

University of Groningen

## F-18-FDG PET/CT in the Diagnostic and Treatment Evaluation of Pediatric Posttransplant Lymphoproliferative Disorders

Montes de Jesus, Filipe; Glaudemans, Andor W J M; Tissing, Wim; Dierckx, Rudi A; Rosati, Stefano; Diepstra, Arjan; Noordzij, Walter; Kwee, Thomas C

*Published in:*  
Journal of Nuclear Medicine

*DOI:*  
[10.2967/jnumed.119.239624](https://doi.org/10.2967/jnumed.119.239624)

**IMPORTANT NOTE: You are advised to consult the publisher's version (publisher's PDF) if you wish to cite from it. Please check the document version below.**

*Document Version*  
Publisher's PDF, also known as Version of record

*Publication date:*  
2020

[Link to publication in University of Groningen/UMCG research database](#)

*Citation for published version (APA):*

Montes de Jesus, F., Glaudemans, A. W. J. M., Tissing, W., Dierckx, R. A., Rosati, S., Diepstra, A., Noordzij, W., & Kwee, T. C. (2020). F-18-FDG PET/CT in the Diagnostic and Treatment Evaluation of Pediatric Posttransplant Lymphoproliferative Disorders. *Journal of Nuclear Medicine*, 61(9), 1307-1313. <https://doi.org/10.2967/jnumed.119.239624>

### Copyright

Other than for strictly personal use, it is not permitted to download or to forward/distribute the text or part of it without the consent of the author(s) and/or copyright holder(s), unless the work is under an open content license (like Creative Commons).

The publication may also be distributed here under the terms of Article 25fa of the Dutch Copyright Act, indicated by the "Taverne" license. More information can be found on the University of Groningen website: <https://www.rug.nl/library/open-access/self-archiving-pure/taverne-amendment>.

### Take-down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

Downloaded from the University of Groningen/UMCG research database (Pure): <http://www.rug.nl/research/portal>. For technical reasons the number of authors shown on this cover page is limited to 10 maximum.

---

---

# <sup>18</sup>F-FDG PET/CT in the Diagnostic and Treatment Evaluation of Pediatric Posttransplant Lymphoproliferative Disorders

Filipe M. Montes de Jesus<sup>1</sup>, Andor W.J.M. Glaudemans<sup>1</sup>, Wim J. Tissing<sup>2</sup>, Rudi A.J.O. Dierckx<sup>1</sup>, Stefano Rosati<sup>3</sup>, Arjan Diepstra<sup>3</sup>, Walter Noordzij<sup>1</sup>, and Thomas C. Kwee<sup>4</sup>

<sup>1</sup>Department of Nuclear Medicine and Molecular Imaging, University of Groningen, University Medical Center Groningen, Groningen, The Netherlands; <sup>2</sup>Department of Pediatric Oncology/Hematology, University of Groningen, University Medical Center Groningen, Groningen, The Netherlands; <sup>3</sup>Department of Pathology and Medical Biology, University of Groningen, University Medical Center Groningen, Groningen, The Netherlands; and <sup>4</sup>Department of Radiology, University of Groningen, University Medical Center Groningen, Groningen, The Netherlands

---

We aimed to evaluate the diagnostic performance of <sup>18</sup>F-FDG PET/CT for the detection of posttransplantation lymphoproliferative disorder (PTLD) in a pediatric population and explore its feasibility during response assessment. **Methods:** This retrospective study included 28 pediatric transplant recipients who underwent a total of 32 <sup>18</sup>F-FDG PET/CT scans due to clinical suspicion of PTLD within an 8-y period. Pathology reports and 2 y of follow-up were used as the reference standard. Twenty-one response assessment <sup>18</sup>F-FDG PET/CT scans were reevaluated according to the Lugano criteria. **Results:** The diagnosis of PTLD was established in 14 patients (49%). Sensitivity, specificity, positive predictive value, and negative predictive value of <sup>18</sup>F-FDG PET/CT for the detection of PTLD in children with a clinical suspicion of this disease were 50% (7/14), 100% (18/18), 100% (7/7), and 72% (18/25), respectively. False-negative results occurred in patients with PTLD in the Waldeyer's ring, cervical lymph nodes, or small bowel with either nondestructive or polymorphic PTLD. Two of 5 interim <sup>18</sup>F-FDG PET/CT scans and 3 of 9 end-of-treatment <sup>18</sup>F-FDG PET/CT scans were false-positive. **Conclusion:** <sup>18</sup>F-FDG PET/CT had good specificity and positive predictive value but low to moderate sensitivity and negative predictive value for the detection of PTLD in a 28-pediatric-patient cohort with a clinical suspicion of this disease. False-negative results were confirmed in the Waldeyer's ring, cervical lymph nodes, and small bowel with either nondestructive or polymorphic PTLD subtypes. <sup>18</sup>F-FDG PET/CT appears to have a limited role in the response assessment setting of pediatric PTLD, given the observed high proportions of false-positives both at interim and at end-of-treatment evaluations.

**Key Words:** posttransplant lymphoproliferative disorder; <sup>18</sup>F-fluorodeoxyglucose PET; <sup>18</sup>F-FDG PET/CT; diagnosis; pediatric

**J Nucl Med 2020; 61:1307–1313**

DOI: 10.2967/jnumed.119.239624

---

**P**osttransplantation lymphoproliferative disorder (PTLD) is a major complication of continued immunosuppressive therapy after

solid-organ or hematopoietic stem cell transplantation. Morphologically, PTLD ranges from Epstein-Barr virus-driven polyclonal lesions to aggressive monoclonal lymphoid proliferations, classified by the World Health Organization as nondestructive, polymorphic, monomorphic, or classic Hodgkin lymphoma PTLD (1).

