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The role of accelerated ageing in aberrant lung tissue repair and remodelling in COPD

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CHAPTER 2

Lung ageing and COPD: is there a role for ageing in abnormal tissue repair?

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ABSTRACT

COPD is the fourth leading cause of death worldwide with increasing prevalence, in particular in the elderly. COPD is characterized by abnormal tissue repair resulting in (small) airways disease and emphysema. There is accumulating evidence that ageing hallmarks are prominent features of COPD. These ageing hallmarks have been described in different subsets of COPD patients, in different lung compartments, and also in a variety of cell types, and thus might contribute to different COPD phenotypes. A better understanding of the main differences and similarities between normal lung ageing and the pathology of COPD may improve our understanding of the mechanisms driving COPD pathology, in particular in those patients that develop the most severe form of COPD at a relatively young age, i.e. severe early onset COPD patients. In this review, after introducing the main concepts of lung ageing and COPD pathology, we focus on the role of (abnormal) ageing in lung remodelling and repair in COPD. We discuss the current evidence for the involvement of ageing hallmarks in these pathologic features of COPD. In the last part, we highlight potential novel treatment strategies and opportunities for future research based on our current knowledge of abnormal lung ageing in COPD.

What is ageing?

As the world indicates, ageing is a process that mainly affects elderly people. With the quickly growing elderly population, the negative aspects of ageing are becoming increasingly apparent. Ageing is defined as the progressive decline in homeostasis after the reproductive phase is complete, which results in increased risk of disease or death (1). As such, ageing is one of the main driving forces of the development and increasing burden of non-communicable diseases (NCDs), i.e. chronic diseases. Worldwide, NCDs are the leading cause of mortality and responsible for 38 million deaths each year. Of these deaths, 4 million can be attributed to respiratory diseases (2). In many of the NCDs including ischaemic heart disease, diabetes, Alzheimer's disease and chronic obstructive pulmonary disease (COPD), it is proposed that acceleration of the normal ageing process is involved in the disease pathogenesis (3).

In this review we will first describe the processes involved in the normal ageing lung and the disease pathology of COPD and then summarize the similarities and the differences. We will specifically focus on the role of abnormal ageing in lung remodelling and repair in COPD and discuss the current evidence for ageing hallmarks in the pathologic features of COPD. Finally, we will discuss potential novel treatment strategies based on the current evidence for lung ageing in COPD.

The ageing lung

On average, the human lung is growing until 10-12 years of age and further matures until it reaches its maximum function at approximately 20 years of age for females and 25 years of age for males (4). From then on lung function progressively declines with increasing age as a consequence of structural and physiological changes of the lung (4).

To start with the structural changes of the ageing lung, we can broadly divide these structural changes in three categories: changes in lung structure, changes in the chest wall and changes in respiratory muscles (5). The changes in the structure of the lung are mainly attributed to an increase in the size of the alveolar space without any inflammation or alveolar wall destruction, so called 'senile emphysema'. This microscopic emphysema increases in a linear fashion with age in non-smokers whereas when smoking a progressive increase in alveolar space size can be observed in specific (susceptible) individuals only (6-8). Senile emphysema might be a consequence of loss of the supporting structure of the lung parenchyma (4,5). Additionally, it has been observed that the elastic recoil of the lung reduces with increasing age. It has been postulated that this phenomenon is rather caused by reduced surface tension forces from the alveoli due to increased individual diameter size than by changes in elastin and collagen in the lung parenchyma (5). Upon increasing age, the compliance of the chest wall decreases progressively, which can be explained by several, synergistically acting, age-related processes. Firstly, the shape of the thorax may change with age due to reduced thickness of intervertebral discs, leading to reduced intra thoracic volume. Secondly, age-associated osteoporosis may cause vertebral fractures resulting in changes in the shape of the thorax. Thirdly, the stiffness of the ribs increases with age, thereby enhancing the forces needed for movement of the chest (5,9). In general, muscle

strength diminishes with age. This loss in muscle strength is also reflected in the diaphragm, the most important respiratory muscle, and thus affects the breathing pattern (5). These structural changes of the ageing lung have a clear effect on the overall lung function and several physiologically parameters are altered upon ageing. Both the forced expiratory volume in one second (FEV_1) and forced vital capacity (FVC) are decreasing with age and the rate of decline has shown to be higher for males than for females. As a consequence of the reduced elastic recoil and compliance of the chest wall, the residual volume (RV) increases, while the vital capacity (VC) decreases (9). Of interest, the total lung capacity (TLC) does not change with increasing age, since the reduction in elastic recoil observed upon ageing is counterbalanced by the decrease in chest wall compliance and muscle strength of the chest (5,9). Although the distribution of alveolar ventilation and perfusion across the lungs is very heterogeneous as a consequence of the decline in alveolar surface area, density of lung capillaries and pulmonary capillary blood volume, the overall transfer capacity of the lung for carbon monoxide (TLCO) is reduced with increasing age. Clinically, this might influence the physical activity and the development of sleep-disordered breathing (5,9).

Next to changes in lung function with increasing age, the natural defence mechanisms of the lungs are also gradually less functional, leading to increased infection risk (4). For example, the antioxidant response to prohibit the accumulation of reactive oxygen species (ROS) is deteriorated in the ageing lung, consistent with an increase in ROS levels upon ageing (10). Furthermore, intercellular communications become less effective with ageing (11) contributing to two phenomena known as immunosenescence and inflammageing. The first relates to dampened immune responses following an infection or injury, and the second term relates to the chronic activation of immune responses in aged subjects in the absence of a real immunologic challenge (12). As a result of immunosenescence, innate and adaptive immune responses decrease with age, which is characterized by an increase in memory and effector cells at the expense of naïve T cells and the overall T cell repertoire (13,14). Of interest, several of the pro-inflammatory mediators associated with inflammageing, like tumour necrosis factor (TN)- α , interleukin (IL)-1 β and IL-6, are present as pro-inflammatory mediators in the senescence-associated secretory phenotype (SASP). Another factor contributing to increased inflammation in aged lungs is poor airway clearance of particles. Over time, muscles become atrophic, resulting in less strength for effective cough (15). Also, mucociliary clearance is known to be compromised with age (16), which might in particular contribute to viral and bacterial inflammation and thus acute exacerbations of lung diseases like COPD.

