

University of Groningen

## The role of estradiol in the maintenance of brain-dead organ donors

Armstrong Junior, Roberto

DOI:  
[10.33612/diss.183298445](https://doi.org/10.33612/diss.183298445)

**IMPORTANT NOTE: You are advised to consult the publisher's version (publisher's PDF) if you wish to cite from it. Please check the document version below.**

*Document Version*  
Publisher's PDF, also known as Version of record

*Publication date:*  
2021

[Link to publication in University of Groningen/UMCG research database](#)

*Citation for published version (APA):*

Armstrong Junior, R. (2021). *The role of estradiol in the maintenance of brain-dead organ donors: from pathophysiology to treatment*. [Thesis fully internal (DIV), University of Groningen]. University of Groningen. <https://doi.org/10.33612/diss.183298445>

**Copyright**

Other than for strictly personal use, it is not permitted to download or to forward/distribute the text or part of it without the consent of the author(s) and/or copyright holder(s), unless the work is under an open content license (like Creative Commons).

The publication may also be distributed here under the terms of Article 25fa of the Dutch Copyright Act, indicated by the "Taverne" license. More information can be found on the University of Groningen website: <https://www.rug.nl/library/open-access/self-archiving-pure/taverne-amendment>.

**Take-down policy**

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

*Downloaded from the University of Groningen/UMCG research database (Pure): <http://www.rug.nl/research/portal>. For technical reasons the number of authors shown on this cover page is limited to 10 maximum.*

CHAPTER

1



# INTRODUCTION

Organ transplantation is the treatment of choice for patients with end-stage failure. In the past few decades, the success rate of transplants has progressively increased due to a better understanding of organ dysfunction, the rejection process, and the development of more appropriate immunosuppressive treatments (1–3).

Brain-dead donors are still the main source of transplantation organs. However, different situations compromise the viability of these organs (1,4), which highlights the importance of donor clinical conditions in the success rate of transplantation (5).

## **Brain death and organ transplantation**

Most of the organs used for transplantation come from donors who have been confirmed to be brain dead. Currently, the biggest challenge in transplantation surgery is the low number of viable organs. Technical advances in donor management and organ preservation have provided improved graft results with clear benefits to the recipient.

In addition to saving lives, heart, liver, and lung transplantations promote significant physical and social rehabilitation of patients, reintegrating them into society with a better quality of life. Kidney transplantations also promote resource savings, as alternative treatments for maintaining life, such as dialysis, are more expensive in the long run (1–3).

Clinical and experimental evidence highlight the influence of BD on the viability of the organs to be transplanted (6–8), pointing to the importance of the donor's physical status in the success rate of transplantation (5). Therefore, an understanding of the alterations caused by BD allows for the optimization of clinical conduct and the acquisition of organs considered viable for transplantation.

BD caused by trauma, infarction, or intracranial hemorrhage occurs due to increased intracranial pressure, and short, intense sympathetic activity. This period is marked by high hemodynamic instability in response to an increase in serum catecholamines (autonomic storm). Catecholamines are released from the adrenal glands and nerve endings, resulting in hypertension, tachycardia, and arrhythmia. However, after the release of catecholamines, there is an imbalance in the consumption and availability of oxygen in the heart, leading to ischemic damage (9,10).

In the lungs, the consequences of a large sympathetic stimulus are edema, hemorrhage, and increased capillary permeability (11).

After a catecholamine storm, an episode of compensatory arterial hypotension occurs. This is accompanied by intense sympathetic stimulation causing severe vasoconstriction, leading to an increase in systemic vascular resistance and tachycardia (12,13). In this phase, the perfusion of peripheral organs decreases, and an inflammatory and immune response develops as a consequence of ischemia, hypoperfusion, and edema. The result is a greater expression of inflammatory mediators, cytokines, chemokines, and adhesion molecules, accompanied by the infiltration of cells in all organs, such as the heart, lung, liver, and kidney (2,4,14).