Compared with adults, pediatric PTLD patients have distinct characteristics regarding incidence and presentation. PTLD is the most common posttransplant malignancy in children, with a higher reported incidence than in adults (2–4). An important risk factor associated with PTLD development is an Epstein-Barr virus status mismatch between seropositive donors and seronegative recipients. The Epstein-Barr virus has a recognized role in the pathogenesis and development of PTLD, particularly related to nondestructive and polymorphic lesions. Because only 20%–25% of the pediatric population is an Epstein-Barr virus carrier by the age of 5 y (unlike 80%–90% in the adult population), children are at an increased risk of developing this disorder after transplantation (5,6). The presentation may be asymptomatic or have a variation of symptoms, including B symptoms, lymphadenopathy, or graft dysfunction. Although it may be localized in any organ system, pediatric PTLD has been reported to occur more frequently in the Waldeyer's ring and gastrointestinal tract (7,8). This location is in contrast to that in the adult PTLD population, for whom lesions have been reported to occur proportionally more often in the transplant allograft and lymph nodes (6,9).

Timely diagnosis of PTLD remains challenging but is crucial for treatment initiation, management, and prognostication. Because reduction of immunosuppression is the first-line intervention in many PTLD cases, prompt therapy, particularly in nondestructive lesions, may be adequate to achieve remission. Nevertheless, this therapy may also jeopardize the transplanted organ (10–12). Biopsy remains necessary for diagnostic confirmation, but imaging may be used to confirm or refute clinical suspicion of PTLD and identify suggestive lesions accessible for biopsy. For treatment evaluation, imaging-based response assessment may be used to monitor lesions in the entire body, circumventing the need for invasive biopsies and their associated complications. <sup>18</sup>F-FDG PET/CT combines metabolic and anatomic information and may be of value in the diagnosis and treatment evaluation of pediatric PTLD. Preliminary literature suggests that <sup>18</sup>F-FDG PET/CT may be helpful in detecting occult lesions and clarifying findings on other imaging modalities (13–19). However, as these previous studies suffered from small sample sizes and frequently mixed pediatric and adult populations, the value of <sup>18</sup>F-FDG PET/CT in pediatric PTLD remains unclear. If <sup>18</sup>F-FDG PET/CT proves

---

Received Nov. 14, 2019; revision accepted Jan. 3, 2020.

For correspondence or reprints contact: Filipe M. Montes de Jesus, Department of Nuclear Medicine and Molecular Imaging, University Medical Center Groningen, Hanzeplein 1 Groningen 9700 RB, The Netherlands.

E-mail: f.m.montes.de.jesus@umcg.nl

Published online Jan. 31, 2020.

COPYRIGHT © 2020 by the Society of Nuclear Medicine and Molecular Imaging.

accurate in detecting PTLT and feasible for treatment evaluation, it may be implemented in future guidelines. In this study, we aimed to determine the diagnostic performance of  $^{18}\text{F}$ -FDG PET/CT for the detection of PTLT in the pediatric population and to explore its feasibility in the therapy response assessment setting.

## MATERIALS AND METHODS

### Study Design and Patients

This retrospective single-center study was conducted at the University Medical Center Groningen. All consecutive patients 18 y old or younger for whom a  $^{18}\text{F}$ -FDG PET/CT scan was requested on clinical suspicion of PTLT between January 2010 until January 2019 were included. The first  $^{18}\text{F}$ -FDG PET/CT scan and in some children a second or third  $^{18}\text{F}$ -FDG PET/CT scan (provided the  $^{18}\text{F}$ -FDG PET/CT scan was requested because of a clinical suspicion of PTLT and there was a minimum interval of 2 y without any evidence of PTLT between these scans) were included for the diagnostic performance analysis. In patients with pathologically proven PTLT, all  $^{18}\text{F}$ -FDG PET/CT scans for treatment evaluation were analyzed to explore the feasibility of  $^{18}\text{F}$ -FDG PET/CT in the response assessment setting. Demographic, relevant clinical data and PTLT morphology or histology were retrieved from the electronic patient charts. Patients who had a complete tumor resection before  $^{18}\text{F}$ -FDG PET/CT, and patients for whom the established reference standard criteria were not fulfilled, were excluded. A waiver was obtained from the local medical ethics committee on September 7, 2017 (study 201700855).

### $^{18}\text{F}$ -FDG PET/CT Acquisition

All  $^{18}\text{F}$ -FDG PET/CT scans were performed on a Biograph 40- or 64-slice mCT (Siemens Healthineers) according to the guidelines for  $^{18}\text{F}$ -FDG PET and PET/CT imaging in pediatric oncology from the European Association of Nuclear Medicine (20). The imaging protocol included a minimum fasting time of 6 h. The  $^{18}\text{F}$ -FDG dose was adjusted according to body weight following European Association of Nuclear Medicine guidelines, and  $^{18}\text{F}$ -FDG PET/CT scans were performed from the mid thigh to the skull base, 60 min after intravenous administration.  $^{18}\text{F}$ -FDG PET/CT images were corrected for scatter and attenuation on the basis of low-dose CT information.