Pathology of COPD

COPD is a heterogeneous disease involving both the alveolar and airway compartment resulting in (small) airways disease and emphysema (Figure 1). The extent of pathologic changes in these different lung compartments is however variable in individual patients (17). The aetiology involves in general exposure to external noxious particles or gases. In the Western world this is in particular by (cigarette) smoking, and in the non-Western world mainly by indoor cooking. COPD pathology is driven by chronic inflammation (18-20), which

is still observed after stopping smoking (21) even after one or more years (22,23). The combination of the exposure and inflammation leads to lung tissue damage resulting in remodelling of the lung. This remodelling shows remarkable features: a common main aetiology, smoke exposure, leads to fibrosis (extracellular matrix increase) with thickening of (large and in a particular small) airway walls with lumen reduction and concurrently to emphysema with ECM destruction in the lung parenchyma. (19,24). Another main histopathologic feature of COPD is seen in the vasculature with in particular increased thickness of the arterioles, resulting in pulmonary hypertension as an important complication of COPD (25).

The chronic inflammation is mainly characterized by macrophages and (CD8⁺) T cells and can also show increased plasma cells, neutrophilic granulocytes and sometimes eosinophilic granulocytes (26-28). B-cells are also found, often in aggregates or small primary or secondary follicles (29-31). These have been described in association with airways as tertiary follicles, or reactive bronchus associated lymphoid tissue (BALT), but also have been observed scattered in the parenchyma (31,32). The presence of such follicles is variable, most pronounced in patients with severe COPD (30,32), and not only seen in COPD patients but also, to a lesser extent, in heavy smokers without COPD (31). As oligoclonality of these follicles has been shown (31). It is most likely that these are induced by local antigen stimulation (33-35). Cigarette smoke components, micro-organisms and matrix components (36) have been considered as etiologic factors for this antigenic stimulation, but none have been convincingly shown yet (34,37).

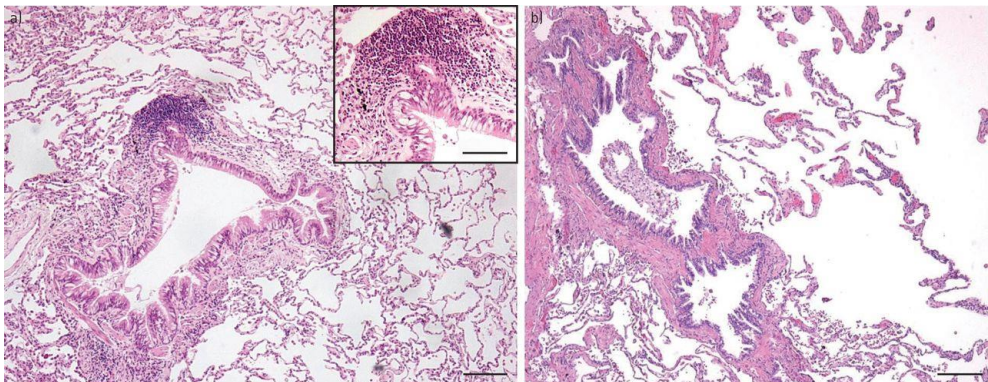


Figure 1: Pathologic changes in COPD. A) Characteristic picture of airways changes, with increase of goblet cells, a thickened airway wall with some adventitial inflammation and a small lymphoid follicle at the left upper side. Emphysema is hardly present here (haematoxylin and eosin, x200). Insert shows the magnification of part of the airway with a lymphoid follicle. **B)** At the left almost longitudinal cross-section of a small airway, while at the right side severe parenchymal destruction by emphysema (haematoxylin and eosin, x200).

In the large and small airways, epithelial changes are observed with increase of goblet cells, basal cell hyperplasia and squamous metaplasia, sometimes with dysplasia (20,38-40). Overall, these changes are in general more pronounced in the large airways, where the primary and most intensive exposure takes place. In the airway lumen increased and more sticky mucus can be present, which is produced by the increased number of goblet cells in combination with the enlarged mucus glands. The airway walls are thickened, caused by increased inflammation, increase of smooth muscle mass, increase in size of mucus glands, and, in due time, further changes with increase of extracellular matrix (ECM). The matrix changes in the airways are in general of a fibrotic nature with increase of collagens in the submucosa but also in the adventitia of smaller airways (41-44). Similar to asthma, a thickening of the basement membrane is seen, although more irregular and of different composition (43). An interesting finding was that in the peribronchial area of small airways in severe COPD an impressive reduction was found for proteoglycans, most prominent for decorin and to a lesser extent biglycan (45,46). A main characteristic of decorin is that it can bind TGF- β , one of the main cytokines regulating matrix production, which is consistently upregulated in COPD (47,48). Another important feature is that decorin is the main proteoglycan connecting collagen fibrils, in this way regulating rigidity of the collagen (48,49). As decorin is reduced in COPD, this will result in a very loose type of fibrosis, contributing to increased airway collapsibility and reduced peribronchial tensile strength of the parenchymal attachments (19,24). An interesting observation by Hogg *et al.* in a limited number of patients was a reduction in the number of small airways with increasing emphysema severity within COPD lungs suggesting that part of the airflow limitation within COPD is primarily caused by destruction of small airways (50,51).