Significant hormonal and metabolic disturbances occur following pituitary failure in experimental models of BD (15). Metabolic disorders are closely linked to the inflammatory process; therefore, these changes are associated with events that precede BD, such as trauma/hemorrhage, and add to the inflammatory response in the organ donor. The development of diabetes insipidus due to antidiuretic hormone reduction occurs in approximately 80% of organ donors. In addition, the significant reduction in cortisol concentration, associated with the interruption of adrenocorticotrophic hormone release by the anterior hypophysis, compromises the donor's endogenous response to the stress generated by BD (16).

BD should not be considered as a static situation, but as a dynamic process with a striking influence on the quality and viability of organs intended for transplantation (2,17). BD leads to gradual organ dysfunction, which is worsened by hemodynamic dysfunction. Additionally, BD triggers the inflammatory status of the potential donor organ, irrespective of the presence of hypotension. The disorders observed may induce the graft to supplementary damage from ischemia/reperfusion in the transplant procedure (2,18). Therefore, organs from donors with BD may possibly be immunologically active before transplantation, which may interfere with the result when comparing them to organs from living donors (1,10).

In the heart, BD induction through increased intracranial pressure leads to a hyperdynamic reaction, culminating in echocardiographic alterations and hemodynamic instability, decreasing myocardial isoenzymes and resulting in histological injury. These consequences of BD are associated with alterations in the expression of myocardial genes, in particular the activation of the  $\alpha$ -adrenergic receptor (19,20). The reduction in cardiac function in brain-dead donors may be caused by an imbalance between oxygen demand and supply to the myocardial tissue or even by direct injury caused by the massive release of catecholamines (21). One of the central causes of death following heart transplantation is primary graft dysfunction, which may be influenced by the donor's characteristics.

The records of the International Society for Heart and Lung Transplantation demonstrated that a sex mismatch between donor and recipient affects heart and lung transplant survival, especially after the first year (22). Other studies confirmed that the donor's gender influences the success of organ transplantation and demonstrated the differences in short-and long-term prognosis in various organ transplantations between sexes (23–27). Some surveys showed worse prognosis in heart and lung transplantation with grafts from women (27,28).

Experimental studies highlighted that acute reduction of female sex hormones caused by BD result in a negative effect on lung inflammation (29,30). Additionally, Simão et al. (31)

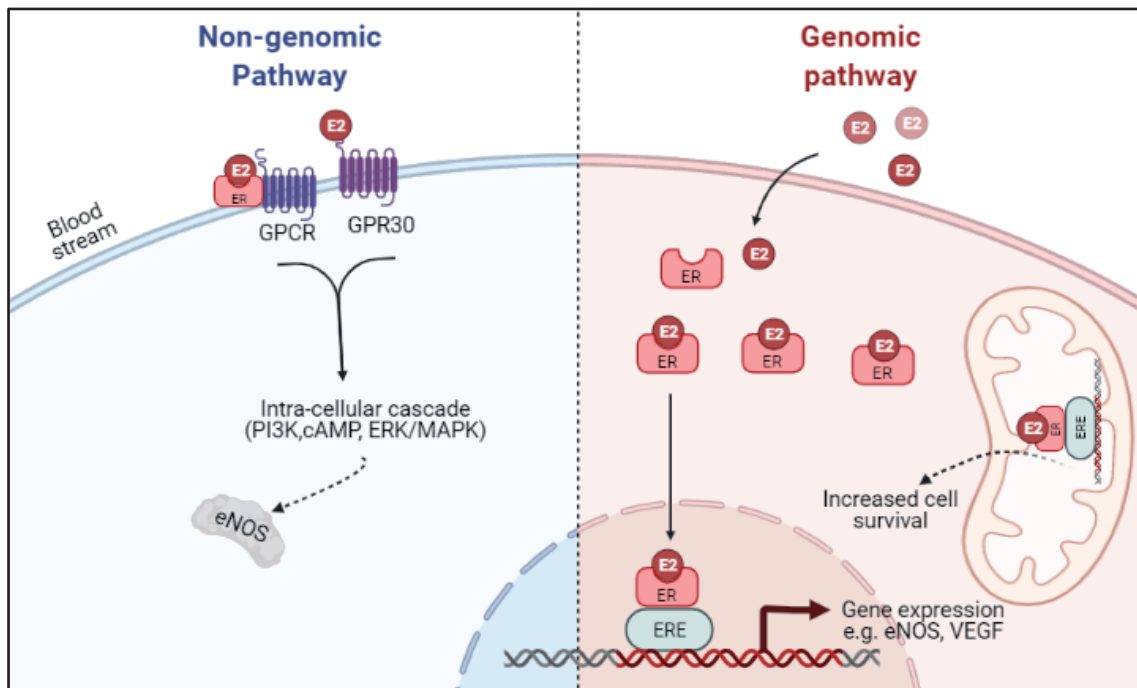
demonstrated inflammatory histological changes in the heart and lungs of female rats subjected to BD, with a remarkable increase in the number of polymorphonuclear and lymphomononuclear cells when compared to males. These data suggest that the inflammation generated by BD may be exacerbated in female animals and that female sex hormones, as modulators of the immune response, may be considered potential modulators of events triggered by BD.