### $^{18}\text{F}$ -FDG PET/CT for PTLT Detection

$^{18}\text{F}$ -FDG PET/CT scans performed for PTLT detection were retrospectively reviewed by 3 readers (2 experienced nuclear medicine physicians and 1 research fellow) using syngo.via software (Siemens Healthineers). The readers reviewed the scans independently from each other and were masked to other imaging findings, pathology results, and clinical findings. Any metabolic active focus that could not be related to physiologic distribution, or any focus with an  $^{18}\text{F}$ -FDG uptake higher than the surrounding tissues and not suggestive of other pathology, was regarded as PTLT-positive. If a metabolic active focus was visualized but could not with certainty be attributed to PTLT or other diseases (such as infectious, inflammatory, or other malignant lesions), the  $^{18}\text{F}$ -FDG PET/CT scan result was considered ambiguous. A differential diagnosis was noted when deemed relevant by the reader. Discordant results between readers were reevaluated in a consensus meeting and conclusively classified as PTLT-positive or PTLT-negative. False-positive and false-negative scans were reevaluated to determine potential patterns. Histopathologic examinations were used as a reference standard for PTLT diagnosis. Two experienced hematopathologists were consulted to clarify morphology for 12 patients whose original pathology report was not sufficiently clear. In the case of a PTLT-negative biopsy or lack of tissue for pathologic examination, a 2-y follow-up period without preemptive PTLT therapy was accepted as the reference standard. In

adults, absence of lymphoma during this period has been shown to be an accurate marker for lack of disease in other lymphomas (21,22). True-positive scans were those interpreted as PTLT-positive on  $^{18}\text{F}$ -FDG PET/CT and confirmed by histopathologic examination to be PTLT within 2 y. True-negative scans were those interpreted as PTLT-negative on  $^{18}\text{F}$ -FDG PET/CT and with no signs of PTLT being identified within a 2-y follow-up. False-positive scans were those interpreted as PTLT-positive on  $^{18}\text{F}$ -FDG PET/CT and with no signs of PTLT being identified within a 2-y follow-up. False-negative scans were those interpreted as PTLT-negative on  $^{18}\text{F}$ -FDG PET/CT but confirmed by histopathologic examination to be PTLT within 2 y.

### $^{18}\text{F}$ -FDG PET/CT for Response Assessment

All  $^{18}\text{F}$ -FDG PET/CT scans performed for response assessment were reevaluated according to the Lugano criteria with masking to other imaging findings, pathology results, and clinical findings (23). Scans with a score of 1–3 were considered indicative of complete remission, whereas scores of 4–5 were considered to represent partial response, stable disease, or progressive disease.  $^{18}\text{F}$ -FDG PET/CT response assessment scans for which a reference standard was available were classified as true-positive, true-negative, false-positive, or false-negative for the presence of PTLT. For interim scans, histopathologic examination was accepted as the reference standard for PTLT confirmation. For end-of-treatment scans, the accepted reference standard for PTLT confirmation was a confirmatory biopsy or high suspicion of death due to PTLT, whereas a negative 2-y follow-up period was accepted as confirmation of absence of disease.

### Statistical Analysis

Baseline patient characteristics were summarized using median  $\pm$  SD with interquartile range. The sensitivity, specificity, positive predictive value, and negative predictive value of  $^{18}\text{F}$ -FDG PET/CT for the detection of PTLT on a patient-based analysis were calculated, along with the 95% confidence interval. Interobserver variability among the 3 observers was calculated using the Fleiss  $\kappa$ . The  $\kappa$ -value was interpreted according to the method of Landis and Koch: poor (0–0.20), fair (0.21–0.40), moderate (0.41–0.60), good (0.61–0.80), and perfect agreement (0.81–1) (24). Because of the relatively small and heterogeneous population, and the inconsistent availability of a reference standard, the diagnostic yield of  $^{18}\text{F}$ -FDG PET/CT in the response assessment setting for PTLT was only descriptively analyzed. Statistical analyses were performed using SPSS, version 23.0 (IBM Corp.).

## RESULTS

### Patients

Thirty-three potentially eligible patients were identified. Four patients were excluded because they did not fulfill the reference standard criteria (3 patients did not have a 2-y follow-up and 1 patient received preemptive treatment with rituximab after a negative biopsy). One patient was excluded because the suspected tumor had been fully resected before  $^{18}\text{F}$ -FDG PET/CT. Because of PTLT suspicion on multiple occasions with an interval of more than 2 y between different  $^{18}\text{F}$ -FDG PET/CT scans, 2 patients had 2 eligible scans and 1 patient had 3 eligible scans. Thus, in total, 32  $^{18}\text{F}$ -FDG PET/CT scans in 28 patients were included. Common indications for requesting an  $^{18}\text{F}$ -FDG PET/CT scan are described in Table 1. There were 13 (46%) boys and 15 (54%) girls (Table 2). Patient age ranged from 1 to 18 y, with a median age of 4 y. Liver was the most frequently transplanted organ ( $n = 20$ , 71.4%), followed by lung ( $n = 3$ , 10.7%), multiple organs ( $n = 2$ , 7.1%), heart ( $n = 1$ , 3.6%), kidney ( $n = 1$ , 3.6%), and small bowel ( $n = 1$ , 3.6%). According to the reference standard, 14

**TABLE 1**  
Indications for <sup>18</sup>F-FDG PET/CT

Indication	n
Blood panel disturbances (e.g., complete blood count and biochemistry)	7 (21.9%)
Epstein-Barr virus DNAemia	19 (59.4%)
Physical symptoms (e.g., B symptoms and enlarged lymph nodes)	13 (40.6%)
After previous examinations	
Colonoscopy	4 (12.5%)
Conventional radiography	1 (3.1%)
CT	2 (6.3%)

B symptoms = fever, night sweats, and weight loss.  
Multiple indications were possible for a single scan.

patients (50%) were diagnosed with PTLD, of which 5 cases (35.7%) were nondestructive, 3 (21.5%) polymorphic, 5 (35.7%) monomorphic, and 1 (7.1%) classic Hodgkin lymphoma.