Emphysema is the characteristic pathology of COPD that occurs in the lung parenchyma. This is characterized by net destruction of alveolar walls, as a result from increased destruction in combination with failing tissue repair (19). The destructive part is caused by an unbalance between exposure to oxygen radicals from cigarette smoke and neutrophils and proteases from macrophages and neutrophils, and their counterparts, oxygen scavengers and anti-proteases (52). Also in the parenchyma inflammation is present, but far less research has been published with respect to this compartment (53,54). Similar to the airways, cytotoxic (CD8+) T cells are important infiltrating cells in alveolar septa and arterioles in COPD when compared to non-COPD controls (53). Whereas neutrophils were and are considered as a main inflammatory cell contributing to emphysema, already in early studies in smokers no association was found between parenchymal neutrophils and the severity of the destructive index (55). In addition to possible direct effects of both smoke components and proteases released from inflammatory cells, indirect effects have been shown to destroy alveolar walls in COPD by inducing apoptosis of endothelial and alveolar type 2 epithelial cells, likely contributing to emphysema (56,57).

The important vascular changes in COPD are mainly seen in the arterioles. Here, intimal thickening with smooth muscle proliferation and increase of collagen and elastin, together with hyperplastic increase of the media have been observed (25,58-61). Initially

this was thought to be the result of hypoxia, but these events are also present in mild and early cases (60). More recently, the vascular changes have been attributed to “endothelial dysfunction” i.e. pathophysiological changes in the normal biochemical function of the endothelium (25,61,62). The end result are arterioles with a thickened wall, increased contraction and reduced lumen, but also with the reduced ability to vasodilate. Functionally, this leads to pulmonary hypertension which is a major cause of morbidity in COPD and a predictor of mortality (25,61).

Similarities and differences between lung ageing and COPD

One of the first reports on senile lungs compared with normal and emphysematous lungs by Verbeke *et al.* (63,64) demonstrated that the airspace enlargement in ageing, although comparable with smoking induced emphysema, differed in the fact that it was more regular in distribution without clear-cut destruction. Furthermore, increased thickening of alveolar septa was observed without inflammation or fibrosis with reduced density of the membranous bronchioles. They proposed the term senile lung for this condition. So, although similar in enlargement of airspaces at least part of the pathogenesis appears to be different. For loss of elasticity it is less clear whether this is a destructive effect in COPD or whether in both conditions there is an underlying defect in elastin fibrillogenesis. The functional effects on small airways in milder forms of emphysema are however comparable with the senile lung with loss of elastic recoil (64).

In the ageing process, Ito and Barnes (1) proposed that with increasing age the lung is less able to maintain organ integrity and protect itself against oxidative injury. Also, Kirkwood (65,66) indicated that cellular defects often cause inflammatory reactions contributing to damage, thereby causing a vicious circle of ongoing microscopic damage in due time with ageing. As yet, it is not readily clear to what extent these events are present during the total life course and when effects on tissue homeostasis become effective. In addition, it is not clear what the variation in the natural course of these events is with regard to their contribution to deterioration of the normal ageing lung. Taking the above mechanisms into account, several components observed in COPD, like the ongoing inflammation, unbalanced oxidative stress, and changes in the ECM are quite comparable as observed in the normal ageing lung. However in COPD, these changes will occur in general at an earlier age and to a larger extent compared to normal lung ageing. In the paragraphs below we will discuss in more detail whether premature or abnormal (lung) ageing aspects may or may not play a role in pathogenesis and natural course of COPD.

Lung ageing and COPD phenotypes

As described above, COPD is a very heterogeneous lung disease presented by different (mixed) phenotypes. Well-known phenotypes in COPD are chronic bronchitis with predominant airway related changes (inflammation and airway wall thickening) and increased mucus production, and emphysema with (severe) alveolar wall destruction, hyperinflation and impaired gas exchange. Other phenotypes of COPD are related to the number of exacerbations (i.e. the frequent exacerbator) (67,68) or the age of onset of the

disease, i.e. severe early onset COPD (SEO-COPD) (69). Given the difference in underlying pathology of these phenotypes, it can be envisaged that lung ageing is more or differently involved in some of these phenotypes than others. As discussed above, senile emphysema is an important hallmark of lung ageing and together with the structural changes, the ageing lung is in particular inclined to develop an emphysema-like phenotype. This is different from the bronchitis phenotype, where, apart from increased inflammation, there are very little similarities with the ageing lung and there is no indication of increased mucus production or airway wall thickening in the ageing lung, although decreased ciliary function with ageing likely contributes to increased coughing and decreased mucociliary clearance (15). The frequent exacerbator is an interesting phenotype as exacerbations are in general linked to infections and the susceptibility for infections increases with age (70,71). Moreover, age is a risk factor for COPD exacerbations (72) and hospital admissions for acute exacerbations of COPD (73).

Severe early-onset COPD is an interesting COPD phenotype with respect to ageing. Patients with this phenotype develop very severe COPD at a relatively young age, i.e. <53 according to Silverman *et al.* (69) and often with a relatively low number of pack years of smoking. This severe early-onset COPD (SEO-COPD) leads to a high personal burden and huge societal costs due to loss in working days and frequent hospitalizations. As these patients progress so quickly, we propose that, if accelerated ageing is an important contributor to COPD pathology, it should be most clear in these SEO-COPD patients. With respect to the pathology these patients are characterized by severe emphysema (69).