### **Estradiol, immune system, and inflammation**

Previous studies have revealed that the repercussions of trauma are less intense in females than in males, who, under treatment with estradiol, also develop less intense injuries (32). In this context, females also have better preservation of hemodynamic parameters after hemorrhagic shock (32,33). Clinical studies show that women, when compared to men, have fewer cases of sepsis, pneumonia, and insufficiency of multiple organs and systems (34–37). Even in cases of cardiovascular disorders, sexually active women are more protected, with the most critical cases being described in postmenopausal women (38).

There is a growing number of investigations on the contribution of female sex hormones to inflammatory processes (39–41). Studies have demonstrated the participation of female sex hormones in modulating the magnitude of the inflammatory response, revealing the complex role of sexual steroids. The gonadal steroid  $17\beta$ -estradiol protects the cardiovascular system against ischemic, inflammatory, and metabolic injuries (39,40). In addition, prior experimental evidence clearly highlights that estrogen and estrogen receptor (ER) agonists are beneficial therapeutic adjuncts in preserving organ function and improving outcomes after trauma-hemorrhage (42).

The beneficial effects of estrogen in preserving organ function are due to a genomic action modulated by the interaction between intracellular receptors ER $\alpha$  and ER $\beta$ , as well as through a non-genomic action mediated by cell-surface estrogen-binding receptors like GPR30 (39) (Fig 1).



**Figure 1.** Overview of genomic and non-genomic effects of estrogen (E2). E2 can modulate the gene expression and activity of signaling molecules by binding to estrogen receptors (ER) through genomic and/or non-genomic pathways.

Estrogen protects endothelial cells against the prevention of atherosclerosis; consequently, it has been used to prevent cardiovascular diseases in postmenopausal women. Estrogen reduces the expression of endothelin-1 (a pro-myogenic and vasoconstriction factor), reduces the expression of adhesion molecules in response to cytokines, and stimulates the synthesis of nitric oxide (NO) by endothelial cells through phosphorylation and activation of endothelial nitric oxide synthase (eNOS) (acute or immediate effect), as well as by regulating the increase in eNOS gene expression (chronic effect) (43,44). Considering the effects of NO on the tone of vascular smooth muscle, its increase may have additional relevance for the maintenance of homeostasis of the cardiovascular system (45). Moreover, there is evidence that the influence of estradiol in



reducing the expression of transcription factors, such as nuclear factor kappa B (NF- $\kappa$ B) and activating protein-1 (AP-1), in the presence of experimental hemorrhagic shock (46).

Acute reductions in the concentration of female sex hormones may cause changes in several systems, including the immune system. Previous studies have shown that the removal of estradiol by ovariectomy has an important impact on gene expression in myocardial tissue, as it reduces the expression of genes that mediate vasodilation and increases the expression of genes relevant to the induction of inflammation, apoptosis, and proteolysis (47).