**Diagnostic Performance of <sup>18</sup>F-FDG PET/CT for PTLD Detection**

After a consensus meeting by the 3 readers, 7 scans were considered as PTLD-positive and 25 as PTLD-negative. A PTLD-positive biopsy, a PTLD-negative biopsy with 2 y of follow-up without preemptive therapy, and a 2-y follow-up without preemptive therapy or biopsy were used as the reference standard for 14 (43.8%), 10 (31.2%), and 8 (25%) of the <sup>18</sup>F-FDG PET/CT

**TABLE 2**  
Patient Characteristics (n = 28)

Characteristic	Data
Age at diagnosis (y)	
Median	4
Range	1–18
Interquartile range	1–12
Sex (n)	
Male	13 (46%)
Female	15 (54%)
Transplanted organ (n)	
Liver	20 (71.4%)
Lung	3 (10.7%)
Multiorgan	2 (7.1%)
Heart	1 (3.6%)
Kidney	1 (3.6%)
Small bowel	1 (3.6%)
Histology (n)	
Nondestructive	5 (35.7%)
Polymorphic	3 (21.5%)
Monomorphic	5 (35.7%)
Classic Hodgkin lymphoma	1 (7.1%)

**TABLE 3**  
Classification of <sup>18</sup>F-FDG PET/CT Scans (n = 32)

Finding	PTLD present (n)	PTLD absent (n)
<sup>18</sup> F-FDG PET/CT-positive	7 (21.9%)	0
<sup>18</sup> F-FDG PET/CT-negative	7 (21.9%)	18 (56.2%)

scans, respectively. According to the reference standard, 18 (56.2%) <sup>18</sup>F-FDG PET/CT scans were true-negative, 7 (21.9%) true-positive, 0 false-positive, and 7 (21.9%) false-negative (Table 3). On a patient-based analysis, the sensitivity of <sup>18</sup>F-FDG PET/CT for the detection of PTLD was 50%, specificity was 100%, positive predictive value was 100%, and negative predictive value was 72% (Table 4).

**Causes of False-Negative <sup>18</sup>F-FDG PET/CT Scans for PTLD Detection**

Seven of 32 (21.9%) <sup>18</sup>F-FDG PET/CT scans performed because of clinical suspicion of PTLD were false-negative (Table 5). Five of the 7 false-negative cases had biopsy-confirmed non-destructive PTLD. On <sup>18</sup>F-FDG PET/CT, 3 patients had symmetric <sup>18</sup>F-FDG uptake (higher than liver <sup>18</sup>F-FDG uptake) in and limited to the Waldeyer's ring, whereas 2 had symmetric <sup>18</sup>F-FDG uptake (higher than liver <sup>18</sup>F-FDG uptake) in the Waldeyer's ring along with <sup>18</sup>F-FDG-avid (higher than liver <sup>18</sup>F-FDG uptake) cervical lymph nodes (Fig. 1). The remaining 2 false-negative patients had biopsy-confirmed polymorphic PTLD in the small intestines, which was interpreted as physiologic intestinal <sup>18</sup>F-FDG uptake in both cases. In these 2 patients, no focal <sup>18</sup>F-FDG-avid lesions were observed; rather, diffuse <sup>18</sup>F-FDG uptake (higher than liver <sup>18</sup>F-FDG uptake) was observed in the gastrointestinal tract. One patient initially had a false-negative scan with polymorphic PTLD in the ileum. After adjustment of immunosuppression and watchful waiting, the patient developed a monomorphic PTLD, which was visualized by subsequent <sup>18</sup>F-FDG PET/CT (Fig. 2). Abdominal diagnostic CT was performed in 1 of these 2 patients, but the clinical radiology report mentioned no signs of PTLD.

**Interobserver Variability of <sup>18</sup>F-FDG PET/CT for PTLD Detection**

From a total of 32 <sup>18</sup>F-FDG PET/CT scans evaluated before the consensus meeting, discordant results were reported for 5. One <sup>18</sup>F-FDG PET/CT scan with symmetric <sup>18</sup>F-FDG uptake in

**TABLE 4**  
Diagnostic Performance of <sup>18</sup>F-FDG PET/CT in PTLD Detection

Analysis	Percentage	95% CI
Sensitivity	50	24–76
Specificity	100	78–100
Positive predictive value	100	56–100
Negative predictive value	72	50–87

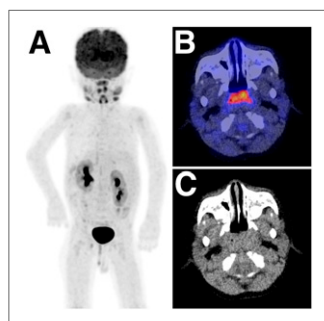
CI = confidence interval.

**TABLE 5**  
Description of False-Negative <sup>18</sup>F-FDG PET/CT Scans (n = 7)

Location	Readers' differential diagnosis*	Final diagnosis
Waldeyer's ring	Physiologic uptake	Nondestructive PTLD
Waldeyer's ring	Physiologic uptake	Nondestructive PTLD
Waldeyer's ring	Physiologic uptake	Nondestructive PTLD
Waldeyer's ring	Physiologic uptake	Nondestructive PTLD
Cervical lymph node	Physiologic uptake; reactive lymph nodes; PTLD	Nondestructive PTLD
Duodenum, mesenteric lymph nodes	Physiologic uptake	Polymorphic PTLD
Ileum	Physiologic uptake	Polymorphic PTLD

\*In order of most likely diagnosis.

the Waldeyer's ring and cervical lymph nodes was considered ambiguous for PTLD by 2 readers, who thought the findings could be interpreted as either reactive lymph nodes or PTLD. One case of <sup>18</sup>F-FDG uptake in the Waldeyer's ring and retroperitoneal lymph nodes was considered to be due to either inflammatory changes or PTLD by 2 readers. In 1 scan with focal <sup>18</sup>F-FDG uptake in the lung, 2 of 3 readers reported difficulties in distinguishing between PTLD and an infectious cause (i.e., fungal). Finally, in 1 scan with localized <sup>18</sup>F-FDG uptake in the cecum and in another scan with <sup>18</sup>F-FDG uptake throughout the whole duodenum and colon, the readers reported difficulty in differentiating whether the <sup>18</sup>F-FDG uptake was physiologic, due to PTLD, or due to other intestinal disease such as colitis. Of the 5 discordant <sup>18</sup>F-FDG PET/CT scans, 2 were true-positive, 2 true-negative, and 1 false-negative. The remaining 6 false-negatives scans were reported as PTLD-negative by all readers. The interobserver variability was found to be good, at a  $\kappa$ -value of 0.74 (95% confidence interval, 0.58–0.86).



