

University of Groningen

## Role of interaction between B cells and epithelial cells in pSS

Pringle, Sarah; Verstappen, Gwenny M; Bootsma, Hendrika; Kroese, Frans G M

*Published in:*  
Annals of the Rheumatic Diseases

*DOI:*  
[10.1136/annrheumdis-2020-219079](https://doi.org/10.1136/annrheumdis-2020-219079)

**IMPORTANT NOTE: You are advised to consult the publisher's version (publisher's PDF) if you wish to cite from it. Please check the document version below.**

*Document Version*  
Publisher's PDF, also known as Version of record

*Publication date:*  
2020

[Link to publication in University of Groningen/UMCG research database](#)

*Citation for published version (APA):*

Pringle, S., Verstappen, G. M., Bootsma, H., & Kroese, F. G. M. (2020). Role of interaction between B cells and epithelial cells in pSS: Correspondence. *Annals of the Rheumatic Diseases*.  
<https://doi.org/10.1136/annrheumdis-2020-219079>

**Copyright**

Other than for strictly personal use, it is not permitted to download or to forward/distribute the text or part of it without the consent of the author(s) and/or copyright holder(s), unless the work is under an open content license (like Creative Commons).

**Take-down policy**

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

*Downloaded from the University of Groningen/UMCG research database (Pure): <http://www.rug.nl/research/portal>. For technical reasons the number of authors shown on this cover page is limited to 10 maximum.*

## Role of interaction between B cells and epithelial cells in pSS

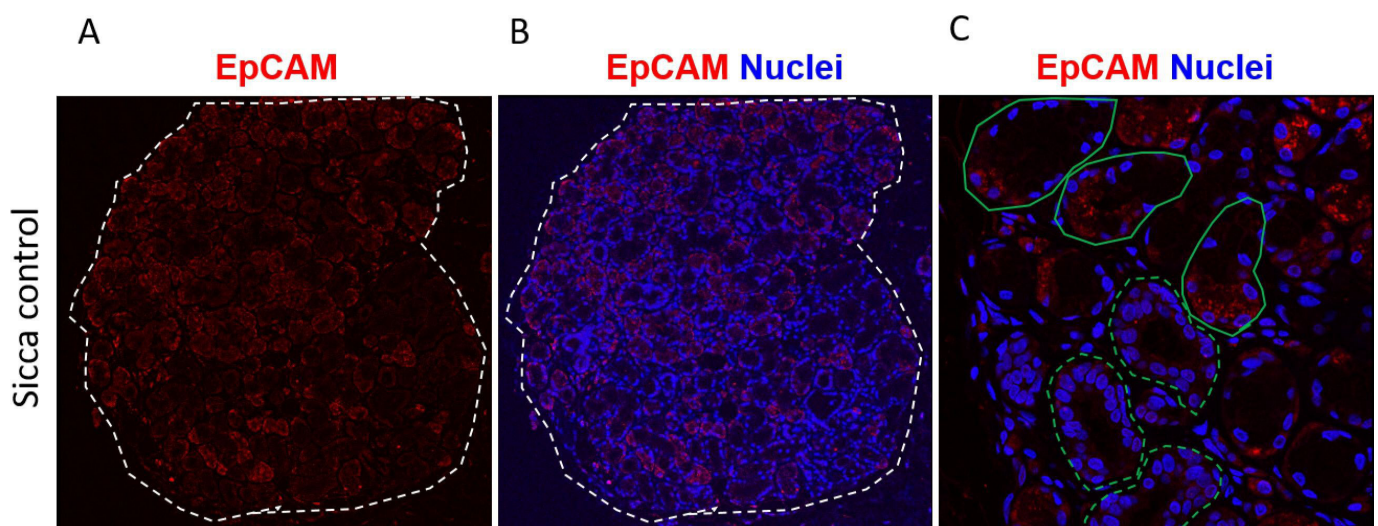
We read with great interest the recent publication by Rivière *et al*, concerning the activation of B cells by salivary gland epithelial cells (SGECs) from patients with primary Sjögren's syndrome (pSS).<sup>1</sup> The authors elegantly show that coculture of CD19<sup>+</sup> peripheral blood B cells with SGECs isolated from labial salivary glands (LSGs) of patients with pSS increases expression of the B cell activation marker CD38, and the memory B cell marker CD27. This effect was the most noticeable with further stimulation of toll-like receptor 3 by means of Poly(I:C) medium supplementation. Treatment of cocultures with agents ablating specific B cell signalling pathways (ibrutinib, blocking BTK signalling), and non-B cell specific pathways (LY294002 blocking the PI3K pathway and leflunomide, inhibiting lymphocyte proliferation) reduced this effect somewhat.