## SCOPE OF THE THESIS

In this context, the aim of this thesis is to describe the effects of female sex hormone modulation on systemic inflammation and organ injury in brain-dead female donors. In **Chapter 2** we analyze the differences in microcirculatory changes caused by BD and inflammatory mediators between sexes.

Considering female sex hormones as immunomodulators and the association between severe organ inflammation in female animals following BD with acute estradiol reduction, we investigated the potential therapeutic effects of 17 $\beta$ -estradiol treatment to mitigate the deleterious process in the heart (**Chapter 3**), lung (**Chapter 4**), and kidney (**Chapter 5**) from brain-dead female rats.

In parallel, preservation solutions and perfusion techniques are important for the maintenance and improvement of the donor organ, which is directly related to morbidity and survival after transplantation. In **Chapter 6**, we compare renal inflammation in male and female patients with BD followed by isolated kidney perfusion.

# REFERENCES

1. Pratschke J, Neuhaus P, Tullius SG. What can be learned from brain-death models? Vol. 18, *Transplant International*. 2005. p. 15–21.
2. Skrabal CA, Thompson LO, Potapov E V, Southard RE, Joyce DL, Youker KA, et al. Organ-specific regulation of pro-inflammatory molecules in heart, lung, and kidney following brain death. *J Surg Res* [Internet]. 2005 Jan [cited 2018 Nov 2];123(1):118–25. Available from: <http://linkinghub.elsevier.com/retrieve/pii/S0022480404005219>
3. McKay DB. The role of innate immunity in donor organ procurement. Vol. 33, *Seminars in Immunopathology*. 2011. p. 169–84.
4. Takada M, Nadeau KC, Hancock WW, Mackenzie HS, Shaw GD, Waaga AM, et al. Effects of explosive brain death on cytokine activation of peripheral organs in the rat. *Transplantation*. 1998 Jun 27;65(12):1533–42.
5. Feng S, Goodrich NP, Bragg-Gresham JL, Dykstra DM, Punch JD, DebRoy MA, et al. Characteristics associated with liver graft failure: The concept of a donor risk index. *Am J Transplant* [Internet]. 2006 Apr [cited 2020 Jun 21];6(4):783–90. Available from: <https://pubmed.ncbi.nlm.nih.gov/16539636/>
6. Terasaki PI, Cecka JM, Gjertson DW, Takemoto S. High Survival Rates of Kidney Transplants from Spousal and Living Unrelated Donors. *N Engl J Med* [Internet]. 1995 Aug 10 [cited 2020 Jun 21];333(6):333–6. Available from: <http://www.nejm.org/doi/abs/10.1056/NEJM199508103330601>
7. Wilhelm MJ, Pratschke J, Laskowski IA, Paz DM, Tilney NL. Brain Death and Its Impact on the Donor Heart—Lessons From Animal Models [Internet]. 2000 [cited 2019 May 24]. Available from: [https://www.jhltonline.org/article/S1053-2498\(00\)00073-5/pdf](https://www.jhltonline.org/article/S1053-2498(00)00073-5/pdf)