**FIGURE 1.** Two-year-old boy 1 y after receiving liver transplant because of biliary atresia. <sup>18</sup>F-FDG PET/CT was requested after prolonged fever. False-negative <sup>18</sup>F-FDG PET/CT scan with biopsy confirmed nondestructive PTLD in adenoid or tonsils. (A) Maximum-intensity-projection <sup>18</sup>F-FDG PET shows almost symmetric uptake in Waldeyer's ring and salivary glands. (B) Axial fused <sup>18</sup>F-FDG PET/CT shows almost symmetric uptake in adenoids. This pattern of <sup>18</sup>F-FDG uptake was interpreted as physiologic. (C) Low-dose CT does not show any suggestive lesions.

#### <sup>18</sup>F-FDG PET/CT for Response Assessment

In all 14 patients who were diagnosed with PTLD, reduction of immunosuppression was the cornerstone therapy. First-line treatment was performed with rituximab in 8 patients; rituximab, cyclophosphamide, vincristine, and prednisone in 2 patients; watchful waiting in 2 patients; rituximab, vincristine, etoposide, prednisone, and doxorubicin in 1 patient; and tumor resection in 1 patient. Two patients were lost to follow-up after diagnosis. <sup>18</sup>F-FDG PET/CT was used for interim response assessment in 6 patients on 12 occasions; of these, biopsy correlation was possible for 5 scans. According to the pathology

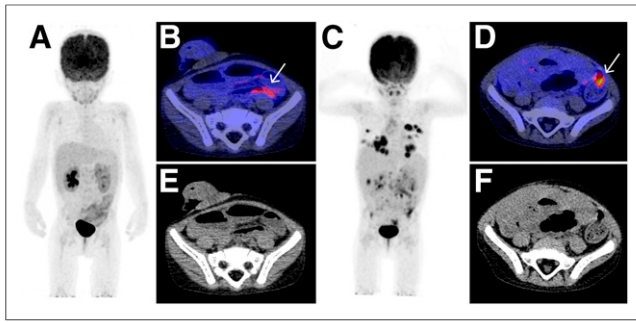
results, there were 3 true-positive and 2 false-positive interim <sup>18</sup>F-FDG PET/CT scans. False-positive scan results were due to therapy-induced reactive changes (Fig. 3). End-of-treatment <sup>18</sup>F-FDG PET/CT was used in 8 patients on 9 occasions, and a reference standard was available on all occasions. There were 1 true-positive, 4 true-negative, 3 false-positive, and 1 false-negative end-of-treatment <sup>18</sup>F-FDG PET/CT scans. In 2 false-positive cases, a negative 2-y follow-up period did not reveal any PTLD, and in 1 case, biopsy revealed follicular hyperplasia without evidence of PTLD. For the false-negative end-of-treatment <sup>18</sup>F-FDG PET/CT, a biopsy obtained 2 mo after a PTLD-negative scan revealed monomorphic PTLD.

#### DISCUSSION

This study aimed to evaluate the diagnostic performance of <sup>18</sup>F-FDG PET/CT for the detection of PTLD in the pediatric population and to explore its feasibility in the response assessment setting. The results suggest that <sup>18</sup>F-FDG PET/CT has a good specificity and positive predictive value but low to moderate sensitivity and negative predictive value for the detection of PTLD in children, especially when disease is localized in the Waldeyer's ring, cervical lymph nodes or gastrointestinal tract. A positive <sup>18</sup>F-FDG PET/CT scan may therefore confirm PTLD suspicion, but a negative <sup>18</sup>F-FDG PET/CT does not rule out PTLD.

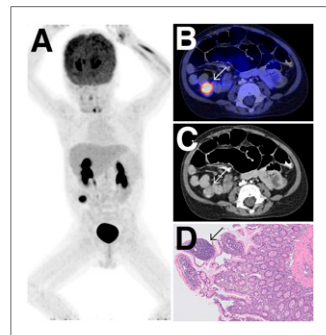
Studies on the clinical utility of <sup>18</sup>F-FDG PET/CT in pediatric PTLD are limited and often combined with adult PTLD cohorts (13,16,17,19). However, considering the essential differences in pathology and presentation of this disease in the 2 population groups, diagnostic performance analyses should be performed separately for children. To date, studies on pediatric PTLD patients have been descriptive in nature, comparing <sup>18</sup>F-FDG PET or PET/CT with other imaging modalities (such as CT and MRI) on a lesion by lesion basis and evaluating how additional detected lesions on <sup>18</sup>F-FDG PET or PET/CT affected staging and treatment (14,15,18,25). Study populations were often small (range, 7–34 patients), and in 2 of the 4 previous studies on this topic, stand-alone <sup>18</sup>F-FDG PET was used instead of the hybrid <sup>18</sup>F-FDG PET/CT (14,18). Furthermore, a diagnostic performance analysis (in terms of sensitivity, specificity, positive predictive value, and negative predictive value) for the detection of PTLD was not performed in any of these previous studies.