We would like to take the opportunity to comment on these interesting findings. First, their main conclusion is that SGECs (by presumption ductal cells) from patients with pSS support the activation and survival of B cells. We would like to add that we have previously demonstrated *in situ* the presence of an epithelium-associated subset of B cells, expressing FcRL4, in both minor (labial) and major (parotid) SGs of patients with pSS.<sup>2</sup> This FcRL4<sup>+</sup> B cell subset is highly proliferative, and located both periductally and intraductally.<sup>2</sup> Follow-up studies by Verstappen *et al* using RNAseq revealed that parotid FcRL4<sup>+</sup> B cells in SGs express genes indicative of both chronic (T-bet, CD11c) and general (TACI, CXCR3, NF- $\kappa$ B signalling genes, IL-6) activation.<sup>3</sup> We further provided some evidence that these chronically activated (intra-)epithelial B cells are involved in epithelial cell proliferation, resulting in the formation of lymphoepithelial lesions (LELs).<sup>2-4</sup> The findings of Rivière *et al* support our notion of an intimate relationship between epithelium and B cells in SGs of patients with pSS, which might be very relevant for the pathogenesis of the disease. We would be extremely curious to see if in the studies of Rivière *et al*, FcRL4

is also differentially expressed in B cells sorted from LSGs of patients with pSS.

Second, Rivière *et al* employed cocultures of bulk (blood-derived) CD19<sup>+</sup> B cells with SGECs. We suggest that the unrefined use of the CD19<sup>+</sup> pool makes the interpretation of which B cell population, for example CD27<sup>-</sup> and CD27<sup>+</sup> naive and memory B cells, respectively, or FcRL4<sup>+</sup> B cells, proliferates (or dies) following SGEC coculture challenging to gauge. Additionally, the accurate recapitulation of pSS-associated B cell/SGEC dynamics in this system can be questioned, considering the lack of crucial B cell stimuli to fully mimic complete B cell activation (TLR7 and/or TLR9 agonism, presence of IL-21 and BCR stimulation). In the same trend, the application of potential therapeutic agents (eg, LY294002 blocking the PI3K pathway) and stimuli (IFN- $\alpha$ , IFN- $\gamma$  and Poly(I:C)) to B cell-SGEC cocultures and read out via B cell viability render the data challenging to interpret in terms of effects on the epithelium. Considering the likely crosstalk between the two cellular entities, we wonder whether the authors plan to probe this in future experiments.

Finally, regarding the study of SGECs alone, the authors employed CD326 (EpCAM) to select epithelial cells from digested LSGs. Transcriptomics analysis of these CD326<sup>+</sup> cells was performed. We were surprised to see expression of immune genes such as *IGHG1*, *BTK* (B cell/monocyte restricted), *CD8A* (NK/T cell restricted) in the CD326<sup>+</sup> SGEC fraction, and were wondering how the authors interpret these findings (Table S3). The lack of pathways reflecting changes in epithelial cell dynamics (cell cycle, proliferation, apoptosis) may also suggest that this SGEC system do not truly mirror the *in vivo* situation suggested by other authors.<sup>5-9</sup> The inference from the paper is that CD326<sup>+</sup> cells equate to ductal cells in the LSG, or least no indication to the contrary is stated. Acinar cells, epithelial cells present in LSGs, also express CD326 (figure 1). Considering their likely involvement in pSS pathogenesis, acinar cells may be partly responsible for the immune profile observed.<sup>10-13</sup> Using SGECs might well be an oversimplification of the SG, and



**Figure 1** Both ductal and acinar epithelial cells express CD326 (EpCAM) in the LSGs. (A) EpCAM immunofluorescence staining of a sicca control LSG. (B) Merged EpCAM immunostaining with nuclear counterstain. (C) High-resolution microscopy of ductal cells (dashed green line) and acinar cells (solid green line), showing EpCAM expression by both. Methodology: paraffin sections of LSGs were dewaxed and rehydrated. Antigen retrieval was performed with a sodium citrate (pH 6.0) buffer containing 0.5% Tween. Sections were blocked in 1% bovine serum albumin (BSA), and incubated overnight at 4°C in mouse anti-human EpCAM conjugated to the fluorophore e660, at 1:100 dilution (eBioscience clone 1B7). Nuclei were counterstained with Hoechst and confocal microscopy performed with the Leica TCS Sp8 confocal microscope. LSG, labial salivary gland.

whether cultured SGEs are representative of in situ epithelial cells remains to be shown.

Although providing a platform for drug screening from which to further expand on and certainly representing an important contribution to the field, the applicability of two-dimensional epithelial cultures to in situ SG architecture remains to be validated. Three-dimensional approaches using SG organoids may represent a step towards true reflection of reality, and have been shown to provide valuable information about epithelial cell dynamics in pSS.<sup>6</sup> While acknowledging the novel and extensive work presented by Rivière *et al*, we look forward to seeing how the more intricate interactions between B cells and the epithelium can be teased apart, including those of LEL development.