8. Pratschke J, Wilhelm MJ, Kusaka M, Laskowski I, Tilney NL. A model of gradual onset brain death for transplant-associated studies in rats. *Transplantation* [Internet]. 2000 Feb 15 [cited 2020 Jun 21];69(3):427–30. Available from: <https://pubmed.ncbi.nlm.nih.gov/10706054/>
9. Seguín C, Devaux Y, Grosjean S, Siaghy EM, Mairose P, De Talancé N, et al. Evidence of functional myocardial ischemia associated with myocardial dysfunction in brain-dead pigs. *Circulation* [Internet]. 2001 Sep 18 [cited 2021 Mar 8];104(SUPPL. 1). Available from: <https://pubmed.ncbi.nlm.nih.gov/11568055/>
10. Bugge JF. Brain death and its implications for management of the potential organ donor [Internet]. Vol. 53, *Acta Anaesthesiologica Scandinavica*. *Acta Anaesthesiol Scand*; 2009 [cited 2021 Mar 8]. p. 1239–50. Available from: <https://pubmed.ncbi.nlm.nih.gov/19681785/>
11. Theodore J, Robin ED. PATHOGENESIS OF NEUROGENIC PULMONARY OEDEMA. *Lancet* [Internet]. 1975 Oct 18 [cited 2021 Mar 8];306(7938):749–51. Available from: <https://pubmed.ncbi.nlm.nih.gov/52777/>
12. Novitzky D. Detrimental effects of brain death on the potential organ donor. *Transplant Proc* [Internet]. 1997 Dec [cited 2020 Mar 27];29(8):3770–2. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/9414920>
13. McKeown DW, Bonser RS, Kellum JA. Management of the heartbeating brain-dead organ donor. *Br J Anaesth* [Internet]. 2012;108(SUPPL. 1):i96–107. Available from: <http://dx.doi.org/10.1093/bja/aer351>
14. Van Der Hoeven JAB, Ploeg RJ, Postema F, Molema I, De Vos P, Girbes ARJ, et al. Induction of organ dysfunction and up-regulation of inflammatory markers in the liver and kidneys of hypotensive brain dead rats: A model to study marginal organ donors. In: *Transplantation* [Internet]. Lippincott Williams and Wilkins; 1999 [cited 2020 Jul 20]. p. 1884–90. Available from: <https://pubmed.ncbi.nlm.nih.gov/10628769/>
15. Roelsgaard K, Bøtker HE, Stødkilde-Jørgensen H, Andreassen F, Jensen SL, Keiding S. Effects of brain death and glucose infusion on hepatic glycogen and blood hormones in the pig. *Hepatology* [Internet]. 1996 Oct 1 [cited 2021 Mar 8];24(4):871–5. Available from: <http://doi.wiley.com/10.1002/hep.510240419>
16. Chen EP, Bittner HB, Kendall SWH, Van Trigt P. Hormonal and hemodynamic changes in a validated animal model of brain death. *Crit Care Med* [Internet]. 1996 [cited 2020 Jul 17];24(8):1352–9. Available from: <https://pubmed.ncbi.nlm.nih.gov/8706491/>
17. Domínguez-Roldán JM, García-Alfaro C, Jimenéz-González PI, Hernández-Hazañas F, Gascón Castillo ML, Egea Guerrero JJ. Muerte encefálica: repercusión sobre órganos y tejidos. *Med Intensiva* [Internet]. 2009 Dec 24 [cited 2020 Jul 14];33(9):434–41. Available from: <https://linkinghub.elsevier.com/retrieve/pii/S0210569109000357>
18. Chamorro C, Falcón JA, Michelena JC. Controversial Points in Organ Donor Management.