Although results from mixed and adult cohorts suggest <sup>18</sup>F-FDG PET/CT as a viable imaging modality for PTLD detection



**FIGURE 2.** Three-year-old girl 2 mo after small-bowel transplantation because of unexplained absorption disorder.  $^{18}\text{F}$ -FDG PET/CT was requested during clinical admission due to fever and leukopenia. (A, B, and E) False-negative  $^{18}\text{F}$ -FDG PET/CT scan with biopsy-confirmed polymorphic PTLD in ileum. Maximum-intensity-projection  $^{18}\text{F}$ -FDG PET (A) and axial fused  $^{18}\text{F}$ -FDG PET/CT (B) show diffuse uptake in small bowel (white arrow), interpreted as physiologic uptake; on low-dose CT (E), distended gas-filled bowels and postoperative ileostomy are shown. (C, D, and F) Same patient 6 mo after reduction in immunosuppression and watchful waiting: true-positive  $^{18}\text{F}$ -FDG PET/CT scan with biopsy-confirmed monomorphic intestinal PTLD. Maximum-intensity-projection  $^{18}\text{F}$ -FDG PET (C) shows multiple intrapulmonary, mesenteric, and intestinal  $^{18}\text{F}$ -FDG-active lesions; axial fused  $^{18}\text{F}$ -FDG PET/CT (D) shows focal  $^{18}\text{F}$ -FDG uptake in small bowel suggestive of PTLD without evident abnormalities on low-dose CT (F).

at diagnosis (sensitivity, 89%–85%; specificity, 91%–89%; positive predictive value, 91%–83%; and negative predictive value, 92%–87%), the high number of false-negative cases in our pediatric patient population impacted the sensitivity and negative predictive value of  $^{18}\text{F}$ -FDG PET/CT for PTLD detection (16,26–28). False-negative results in our current study were confirmed in the Waldeyer's ring ( $n = 4$ ), cervical lymph nodes ( $n = 1$ ), and small bowel ( $n = 2$ ), which were interpreted as physiologic uptake but proved to be either nondestructive or polymorphic PTLD.



**FIGURE 3.** Three-year-old boy 2 y after liver transplantation because of biliary atresia.  $^{18}\text{F}$ -FDG PET/CT was requested after 3 cycles of rituximab, cyclophosphamide, vincristine, and prednisone therapy. False-positive interim  $^{18}\text{F}$ -FDG PET/CT scan confirmed after biopsy via colonoscopy revealed therapy-induced reactive changes in the cecum. (A and B) Maximum-intensity-projection  $^{18}\text{F}$ -FDG PET (A) and axial fused  $^{18}\text{F}$ -FDG PET/CT (B) show  $^{18}\text{F}$ -FDG-avid lesion in cecum (arrow). (C) On diagnostic CT, spheric mass is seen in cecum (arrow). (D)  $\times 100$  magnification with hematoxylin and eosin staining shows lymphoid infiltration without abnormal cells (arrow).

In pediatric patients particularly, attention should also be paid to the head and neck region. Concerns about false-negative results in the tonsils have been previously reported by Vali et al. (18). Nondestructive PTLD tends to occur at a younger age and is also often limited to the Waldeyer's ring (1,7). However, uptake in the Waldeyer's ring is commonly reported in children and not necessarily indicative of pathology, leading to potential misinterpretation of uptake in this area

as physiologic (29). Additionally, although reactive  $^{18}\text{F}$ -FDG-avid lymph nodes in the cervical region are also often reported in children, cervical malignant lymphadenopathy seems to occur more frequently in PTLD patients than in immunocompetent lymphoma patients (30). The gastrointestinal tract is also a commonly reported PTLD location in pediatric patients (7,31). Physiologic uptake in the gastrointestinal tract may obscure or mimic pathology and give rise to false-negative results (32).

Despite a low to moderate sensitivity and negative predictive value for the detection of PTLD at diagnosis,  $^{18}\text{F}$ -FDG PET/CT retains clinical utility in the management of pediatric PTLD patients. Because of the high number of false-negative scans in the tonsils or adenoids, physicians must remain alert for signs that might indicate the presence of disease, such as a high Epstein-Barr virus DNA load and tonsillar hypertrophy (4,33). Nevertheless, if a biopsy is positive for nondestructive PTLD in the tonsils or adenoids but the  $^{18}\text{F}$ -FDG PET/CT findings are interpreted as PTLD-negative, the disease might be focal and therapy limited to reduction of immunosuppression (or potentially rituximab) and clinical follow-up. With regard to uptake in the gastrointestinal tract, 1 study has demonstrated that patient preparation with *N*-butylscopolamine (Buscopan; Boehringer Ingelheim) reduces artifacts in the bowel and improves accuracy (34). Furthermore, CT has also been suggested as a more sensitive modality for PTLD lesion detection in bowel and stomach (18). Patient-specific preparation and an abdominal diagnostic CT scan may be necessary in a selected group of patients if lower-gastrointestinal-tract PTLD is suspected. The high specificity and positive predictive value of  $^{18}\text{F}$ -FDG PET/CT in the disease detection setting are clinically relevant, because concerns about false-positive  $^{18}\text{F}$ -FDG PET/CT scans, predominantly due to inflammation or other malignancies, are often encountered in the literature (26,35). However, compared with adults, the risk of a malignancy (other than PTLD) is decreased in pediatric transplant patients—a fact that may explain the lack of false-positive scans in the disease detection setting in this study (36).

Regarding the potential contribution of  $^{18}\text{F}$ -FDG PET/CT during treatment evaluation in pediatric PTLD, there were 40% (2/5) false-positive interim  $^{18}\text{F}$ -FDG PET/CT scans. For end-of-treatment  $^{18}\text{F}$ -FDG PET/CT, there were 33% (3/9) false-positive and 11% (1/9) false-negative scans. Interim false-positive results were predominantly due to therapy-induced reactive changes. This finding is in line with a systematic review of immunocompetent lymphoma patients by Adams et al. (37), who raised concerns about high proportions of false-positives, with false-positive results reported in 55.7% of all  $^{18}\text{F}$ -FDG-avid lesions that were biopsied during and at the end of treatment (most being due to inflammatory changes).