Ageing hallmarks in COPD

The main hallmarks of ageing were recently summarized in a review by Lopez-Orin (11) and this was followed by an overview of these hallmarks in lung ageing and lung disease (74). Broadly, the ageing hallmarks can be divided in processes affecting transcription (genomic instability, telomere attrition and epigenetic alterations), processes affecting the metabolism (loss of proteostasis, deregulated nutrient sensing and mitochondrial dysfunction) and cellular processes (cellular senescence, stem cell exhaustion and altered intracellular communication). We will now discuss the current knowledge about the possible role for these ageing hallmarks in COPD and mainly focus on findings in structural cells (alveolar and bronchial epithelial cells, smooth muscle cells and fibroblasts) and lung tissue. Subsequently, we will summarize the main evidence regarding the role of ageing hallmarks in disturbed repair and remodelling in COPD. All findings discussed in the paragraphs below are summarized in Table 1.

Transcription

Genomic instability

Ageing leads to increased DNA damage and to impaired ability to prevent and repair DNA damage. Several markers related to these features have also been demonstrated in COPD lungs and may contribute to the pathologic processes.

The DNA damage marker gamma- H2A histone family member X (γ -H2A.X) was increased in alveolar walls, including type I and type II epithelial cells and endothelial cells (75), as well as in small airways of COPD patients compared to controls (75,76). Another study, however, showed no differences in small airways in COPD versus control (77).

Smoke exposure increases γ -H2A.X levels in experimental animal models and cigarette smoke extract (CSE) treatment increases γ -H2A.X levels in bronchial epithelial cells and fibroblasts *in vitro* (76,78,79), suggesting an important role for oxidative stress. In addition, the anti-ageing protein sirtuin 6 (SIRT6) is considered to be protective against DNA damage and senescence. SIRT6 levels are decreased in lung tissue homogenates from COPD patients and overexpression and knockdown of SIRT6 in bronchial epithelial cells resulted in a decrease and an increase in γ H2A.X levels, respectively (79).

The DNA repair marker Ku86 was decreased in parenchymal lung tissue of COPD patients, including small airways (77,80), while no differences were observed in Ku70 expression in these samples. Ku70 is another DNA repair marker which was decreased in leukocytes derived from COPD patients and its expression was negatively correlated with age (80).

Table 1: Evidence for ageing hallmarks in COPD

Hallmark	Marker	COPD vs non-COPD	COPD cell origin	CSE treated cells	References
Genomic instability	γ-H2A.X	↑	Lung tissue sections, AT1, AT2, HBEC & PV-EC	HBEC, HFL1 & MRC-5	(75,76,78,79)
	Ku70	↓	Peripheral leukocytes		(80)
Telomere shortening	Ku80	↓	Lung homogenates		(77)
	length	↓	Lung homogenates AT2, PA-SMC, PV-EC & peripheral leukocytes	SAEC (COPD) & HFL1	(78,81-86)
	telomerase	↓	PV-EC		(84)
Epigenetic changes	TPP1	↓	Lung homogenates	SAEC & HFL1	(78)
	HDAC activity	↓	Lung homogenates and bronchial biopsies		(92)
Loss of proteostasis	SIRT-1 & -6	↓	Lung homogenates		(94,95)
	Autophagy	↑	HBEC	SAEC	(100)
	Autophagy	↓		HBEC (COPD)	(100)
	Autophagosomes	↑	Lung homogenates	HBEC & BEAS-2B	(79,99,100)
Deregulated nutrient sensing	Ubiquitin	↑	Lung homogenates		(98,100)
	p62	↑	Lung homogenates		(79,100)
	S6K (mTOR)	↑*	Lung homogenates & peripheral leukocytes	HBEC	(79,103)
	IGF1	↑	SAEC		(104)
Mitochondrial dysfunction	ROS	↑		HBEC, BEAS-2B & MRC-5	(76,108,109)
	Ox-DNA	↑	Lung homogenates		(77)
	lipid peroxidation	↑	Lung homogenates		(105,106)
	NO	↑	Lung homogenates		(106)
	mitophagy	↑	Lung homogenates	HBEC & BEAS-2B	(108,109)
	Antioxidant	↓	Lung homogenates & HBEC		(107)
	Mitochondrial membrane potential	↓		BEAS-2B	(108)
Immune dysregulation	Klotho	↓	Lung homogenates	HBEC	(112)
	NF-κB	↑	Lung homogenates		(113)
	pro-inflammatory cytokines	↑	Lung homogenates		(113)
Senescence	SA-β-gal	↑	SAEC, PA-SMC, PV-EC & fibroblasts	SAEC (COPD), HBEC, A549, HFL1 & MRC-5	(76,78,79,84-86,115-117)
	p16	↑	Lung tissue sections, AT1, AT2, PA-SMC, PV-EC & fibroblasts	HFL1	(75,76,78,84-86,116,117)
	p21	↑	Lung homogenates, AT2, PA-SMC, PV-EC & peripheral leukocytes	HBEC, A549 & HFL1	(78-80,84-86,117)
	IL-6 & IL-8	↑	Lung tissue sections, AT1, AT2, PA-SMC & PV-EC	MRC-5	(75,76,84,85)
ECM dysregulation	ECM proteins	↑	Lung homogenates		(42)
	Elastogenesis genes	↑	Lung homogenates		(125)
	MMP/TIMP dysregulation	↑	Lung homogenates		(123)
Stem cell exhaustion	Circulating progenitor cells	↓*	Endothelial and haemopoietic progenitor cells		(137,138)
	Regenerative capacity	↓	Basal progenitor cells		(135)
	Stem cell function	↓	HBEC		(142)
	WNT signalling	↓*	Lung homogenates, AT2 & SAEC		(144,147)
	Notch pathway	↓*	SAEC		(143)

CSE: cigarette smoke extract; AT1: type I alveolar cells; AT2: type II alveolar cells; HBEC: human bronchial epithelial cells; PV-EC: pulmonary vascular endothelial cells; HFL1: foetal lung fibroblasts; MRC-5: foetal lung fibroblasts; TPP1: telomere protection protein 1; PA-SMC: pulmonary artery smooth muscle cells; SAEC: small airway epithelial cells; HDAC: histone deacetylase; SIRT: sirtuin; BEAS-2B: bronchial epithelial cell line (virus); mTOR: mechanistic target of rapamycin; IGF1: insulin-like growth factor 1; ROS: reactive oxygen species; SA-β-gal: senescence-associated-β-galactosidase; IL: interleukin; A549: alveolar basal epithelial cell line (carcinoma); ECM: extracellular matrix; MMP: matrix metalloproteinase; TIMP: tissue inhibitor of metalloproteinase. #: mean age was significantly different between COPD and control group; for the other studies, information was not available or mean age was not different between groups.