- Transplant Proc [Internet]. 2009 Oct [cited 2020 Mar 27];41(8):3473–5. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/19857774>
19. Yeh T. J, Wechsler AS, Graham LJ, Loesser KE, Sica DA, Wolfe L, et al. Acute brain death alters left ventricular myocardial gene expression. *J Thorac Cardiovasc Surg*. 1999;117(2):365–74.
  20. Silva IA, Correia CJ, Simas R, Correia C d. J, Cruz JWMC, Ferreira SG, et al. Inhibition of Autonomic Storm by Epidural Anesthesia Does Not Influence Cardiac Inflammatory Response After Brain Death in Rats. *Transplant Proc* [Internet]. 2012 Sep 1 [cited 2019 May 24];44(7):2213–8. Available from: <https://www.sciencedirect.com/science/article/pii/S0041134512008068?via%3Dihub>
  21. Halejcio-Delophont P, Siaghy EM, Devaux Y, Richoux JP, Bischoff N, Carteaux JP, et al. Consequences of brain death on coronary blood flow and myocardial metabolism. In: *Transplantation Proceedings*. Elsevier; 1998. p. 2840–1.
  22. Khush KK, Kubo JT, Desai M. INFLUENCE OF DONOR AND RECIPIENT SEX MISMATCH ON HEART TRANSPLANT OUTCOMES: ANALYSIS OF THE ISHLT REGISTRY. 2012 [cited 2018 Nov 12]; Available from: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3322315/pdf/nihms355637.pdf>
  23. Zeier M, Döhler B, Opelz G, Ritz E. The effect of donor gender on graft survival. *J Am Soc Nephrol* [Internet]. 2002 Oct 1 [cited 2021 Mar 8];13(10):2570–6. Available from: <https://pubmed.ncbi.nlm.nih.gov/12239247/>
  24. Neugarten J, Srinivas T, Tellis V, Silbiger S, Greenstein S. The effect of donor gender on renal allograft survival. *J Am Soc Nephrol*. 1996;7(2).
  25. Sato M, Gutierrez C, Kaneda H, Liu M, Waddell TK, Keshavjee S. The Effect of Gender Combinations on Outcome in Human Lung Transplantation: The International Society of Heart and Lung Transplantation Registry Experience. 2006;
  26. Hibi T, Sageshima J, Molina E, Ciancio G, Nishida S, Chen L, et al. Predisposing factors of diminished survival in simultaneous liver/kidney transplantation. *Am J Transplant* [Internet]. 2012 Nov [cited 2018 Nov 2];12(11):2966–73. Available from: <http://doi.wiley.com/10.1111/j.1600-6143.2012.04121.x>
  27. Kaczmarek I, Meiser B, Beiras-Fernandez A, Guethoff S, Überfuhr P, Angele M, et al. Gender Does Matter: Gender-Specific Outcome Analysis of 67,855 Heart Transplants. *Thorac Cardiovasc Surg* [Internet]. 2012 Dec 20 [cited 2019 May 24];61(01):029–36. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/23258761>
  28. Weiss ES, Allen JG, Patel ND, Russell SD, Baumgartner WA, Shah AS, et al. The impact of donor-recipient sex matching on survival after orthotopic heart transplantation: Analysis of 18 000 transplants in the modern era. *Circ Hear Fail*. 2009;2(5):401–8.
  29. Bonnano Abib AL de O, Correia C de J, Armstrong-Jr R, Ricardo-da-Silva FY, Ferreira SG, Vidal-dos-Santos M, et al. The influence of female sex hormones on lung inflammation after brain death - an experimental study. *Transpl Int* [Internet]. 2020 Mar 1 [cited 2020 Jul 20];33(3):279–87. Available from: <https://pubmed.ncbi.nlm.nih.gov/31701582/>
  30. Breithaupt-Faloppa AC, Ferreira SG, Kudo GK, Armstrong R, Tavares-de-Lima W, da Silva LFF, et al. Sex-related differences in lung inflammation after brain death. *J Surg Res* [Internet]. 2016 Feb [cited 2019 May 24];200(2):714–21. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/26547667>
  31. Simão RR, Ferreira SG, Kudo GK, Armstrong Junior R, da Silva LFF, Sannomiya P, et al. Sex differences on solid organ histological characteristics after brain death. *Acta Cir Bras*. 2016;31(4).
  32. Deitch EA, Chu H, Xu DZ. Organ blood flow and the central hemodynamic response are better preserved in female, as opposed to the male rats, after trauma-hemorrhagic shock. *J Trauma - Inj Infect Crit Care* [Internet]. 2008 Sep [cited 2020 Nov 24];65(3):566–72. Available from: <https://pubmed.ncbi.nlm.nih.gov/18784569/>
  33. Deitch EA, Feketeova E, Lu Q, Zaets S, Berezina TL, MacHiedo GW, et al. Resistance of the female, as opposed to the male, intestine to I/R-mediated injury is associated with increased resistance to gut-induced distant organ injury. *Shock* [Internet]. 2008 Jan [cited 2021 Mar 8];29(1):78–83. Available from: <https://pubmed.ncbi.nlm.nih.gov/17666951/>
  34. Deitch EA, Feketeova E, Lu Q, Zaets S, Berezina TL, Machiedo GW, et al. RESISTANCE OF THE FEMALE, AS OPPOSED TO THE MALE, INTESTINE TO I/R-MEDIATED INJURY IS ASSOCIATED WITH INCREASED RESISTANCE TO GUT-INDUCED DISTANT ORGAN INJURY. *Shock* [Internet]. 2007 Jul [cited 2020 Nov 24];PAP(1):78–83. Available from: <http://journals.lww.com/00024382-900000000-99892>
  35. Grossman CJ. Interactions between the gonadal steroids and the immune system. *Science* (80- ) [Internet]. 1985 [cited 2021 Mar 8];227(4684):257–61. Available from: <https://pubmed.ncbi.nlm.nih.gov/3871252/>
  36. OLSEN NJ, KOVACS WJ. Gonadal Steroids and Immunity\*. *Endocr Rev* [Internet]. 1996 Aug [cited 2020 Mar 27];17(4):369–84. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/8854050>
  37. Schröder J, Kahlke V, Book M, Stüber F. Gender differences in sepsis: Genetically determined? *Shock*. 2000;14(3):307–11.