**Sarah Pringle** , **Gwenny M Verstappen, Hendrika Bootsma, Frans G M Kroese**

Rheumatology and Clinical Immunology, University Medical Center Groningen, University of Groningen, Groningen, Netherlands

**Correspondence to** Dr Sarah Pringle, Rheumatology & Clinical Immunology, University Medical Center Groningen, 9713 GZ Groningen, Netherlands; s.a.pringle@umcg.nl

**Twitter** Sarah Pringle @SjogrensUMCG; @springles82

**Contributors** SP, GMV and FGMK generated data, and formulated the manuscript first draft. HB read the manuscript critically.

**Funding** ReumaNederland (LLP-29, T015-052).

**Competing interests** None declared.

**Patient and public involvement** Patients and/or the public were not involved in the design, or conduct, or reporting or dissemination plans of this research.

**Patient consent for publication** Not required.

**Provenance and peer review** Not commissioned; internally peer reviewed.

© Author(s) (or their employer(s)) 2020. No commercial re-use. See rights and permissions. Published by BMJ.



**To cite** Pringle S, Verstappen GM, Bootsma H, *et al*. *Ann Rheum Dis* Epub ahead of print: [please include Day Month Year]. doi:10.1136/annrheumdis-2020-219079

Received 8 September 2020

Accepted 9 September 2020



► <http://dx.doi.org/10.1136/annrheumdis-2020-219096>

*Ann Rheum Dis* 2020;0:1–2. doi:10.1136/annrheumdis-2020-219079

#### ORCID iD

Sarah Pringle <http://orcid.org/0000-0002-0779-1680>

#### REFERENCES

- Rivière E, Pascaud J, Tchitchek N, *et al*. Salivary gland epithelial cells from patients with Sjögren's syndrome induce B-lymphocyte survival and activation. *Ann Rheum Dis* 2020;annrheumdis-2019-216588.
- Haacke EA, Bootsma H, Spijkervet FKL, *et al*. FCRL4<sup>+</sup> B-cells in salivary glands of primary Sjögren's syndrome patients. *J Autoimmun* 2017;81:90–8.
- Verstappen GM, Ice JA, Bootsma H, *et al*. Gene expression profiling of epithelium-associated FCRL4<sup>+</sup> B cells in primary Sjögren's syndrome reveals a pathogenic signature. *J Autoimmun* 2020;109:102439.
- van Ginkel MS, Haacke EA, Bootsma H, *et al*. Presence of intraepithelial B-lymphocytes is associated with the formation of lymphoepithelial lesions in salivary glands of primary Sjögren's syndrome patients. *Clin Exp Rheumatol* 2019;118:42–8.
- Wang X, Shaalan A, Liefers S, *et al*. Dysregulation of NF-κB in glandular epithelial cells results in Sjögren's-like features. *PLoS One* 2018;13:e0200212.
- Pringle S, Wang X, Verstappen GMPJ, *et al*. Salivary gland stem cells age prematurely in primary Sjögren's syndrome. *Arthritis Rheumatol* 2019;71:133–42.
- Vakrakou AG, Boiu S, Ziakas PD, *et al*. Systemic activation of NLRP3 inflammasome in patients with severe primary Sjögren's syndrome fueled by inflammagenic DNA accumulations. *J Autoimmun* 2018;91:23–33.
- Manoussakis MN, Spachidou MP, Maratheftis CI. Salivary epithelial cells from Sjögren's syndrome patients are highly sensitive to anoikis induced by TLR-3 ligation. *J Autoimmun* 2010;35:212–8.
- Manoussakis MN, Kapsogeorgou EK. The role of intrinsic epithelial activation in the pathogenesis of Sjögren's syndrome. *J Autoimmun* 2010;35:219–24.
- Sung HH, Castro I, González S, *et al*. Muc1/Sec and MUC1/Y overexpression is associated with inflammation in Sjögren's syndrome. *Oral Dis* 2015;21:730–8.
- Ewert P, Aguilera S, Alliende C, *et al*. Disruption of tight junction structure in salivary glands from Sjögren's syndrome patients is linked to proinflammatory cytokine exposure. *Arthritis Rheum* 2010;62:1280–9.
- Castro I, Albornoz N, Aguilera S, *et al*. Aberrant MUC1 accumulation in salivary glands of Sjögren's syndrome patients is reversed by TUDCA in vitro. *Rheumatology* 2020;59:742–53.
- Barrera M-J, Aguilera S, Veerman E, *et al*. Salivary mucins induce a Toll-like receptor 4-mediated pro-inflammatory response in human submandibular salivary cells: are mucins involved in Sjögren's syndrome? *Rheumatology* 2015;54:1518–27.