Telomere shortening

Telomere shortening is an important inducer of senescence and a well-known phenomenon in ageing. Reduced telomere length in circulating leukocytes in COPD has been demonstrated in several studies (80-83), while data regarding telomere shortening in structural cells is still scarce.

Reduced telomere length was demonstrated in pulmonary vascular endothelial cells and pulmonary artery smooth muscle cells that were derived from COPD patients when compared to cells derived from smoking controls (84,85). Tsui *et al.* used fluorescent in situ hybridization (FISH) to assess telomere length in alveolar type II and endothelial cells and demonstrated decreased telomere length in COPD patients when compared to non-smoking controls, but not compared to smoking controls (86). A recent study from Ahmad *et al.* assessed telomere length in lung tissue and reported an association with levels of telomere protection protein 1 (TPP1) (78). Both telomere length and TPP1 levels were reduced in lung homogenates from COPD patients compared to non-smoking controls, but not compared to smoking controls. This was further supported by decreased TPP1 levels and telomere length in CSE treated airway epithelial cells and lung fibroblasts (78).

The above findings of Tsui *et al.* and Ahmad *et al.* (78,86) suggest an association with smoking rather than being COPD specific, although given that most COPD-patients are (ex-) smokers, this might be a contributing factor to disease risk and development.

Epigenetic changes

Epigenetic alterations caused by DNA methylation, histone modifications and noncoding RNA's are highly dynamic and influenced by ageing (87). It has even been postulated that the DNA methylation status of particular CpG-sites, also known as the 'epigenetic clock', can be used in an algorithm to predict the biological age (88,89). However, as far as we know, this algorithm has not been applied yet to COPD patients to test if the biological age of COPD patients determined by their methylation status is indeed increased compared to controls as would be expected. The majority of epigenome-wide methylation studies have been performed in whole blood and not much data is available on DNA methylation in whole lung tissue and lung tissue-specific cell types. While it has been widely established that cigarette smoke affects DNA methylation (90) and that COPD is highly associated with cigarette smoke exposure, in a recent systematic review by Machin *et al.*, no consistent differences were found in DNA methylation in peripheral blood in association with COPD or lung function (91). Therefore, the role of DNA methylation in COPD and thereby the role of age-associated differences in DNA methylation in COPD remains unclear.

Histone deacetylase (HDAC) enzymes can reduce the acetylation of histones, leading to enhanced expression of inflammatory genes involved in the disease pathogenesis of COPD. It has been shown that the HDAC activity is reduced in peripheral lung tissue, alveolar macrophages and bronchial biopsies of COPD patients compared to controls and this activity is further associated with the disease severity of COPD in peripheral lung tissue (92). The NAD⁺-dependent Class III protein deacetylases known as the Sirtuin family are frequently described as anti-ageing enzymes (93). The fact that SIRT1 and 6 have been

shown to be decreased in peripheral lung tissue, and SIRT1 also in serum, of COPD patients compared to controls and, suggests age-associated acetylation differences in COPD (94,95). *Baker et al.* postulate that the reduced expression of both of the Sirtuins is regulated by the micro-RNA MiR-34a, a small endogenous non-coding RNA, which appears to be increased in COPD patients compared to controls. While the role of micro-RNAs in COPD has been extensively reviewed (96) the role of micro-RNAs in accelerated lung ageing is not extensively investigated and remains rather elusive (97).

Metabolism

Loss of proteostasis

Ageing cells are less able to maintain the homeostasis of proteins and contain more damaged proteins. In COPD lungs proteostasis of cells is decreased as well, which results in accumulation of damaged proteins. Accumulation of ubiquitinated proteins and the de-ubiquitinating enzyme and aggregation marker, ubiquitin C-terminal hydrolase L1 (UCH-L1), is increased in lung tissue of patients with severe COPD, and these levels negatively correlate with FEV₁ % predicted (98). Furthermore, several autophagy markers are increased in COPD lung tissue including p62, microtubule-associated proteins 1A/1B light chain 3B (LC3-II), autophagy related 4 (Atg4), Atg5-Atg12 and Atg7 (79,99).

Functional studies showed that autophagy activity (LC3-II flux) is increased in bronchial epithelial cells of COPD patients, without further increase upon CSE treatment (100). However, CSE treatment does increase the amount of autophagosomes in airway and bronchial epithelial cells (79,99). Inhibition of autophagy in bronchial epithelial cells results in accumulation of ubiquitinated protein and p62 (100). Again, the anti-ageing molecule SIRT6 may also regulate autophagy, as SIRT6 overexpression and knockdown resulted in an increase and a decrease of autophagosomes respectively (79).

Deregulated nutrient sensing

Nutrient sensing is a cell's ability to recognize and respond to fuel substrates such as glucose and recent findings suggest that nutrient sensing is increased in COPD lungs. Caloric restriction is strongly associated with longevity, and this is possibly mediated via two main pathways involved in nutrient sensing: mechanistic target of rapamycin (mTOR) and insulin like growth factor (IGF1)-signalling (101,102). The activity of mTOR, an important protein kinase in cell metabolism and nutrient sensing, is increased in total lung tissue and leukocytes of COPD patients (103) as well as in CSE-treated bronchial epithelial cells (79). In addition, SIRT6 is considered to attenuate the IGF1-mTOR pathway. The IGF1 pathway is important in cell growth and interacts with mTOR in the regulation of energy metabolism. Here, SIRT6 may play a role, as overexpression and knockdown of *SIRT6* resulted in a decrease in mTOR activity and increase in IGF1 signalling, respectively (79). Of interest, IGF1 protein levels were found to be increased in airway epithelial cells of patients with chronic bronchitis (104).