38. Stangl V, Baumann G, Stangl K. Coronary atherogenic risk factors in women [Internet]. Vol. 23, European Heart Journal. 2002 [cited 2020 Mar 27]. p. 1738–52. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/12419293>
39. Iorga A, Cunningham CM, Moazeni S, Ruffenach G, Umar S, Eghbali M. The protective role of estrogen and estrogen receptors in cardiovascular disease and the controversial use of estrogen therapy. Vol. 8, Biology of sex differences. 2017. p. 33.
40. Knowlton AA, Korzick DH. Estrogen and the female heart. Mol Cell Endocrinol [Internet]. 2014 May 25 [cited 2019 May 24];389(1–2):31–9. Available from: <https://www.sciencedirect.com/science/article/pii/S0303720714000045?via%3Dihub>
41. Gee AC, Sawai RS, Differding J, Muller P, Underwood S, Schreiber MA. The influence of sex hormones on coagulation and inflammation in the trauma patient. Shock [Internet]. 2008 Mar [cited 2021 Mar 8];29(3):334–41. Available from: <https://pubmed.ncbi.nlm.nih.gov/18437714/>
42. Kawasaki T, Chaudry IH. The effects of estrogen on various organs: Therapeutic approach for sepsis, trauma, and reperfusion injury. Part 1: Central nervous system, lung, and heart. J Anesth [Internet]. 2012 Dec [cited 2020 Mar 27];26(6):883–91. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/22729227>
43. Sumi D, Ignarro LJ. Estrogen-related receptor  $\alpha 1$  up-regulates endothelial nitric oxide synthase expression. Proc Natl Acad Sci U S A. 2003 Nov 25;100(SUPPL. 2):14451–6.
44. Florian M, Lu Y, Angle M, Magder S. Estrogen induced changes in Akt-dependent activation of endothelial nitric oxide synthase and vasodilation. Steroids. 2004 Sep 1;69(10):637–45.
45. Hogg ME, Vavra AK, Banerjee MN, Martinez J, Jiang Q, Keefer LK, et al. The Role of Estrogen Receptor  $\alpha$  and  $\beta$  in Regulating Vascular Smooth Muscle Cell Proliferation is Based on Sex. In: Journal of Surgical Research [Internet]. J Surg Res; 2012 [cited 2021 Mar 8]. Available from: <https://pubmed.ncbi.nlm.nih.gov/22099601/>
46. Shimizu T, Yu HP, Suzuki T, Szalay L, Hsieh YC, Choudhry MA, et al. The role of estrogen receptor subtypes in ameliorating hepatic injury following trauma-hemorrhage. J Hepatol [Internet]. 2007 Jun [cited 2021 Mar 8];46(6):1047–54. Available from: <https://pubmed.ncbi.nlm.nih.gov/17336418/>
47. Hamilton KL, Lin L, Wang Y, Knowlton AA. Effect of ovariectomy on cardiac gene expression: Inflammation and changes in SOCS gene expression. Physiol Genomics [Internet]. 2008 Jan 17 [cited 2021 Mar 8];32(2):254–63. Available from: <https://pubmed.ncbi.nlm.nih.gov/17986523/>