratieve ziekten samen, bespreekt mogelijke tekortkomingen en schetst toekomstperspectieven. In dit proefschrift hebben we postmortem menselijk hersenweefsel gebruikt om te onderzoeken hoe verschillende cellen worden beïnvloed door de onderliggende neurodegeneratieve pathologie. Het bestuderen van postmortaal menselijk hersenweefsel heeft verschillende valkuilen, waaronder beperkte weefselbeschikbaarheid, het bestuderen van ziekte in het eindstadium en het onvermogen om het ziekteproces experimenteel te beïnvloeden. Aan de andere kant recapituleren alternatieve benaderingen, zoals muismodellen, vaak niet volledig de ziekte, en de hersenen van muizen en mensen zijn significant verschillend. Bovendien kan een breed scala aan technische factoren de experimentele resultaten beïnvloeden en uit vergelijking van gerapporteerde differentieel tot expressie gebrachte genen tussen onderzoeken bleek er een lage reproduceerbaarheid te zijn. Daarom moet men zich bewust zijn van deze beperkingen bij het interpreteren van wetenschappelijke data.

Moleculaire biogietechnieken ontwikkelen zich snel en met name scRNAseq en snRNAseq hebben het bestuderen van cellulaire heterogeniteit van de hersenen mogelijk gemaakt. Het nadeel van sc/snRNAseq is het verlies van ruimtelijke informatie en mogelijk selectie bias door de gebruikte procedures voor cel/kern isolatie. Met de komst van 'spatial transcriptomics' technieken, waarbij RNA-expressie *in situ* wordt bestudeerd, kunnen genexpressie analyses worden uitgevoerd zonder voorafgaande isolatie van cellen of kernen, waarbij ruimtelijke informatie behouden blijft. Dit opent veel nieuwe wegen, vooral wanneer de techniek op sub cellulair niveau kan worden toegepast. Voorbeelden van toepassing van deze techniek zijn het in kaart brengen van cellulaire subtypes geïdentificeerd met sc/snRNAseq op het weefsel in relatie tot pathologie (bijv. lokalisatie van AD1 microglia met amyloïde- β plaques), of het bestuderen van cel-cel interacties. Ten slotte blijven er open vragen over hoe inductie van cellulaire subtypen geïdentificeerd met sc/snRNAseq wordt gereguleerd en of deze subtypen zich vertalen in functionele subtypen. Multimodale studies, waarbij genexpressie analyses worden gecombineerd met epigenetica of eiwit analyses, kunnen helpen bij het beantwoorden van deze vragen.

Publications and papers in progress

First authorship (this thesis)

Gerrits, E.*, Heng, Y.*, Boddeke, H.W.G.M., Eggen, B.J.L. (2020). Transcriptional profiling of microglia; current state of the art and future perspectives. *Glia*, 68(4), 740-755. <https://doi.org/10.1002/glia.23767>. (**Chapter 3**) *shared first authorship

Gerrits, E., Brouwer, N., Kooistra, S.M., Woodbury, M.E., Vermeiren, Y., Lambourne, M., Mulder, J., Kummer, M., Möller, T., Biber, K., den Dunnen, W.F.A., de Deyn, P.P., Eggen, B.J.L.*, Boddeke, H.W.G.M.* (2021). Distinct amyloid- β and tau-associated microglia profiles in Alzheimer's disease. *Acta Neuropathologica*, 141(5), 1-16. 2021; <https://doi.org/10.1007/s00401-021-02263-w>. (**Chapter 4**)

Gerrits, E.*, Miedema, A.*, Brouwer, N., Jiang, Q., Kracht, L., Meijer, M., Pijnacker, A.T.E., Wesseling, E., Wijering, M.H.C., Gabius, H-J., Amor, S., Eggen, B.J.L., Kooistra, S.M. (2022). Brain macrophages acquire distinct transcriptomic profiles in multiple sclerosis lesions and normal appearing white matter. *In press, Acta Neuropathologica Communications*. (**Chapter 2**) *shared first authorship

Gerrits, E., Giannini, L.A.A., Brouwer, N., Melhelm, S., Seilhean, D., Le Ber, I., Kamermans, A., Kooij, G., de Vries, H.E., Boddeke, H.W.G.M., Seelaar, H., van Swieten, J., Eggen, B.J.L. Neurovascular unit disruption underlies GRN-associated frontotemporal dementia. *In revision*. (**Chapter 5**)

Second authorship

Talma, N., **Gerrits, E.**, Wang, B., Eggen, B.J.L.*, Demaria, M.* (2021). Identification of distinct and age-dependent p16^{high} microglia subtypes. *Aging Cell*, 20(10), e13450. <https://doi.org/10.1111/accel.13450>.

Van Wageningen, T.A., **Gerrits, E.**, Palacin i Bonson, S., Huitinga, I., Eggen, B.J.L., van Dam, A.M. (2021). Exploring reported genes of microglia RNA-sequencing data: uses and considerations. *Glia*, 69(12), 2933-2946. <https://doi.org/10.1002/glia.24078>.

Van Wageningen, T.A., **Gerrits, E.**, Geleijnse, A., Brouwer, N., Geurts, J.G., Eggen, B.J.L., Boddeke, H.W.G.M., van Dam, A.M. (2022). Distinct gene expression profiles in leukocortical demyelinated white and grey matter areas of multiple sclerosis patients. *In press, Brain Communications*.

Other

Alsema, A.M.*, Jiang, Q.*, Kracht, L.*, **Gerrits, E.**, Dubbelaar, M., Miedema, A., Brouwer, N., Woodbury, M.E., Wachter, A., Xi, S., Moeller, M., Biber, K., Kooistra, S.M., Boddeke, H.W.G.M., Eggen, B.J.L. (2020). Profiling microglia from Alzheimer's disease donors and non-demented elderly in acute human post-mortem cortical tissue. *Frontiers of Molecular Neuroscience*, 13(134). <https://doi.org/10.3389/fnmol.2020.00134s>.

Zhang, X.*, Heng, Y.*, Kooistra, S.M., van Weering, H., Brummer, M., **Gerrits, E.**, Wesseling, E., Brouwer, N., Nijboer, T., Sierra, A., Dubbelaar, M.L., Boddeke, H.W.G.M., Eggen, B.J.L. (2020). Intrinsic DNA damage repair deficiency results in progressive microglia loss and replacement. *Glia*, 69(3), 729-745. <https://doi.org/10.1002/glia.23925>.

Amor, S., McNamara, N.B., **Gerrits, E.**, Marzin, M., Kooistra, S.M., Miron, V.E., Nutma, E. (2021). White matter microglia heterogeneity in the CNS. *Acta Neuropathologica*. <https://doi.org/10.1007/s00401-021-02389-x>.

Huizing, W., Dekker, S.L., van der Lienden, J.C.J, Mergener, R., Muszkopf, M.K., Furtado, G.V., **Gerrits, E.**, Coit, D., Oghbaie, M., Di Stefano, L.H., Schepers, H., van Waarde-Verhagen, M.A.W.H., Couzijn, S., Barazzuol, L., LaCava, J., Kampinga, H.H., Bergink, S. Targeting DNA topoisomerases or checkpoint kinases results in an overload of chaperone systems, triggering aggregation of a metastable subproteome. *In revision*.

Reitsema, V.A.* , Schreuder, L.* , **Gerrits, E.**, Eggen, B.J.L., Laman, J.D., de Rooij, S.E., Wesseling, E.M., Bouma, H.R., Henning, R.H. Calorie restriction increases vulnerability of progeroid *Erccl1^{Δ/-}* mice to acute (neuro)inflammation. *In revision*.

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