Mitochondrial dysfunction

With ageing, the function of the mitochondria decreases, which can lead to oxidative stress. Of interest, increased levels of oxidative stress are observed in COPD lungs as well. In whole lung tissue of COPD patients increased oxidative stress was found, as determined by reactive oxygen species (ROS) levels, oxidized-DNA, lipid peroxidation and nitric oxide (NO) levels (77,105,106). Moreover, lipid peroxidation correlated negatively with FEV₁ % predicted (105). In addition, gene expression and protein levels of the anti-oxidant nuclear factor, erythroid 2 like 2 (NRF2) were decreased in total lung tissue and bronchial epithelial cells of COPD patients (107) and *NRF2* expression was positively correlated with airway obstruction (FEV₁/FVC). Mitophagy, the degradation of mitochondria by autophagy, was increased in total lung tissue of COPD patients (108).

Furthermore, CSE treatment of bronchial epithelial cells resulted in increased ROS levels and mitophagy and decreased mitochondrial potential (108,109). Also in fibroblasts CSE treatment resulted in higher ROS levels (76).

Cellular processes

Immunosenescence and inflammaging

As described above, ageing is associated with immunosenescence and inflammaging. These two definitions underlie most of age-associated diseases and are important during COPD development in aged individuals. Several recent studies have investigated how ageing might affect immune dysregulation in COPD. In a study from 2016, John-Schuster *et al.* (110) demonstrated that aged mice exposed to cigarette-smoke are more susceptible to develop emphysema than younger mice. Aged animals had increased lung inflammation, with higher levels of inflammatory cells and mediators associated with lower repair. Two studies, also from 2015, observed that *Klotho*, an anti-ageing protein with anti-inflammatory properties, is reduced in alveolar macrophages (111) and airway epithelial cells of patients with COPD (112). The reduction was associated with high levels of oxidative stress, inflammation and apoptosis (111,112). Furthermore, decreased expression of miR-125a and b levels in COPD have been linked to inflammation and an impaired immune response. MiR-125a reduction resulted in NF- κ B activation with a classical induction of pro-inflammatory cytokines, while in parallel, low levels of miR-125a and b suppress viral clearance (113). These data underscore the potential of targeting inflammation and at the same time increasing resistance to infections in the aged individual with COPD.

Cellular senescence

Cellular senescence is a cell state in which normal cells stop to divide as a mechanism to prevent tumorigenesis and tissue damage. Senescent cells can be cleared by the immune system, however upon ageing the number of senescent cells is accumulating in tissues. In here, these cells can have detrimental effects as they secrete several inflammatory factors and may disturb normal tissue homeostasis and repair due to the loss of their proliferative

capacity and normal physiologic function (114). Evidence is accumulating for increased cellular senescence in COPD lungs.

The percentage of senescent-associated β -galactosidase positive cells was increased in multiple cell types in COPD patients, including airway epithelial cells, smooth muscle cells, endothelial cells and fibroblasts as well as in CSE treated alveolar and bronchial epithelial cells (76,78,79,84,85,115,116). Another senescence marker, the cell cycle inhibitor p16, was found to be increased in total lung tissue, alveolar cells, airway epithelial cells, smooth muscle cells, endothelial cells and fibroblasts of COPD patients (75,76,84-86,116). Similarly, the presence of p21, another cell cycle inhibitor, was increased in total lung tissue, alveolar cells, smooth muscle cells, endothelial cells and leukocytes of COPD patients (79,80,84-86). P21 was also increased in CSE treated bronchial epithelial cells and fibroblasts (78,79,117). Moreover, the percentages of p16 and p21 positive cells were negatively correlated with FEV₁% predicted in alveolar type II and endothelial cells (86). The levels of IL-6 and IL-8, two important cytokines that are secreted by senescent cells as part of the senescence-associated secretory phenotype, were increased in total lung tissue, alveolar cells, smooth muscle cells and endothelial cells of COPD patients as well as in CSE treated fibroblasts (75,76,84,85). Though these cytokines can also be the result of ongoing inflammation in COPD, these observations cannot directly be related to an increase in cellular senescence.

ECM dysregulation

Age-related changes in the lung can also be observed at the extracellular levels. Comparable to the ageing lung (118,119) the extracellular matrix (ECM) is altered in COPD (120). The main alterations in COPD include increases of several ECM proteins such as collagens, fibronectin and laminin (42), changes in the structural organization of collagen with more disorganized collagen fibres (121), and also a reduction in elastic fibres (122). An important contributing factor to these ECM changes is the imbalance between proteases, such as matrix metalloproteinase 12 (MMP12) and neutrophil elastase, and anti-proteases, like α -1 antitrypsin and tissue inhibitor of metalloproteinase (TIMP) 1-4, as reviewed by Navratilova *et al.* (123).

Elastin degradation plays an important role in the pathogenesis of COPD. Elastin fragments alone are known to induce inflammation, leading to destruction of lung tissue (124). It was previously thought that in COPD the lung loses its ability to repair, however, it has become increasingly more evident that there may be aberrant attempts at repair. A number of genes encoding for elastogenesis components, such as fibulin-5 (*FBLN5*), microfibril associated protein 4 (*MFAP4*), latent transforming growth factor binding protein 2 (*LTBP2*) and elastin (*ELN*) itself were identified in a large COPD patient cohort to be higher expressed (125). Whether these components are beneficial or further drive disease pathogenesis remains unclear, as extracellular proteins have the potential to interfere with different cellular pathways (126,127).