The retrospective nature of this study constitutes a significant limitation. Important variables such as patient selection and the timing of  $^{18}\text{F}$ -FDG PET/CT could not be controlled. Because there are currently no guidelines on the use of  $^{18}\text{F}$ -FDG PET/CT for the diagnosis of PTLD in pediatric patients, each medical department defined its own criteria for requesting a scan. Previous examinations performed before  $^{18}\text{F}$ -FDG PET/CT in the included patients may have influenced the a priori incidence of PTLD and, therefore, diagnostic performance. In addition to potentially inducing a selection bias, the lack of control on patient management variables may also have affected  $^{18}\text{F}$ -FDG PET/CT diagnostic performance during treatment evaluation. Taking into consideration the lack of

literature and the limitations of retrospective studies, future research on this topic should focus on prospective and multicenter studies.

## CONCLUSION

<sup>18</sup>F-FDG PET/CT showed a good specificity and positive predictive value but a low to moderate sensitivity and negative predictive value for the detection of PTLD in a 28-pediatric-patient cohort with clinical suspicion of this disease. False-negative results were confirmed in the Waldeyer's ring, cervical lymph nodes, or small bowel with either nondestructive or polymorphic PTLD subtypes. <sup>18</sup>F-FDG PET/CT appears to have a limited role in the setting of response assessment for pediatric PTLD, given the observed high proportions of false-positives both at interim and end-of-treatment evaluations.

## DISCLOSURE

No potential conflict of interest relevant to this article was reported.

## KEY POINTS

**QUESTION:** Is <sup>18</sup>F-FDG PET/CT an accurate imaging modality for PTLD detection in pediatric patients with suspicion of the disorder?

**PERTINENT FINDINGS:** In this single-center retrospective study including 28 patients and 32 scans, the sensitivity, specificity, positive predictive value, and negative predictive value of <sup>18</sup>F-FDG PET/CT for the detection of PTLD in children with a clinical suspicion of this disease were 50% (7/14), 100% (18/18), 100% (7/7), and 72% (18/25), respectively. False-negative results were confirmed in the Waldeyer's ring, cervical lymph nodes, and small bowel with either nondestructive or polymorphic PTLD subtypes.

**IMPLICATIONS FOR PATIENT CARE:** Clinicians should be aware of the inherent limitations of <sup>18</sup>F-FDG PET/CT, paying particular attention to the potential for a focus of disease in the Waldeyer's ring, cervical lymph nodes, and gastrointestinal tract of pediatric patients.