Another interesting observation in COPD is the change in lung fibroblast responses *in vitro*. Lung fibroblasts are the main cells involved in ECM homeostasis and repair in the

lung and several studies have shown differences in terms of ECM production when comparing COPD fibroblasts to those derived from non-COPD controls (128-131), suggesting a disturbed or abnormal repair capacity of these cells.

Stem cell exhaustion

Adult lung tissue is thought to reside in a quiescent state. Upon injury, (stem) cells can get activated and are able to proliferate and (trans) differentiate into other cell types, according to their plasticity (132). Indeed, the lung harbours different cell populations including stem cells, responsible for its unique homeostatic capacity to ensure gas exchange (133,134). Airway basal cells represent a well-characterized stem cell population located in the trachea and bronchi. These cells have the ability to self-renew and give rise to secretory, ciliated and neuroendocrine cells (135). In the distal lung, alveolar type II cells (ATII) (136) have been shown to be able to replenish lost ATII and trans differentiate to alveolar type I cells (ATI), thus ensuring proper gas exchange (132,133). It is most likely that other progenitor or stem cell subpopulations exist, which is indicated by several studies in mouse tissue over the past years, however, the existence of these cell in the human tissue and the relevance for tissue injury and potentially impaired repair, remains elusive.

Moreover, COPD has been associated with reduced numbers and dysfunction of circulating progenitor cells (137,138). Cigarette-smoke, a major risk factor for COPD, was shown to reduce the repair potential of endothelial progenitor cells (139), and bone marrow mesenchymal stem cells by interfering with cell homing and proliferation capacities (140). Thus, stem cell exhaustion might contribute to COPD pathogenesis by reducing the endogenous renewal and repair capacity of the lung by local as well as recruited cells. Stem cell niches fail to respond effectively to additional demands for cell-turnover, moreover, deranged metabolic signalling and premature senescence might occur (141). In line with this, reduced regenerative capacity of basal progenitor cells has been reported in COPD (135). In addition, in a different study, an abnormal population of TRP63⁺ KRT5⁺ KRT14⁺ basal cells was identified in regions of hyperplasia from sections of COPD human airways (142), suggesting abnormal stem cell function. Developmental pathways, such as WNT, Sonic Hedgehog and Notch, are important susceptibility factors for COPD (143-145) and are associated with the regulation of different stem cell functions (146). Canonical WNT signalling, which relies on stabilization of β -catenin for transcriptional activation, is decreased in COPD (144,147,148). Notably, pharmaceutical activation of the pathway led to an increase in surfactant protein C production and secretion along with increased alveolar type I cell marker expression in COPD lung tissue *ex vivo*, thus suggesting that the initiation of stem cell mediated repair in the COPD lung is possible (149).

Evidence for ageing hallmarks and abnormal tissue repair in COPD

Increased levels of DNA damage and decreased levels of DNA repair markers have been demonstrated in COPD, in particular in the alveolar compartment. Although the data is derived from a limited number of studies, it does indicate a role for these ageing markers

in COPD, in particular in relation to emphysema development with increased alveolar wall destruction and lack of repair.

Another key hallmark in ageing is telomere shortening. Since information on telomere length in structural cells is mostly lacking it is difficult to speculate on a role of telomere shortening in relation to tissue repair and remodelling in COPD. Reduced TPP1 levels in relation to smoking in lung tissue and structural cells however does suggest an effect of smoking. Whether this also relates to smoking-induced COPD remains to be elucidated.

Regarding epigenetic changes in COPD, solid data on DNA methylation is lacking, and thus it is yet not possible to infer a role for DNA methylation in abnormal tissue repair in COPD. Several studies however have indicated involvement of histone modification (HDAC and Sirtuins) and miRNAs in COPD and it is of great interest to further evaluate if and how these changes contribute to accelerated lung ageing and abnormal tissue repair in COPD.

With respect to the metabolic changes in COPD, increased autophagy and accumulation of damaged proteins reflects ongoing tissue damage and high protein turnover in COPD. Whether this is cause or consequence (or both) of the abnormal repair response is currently unclear. Similarly, disturbed nutrient sensing (IGF-1-mTOR) and the oxidant anti-oxidant imbalance, indicate that, as with normal ageing, cell homeostasis is disturbed, which makes the cells vulnerable to disease. However, whether and how this contributes to abnormal repair should be evaluated by further studies.

Of all ageing hallmarks in COPD, the changes in cellular processes are probably the best studied. Multiple studies have shown increased cellular senescence and changes in ECM regulation in COPD, in particular in structural cells, including epithelial cells, smooth muscle cells and fibroblasts. The latter are of particular interest, as fibroblasts are the main regulators of tissue repair in the lung and changes in these cells possibly underlie the abnormal tissue repair responses in COPD. Together with the reduced numbers, dysfunction and regenerative capacity of progenitor cells in COPD these age-related cellular changes may very well explain the disturbed repair and remodelling capacity of COPD lungs, both in the alveolar and airway compartment.

Implications for treatment

COPD exacerbations are of major concerns in elderly, as they are highly susceptible to infections. Due to a dampened immune system, vaccination is not considered a successful preventive measure (150). Having this in mind, strategies which boost the immune system have been proposed for lung disease treatment. One of the strategies is the interference with gut microbiota (151). Local microbiota influences immunity at distal sites and organs. *Bifidobacterium breve* and *Lactobacillus rhamnosus* have been shown to reduce inflammatory responses in macrophages that were exposed to cigarette smoke extract *in vitro* (152). Another potential future therapy is based on the application and usage of stem cells. In 2013, bone marrow-derived mesenchymal stem cells (MSCs) were first transplanted to patients with no adverse effects observed in older patients (153). More recently, an immunomodulatory mechanism has been associated with MSCs treatment which

decreased lung inflammation and improved lung function (154). Furthermore, as mentioned above, pharmaceutical activation of the WNT pathway showed promising effects *ex vivo* and indicates the opportunities to induce endogenous stem cell mediated repair in COPD (149).