## REFERENCES

1. Swerdlow SH, Campo E, Harris NL, et al. Post-transplant lymphoproliferative disorders (PTLD). In: Swerdlow S, Campo E, Harris N, et al., eds. *World Health Organization Classification of Tumours of Haematopoietic and Lymphoid Tissues*. Vol 2. 4th ed. Lyon, France: International Agency for Research on Cancer;2017:453–462.
2. Taylor AL, Marcus R, Bradley JA. Post-transplant lymphoproliferative disorders (PTLD) after solid organ transplantation. *Crit Rev Oncol Hematol*. 2005;56:155–167.
3. Dierickx D, Habermann TM. Post-transplantation lymphoproliferative disorders in adults. *N Engl J Med*. 2018;378:549–562.
4. Parker A, Bowles K, Bradley JA, et al. Diagnosis of post-transplant lymphoproliferative disorder in solid organ transplant recipients: BCSH and BTS guidelines. *Br J Haematol*. 2010;149:675–692.
5. Llaurador G, McLaughlin L, Wistinghausen B. Management of post-transplant lymphoproliferative disorders. *Curr Opin Pediatr*. 2017;29:34–40.
6. Opelz G, Döhler B. Lymphomas after solid organ transplantation: a collaborative transplant study report. *Am J Transplant*. 2004;4:222–230.
7. L'Huillier AG, Dipchand AI, Ng VL, et al. Posttransplant lymphoproliferative disorder in pediatric patients: survival rates according to primary sites of occurrence and a proposed clinical categorization. *Am J Transplant*. 2019;19:2764–2774.
8. Allen U, Hebert D, Moore D, Dror Y, Wasfy S. Epstein-Barr virus-related post-transplant lymphoproliferative disease in solid organ transplant recipients, 1988–97: a Canadian multi-centre experience. *Pediatr Transplant*. 2001;5:198–203.
9. Al-Mansour Z, Nelson BP, Evens AM. Post-transplant lymphoproliferative disease (PTLD): risk factors, diagnosis, and current treatment strategies. *Curr Hematol Malig Rep*. 2013;8:173–183.
10. Ligeti K, Müller LP, Müller-Tidow C, Weber T. Risk factors, diagnosis, and management of posttransplant lymphoproliferative disorder: improving patient outcomes with a multidisciplinary treatment approach. *Transpl Res Risk Manag*. 2017;9:1–14.
11. Styczynski J, Gil L, Tridello G, et al. Response to rituximab-based therapy and risk factor analysis in Epstein Barr virus-related lymphoproliferative disorder after hematopoietic stem cell transplant in children and adults: a study from the Infectious Diseases Working Party of the European Group for Blood and Marrow Transplantation. *Clin Infect Dis*. 2013;57:794–802.
12. Végso G, Hajdu M, Sebestyén A. Lymphoproliferative disorders after solid organ transplantation: classification, incidence, risk factors, early detection and treatment options. *Pathol Oncol Res*. 2011;17:443–454.
13. Blaes AH, Cioc AM, Froelich JW, Peterson BA, Dunitz JM. Positron emission tomography scanning in the setting of post-transplant lymphoproliferative disorders. *Clin Transplant*. 2009;23:794–799.
14. von Falck C, Maecker B, Schirg E, et al. Post transplant lymphoproliferative disease in pediatric solid organ transplant patients: a possible role for <sup>18</sup>F-FDG-PET/CT in initial staging and therapy monitoring. *Eur J Radiol*. 2007;63:427–435.
15. Guerra-García P, Hirsch S, Levine DS, Taj MM. Preliminary experience on the use of PET/CT in the management of pediatric post-transplant lymphoproliferative disorder. *Pediatr Blood Cancer*. 2017;64:e26685.
16. Panagiotidis E, Quigley A-M, Pencharz D, et al. <sup>18</sup>F-fluorodeoxyglucose positron emission tomography/computed tomography in diagnosis of post-transplant lymphoproliferative disorder. *Leuk Lymphoma*. 2014;55:515–519.
17. Takehana CS, Twist CJ, Mosci C, Quon A, Mittra E, Iagaru A. <sup>18</sup>F-FDG PET/CT in the management of patients with post-transplant lymphoproliferative disorder. *Nucl Med Commun*. 2014;35:276–281.
18. Vali R, Punnett A, Bajno L, Moineddin R, Shammam A. The value of <sup>18</sup>F-FDG PET in pediatric patients with post-transplant lymphoproliferative disorder at initial diagnosis. *Pediatr Transplant*. 2015;19:932–939.
19. O'Conner AR, Franc BL. FDG PET imaging in the evaluation of post-transplant lymphoproliferative disorder following renal transplantation. *Nucl Med Commun*. 2005;26:1107–1111.
20. Stauss J, Franzius C, Pfluger T, et al. Guidelines for <sup>18</sup>F-FDG PET and PET-CT imaging in paediatric oncology. *Eur J Nucl Med Mol Imaging*. 2008;35:1581–1588.
21. El-Galaly TC, Jakobsen LH, Hutchings M, et al. Routine imaging for diffuse large B-cell lymphoma in first complete remission does not improve post-treatment survival: a Danish-Swedish population-based study. *J Clin Oncol*. 2015;33:3993–3998.
22. Maurer MJ, Ghesquière H, Jais JP, et al. Event-free survival at 24 months is a robust end point for disease-related outcome in diffuse large B-cell lymphoma treated with immunochemotherapy. *J Clin Oncol*. 2014;32:1066–1073.
23. Cheson BD, Fisher RI, Barrington SF, et al. Recommendations for initial evaluation, staging, and response assessment of Hodgkin and non-Hodgkin lymphoma: the Lugano classification. *J Clin Oncol*. 2014;32:3059–3068.
24. Landis JR, Koch GG. The measurement of observer agreement for categorical data. *Biometrics*. 1977;33:159–174.
25. Yang J, Zhuang H. The role of <sup>18</sup>F-FDG PET/CT in the evaluation of pediatric transplant patients. *Hell J Nucl Med*. 2015;18:136–139.
26. Dierickx D, Tousseyn T, Requilé A, et al. The accuracy of positron emission tomography in the detection of posttransplant lymphoproliferative disorder. *Hematologica*. 2013;98:771–775.
27. Montes de Jesus FM, Kwee TC, Kahle XU, et al. Diagnostic performance of FDG-PET/CT of post-transplant lymphoproliferative disorder and factors affecting diagnostic yield. *Eur J Nucl Med Mol Imaging*. 2020;47:529–536.
28. Treglia G, Ceriani L. <sup>18</sup>F-FDG PET/CT in patients with post-transplant lymphoproliferative disorders: so far so good. *Eur J Nucl Med Mol Imaging*. 2020; 47:523–524.
29. Shammam A, Lim R, Charron M. Pediatric FDG PET/CT: physiologic uptake, normal variants, and benign conditions. *Radiographics*. 2009;29:1467–1486.
30. Vali R, Bakari A, Marie E, et al. FDG uptake in cervical lymph nodes in children without head and neck cancer. *Pediatr Radiol*. 2017;47:860–867.

31. McDonald RA, Smith JM, Ho M, et al. Incidence of PTLD in pediatric renal transplant recipients receiving basiliximab, calcineurin inhibitor, sirolimus and steroids. *Am J Transplant.* 2008;8:984–989.
32. Prabhakar HB, Sahani DV, Fischman AJ, Mueller PR, Blake MA. Bowel hot spots at PET-CT. *Radiographics.* 2007;27:145–159.
33. Marques HH, Shikanai-Yasuda MA, de Azevedo LSF, et al. Management of post-transplant Epstein-Barr virus-related lymphoproliferative disease in solid organ and hematopoietic stem cell recipients. *Rev Soc Bras Med Trop.* 2014;47:543–546.
34. Emmott J, Sanghera B, Chambers J, Wong WL. The effects of N-butylscopolamine on bowel uptake: an  $^{18}\text{F}$ -FDG PET study. *Nucl Med Commun.* 2008;29:11–16.
35. Montes HH, de Jesus FM, Kwee TC, Nijland M, et al. Performance of advanced imaging modalities at diagnosis and treatment response evaluation of patients with post-transplant lymphoproliferative disorder: a systematic review and meta-analysis. *Crit Rev Oncol Hematol.* 2018;132:27–38.
36. Bhat M, Mara K, Dierkhising R, Watt KDS. Immunosuppression, race, and donor-related risk factors affect de novo cancer incidence across solid organ transplant recipients. *Mayo Clin Proc.* 2018;93:1236–1246.
37. Adams HJA, Kwee TC. Proportion of false-positive lesions at interim and end-of-treatment FDG-PET in lymphoma as determined by histology: systematic review and meta-analysis. *Eur J Radiol.* 2016;85:1963–1970.

### Erratum

In the article “ $^{11}\text{C}$ -PBR28 and  $^{18}\text{F}$ -PBR111 Detect White Matter Inflammatory Heterogeneity in Multiple Sclerosis,” by Datta et al. (*J Nucl Med.* 2017;58:1477–1482), the name of one of the authors was misspelled. “Nicola D. Stefano” should be “Nicola De Stefano.” The authors regret the error.