Although stem cell therapies hold promise as future therapeutic options, more regulations and clinical trials on the matter are needed to optimize therapeutic schemes, dosages, infusion rates and further identify possible risk groups and specific adverse effects (155). Ultimately, understanding the molecular biology of ageing in the lung is crucial for finding new ways of managing COPD in older but also younger (SEO-COPD) patients.

Summary and conclusions

As summarized in this review, main ageing hallmarks are present in COPD and this supports the hypothesis that (abnormal) ageing contributes to COPD development. With respect to the role of abnormal ageing in tissue repair in COPD, the strongest indications come from cellular changes, i.e. increased cellular senescence, ECM dysregulation and stem cell exhaustion. Yet, to be able to answer our question whether accelerated or abnormal ageing is causally contributing to COPD pathogenesis and in particular impaired tissue repair, we need to integrate all findings and assess how age-related changes affect ECM homeostasis and tissue repair in the lung. Ideally this should not be restricted to single-cell culture models with primary lung cells, but also involve more complex co-culture and organoid models, lung tissue slices and/or lab-on-a-chip approaches. An important aspect that needs to be taken into account is the age-matching between the control and COPD groups. Indeed for some studies discussed in this review, the mean age was significantly different between the control and COPD group (indicated in Table 1). This may come as a challenge to distinguish the effects which are related to age and which are related to COPD. Finally, translation and comparison to *in vivo* models and to what happens in the lungs of the actual COPD patients is important for the identification of potential new therapeutic approaches. Evaluation in well-defined clinical samples is crucial to understand the clinical implications and potential benefit for COPD patients. This information may also guide us towards novel approaches aiming to stop or at least slow down accelerated lung ageing in COPD. Providing a future perspective for the most vulnerable group of COPD patients that suffers from the highest disease burden and lacks adequate treatment; severe early onset COPD.

REFERENCES

- (1) Ito K, Barnes PJ. COPD as a disease of accelerated lung aging. *Chest* 2009 01;135(1931-3543; 1):173-180.
- (2) WHO. WHO factsheet Noncommunicable diseases. 2017; Available at: <http://www.who.int/mediacentre/factsheets/fs355/en/>, 2017.
- (3) Mercado N, Ito K, Barnes PJ. Accelerated ageing of the lung in COPD: new concepts. *Thorax* 2015 May;70(5):482-489.
- (4) Rojas M, Meiners S, Le Saux CJ. *Molecular Aspects of Aging: Understanding Lung Aging*. : Wiley-Blackwell; 2014.
- (5) Miller MR. Structural and physiological age-associated changes in aging lungs. *Semin Respir Crit Care Med* 2010 Oct;31(5):521-527.
- (6) Thurlbeck WM, Angus GE. Growth and aging of the normal human lung. *Chest* 1975 Feb;67(2 Suppl):3S-6S.
- (7) Lamb D, Gillyool M, Farrow AS. Microscopic emphysema and its variations with age, smoking, and site within the lungs. *Ann N Y Acad Sci* 1991;624:339-340.
- (8) Gillyool M, Lamb D. Airspace size in lungs of lifelong non-smokers: effect of age and sex. *Thorax* 1993 Jan;48(1):39-43.
- (9) Janssens JP, Pache JC, Nicod LP. Physiological changes in respiratory function associated with ageing. *Eur Respir J* 1999 Jan;13(1):197-205.
- (10) MacNee W. Is Chronic Obstructive Pulmonary Disease an Accelerated Aging Disease? *Ann Am Thorac Soc* 2016 Dec;13(Supplement_5):S429-S437.
- (11) Lopez-Otin C, Blasco MA, Partridge L, Serrano M, Kroemer G. The hallmarks of aging. *Cell* 2013 Jun 6;153(6):1194-1217.
- (12) Franceschi C, Bonafe M, Valensin S, Olivieri F, De Luca M, Ottaviani E, et al. Inflamm-aging. An evolutionary perspective on immunosenescence. *Ann N Y Acad Sci* 2000 Jun;908:244-254.
- (13) Boe DM, Boule LA, Kovacs EJ. Innate immune responses in the ageing lung. *Clin Exp Immunol* 2017 Jan;187(1):16-25.
- (14) De Martinis M, Franceschi C, Monti D, Ginaldi L. Inflamm-aging and lifelong antigenic load as major determinants of ageing rate and longevity. *FEBS Lett* 2005 Apr 11;579(10):2035-2039.
- (15) Lowery EM, Brubaker AL, Kuhlmann E, Kovacs EJ. The aging lung. *Clin Interv Aging* 2013;8(1178-1998; 1176-9092):1489-1496.
- (16) Bailey KL, Bonasera SJ, Wilderdyke M, Hanisch BW, Pavlik JA, DeVasure J, et al. Aging causes a slowing in ciliary beat frequency, mediated by PKCepsilon. *Am J Physiol Lung Cell Mol Physiol* 2014 Mar 15;306(6):L584-9.
- (17) Hogg JC, Timens W. The pathology of chronic obstructive pulmonary disease. *Annu Rev Pathol* 2009;4:435-459.
- (18) Willemsse BW, Ten Hacken NH, Rutgers B, Postma DS, Timens W. Association of current smoking with airway inflammation in chronic obstructive pulmonary disease and asymptomatic smokers. *Respir Res* 2005;6(1465-993):38.
- (19) Hogg JC, Timens W. The pathology of chronic obstructive pulmonary disease. *Annu.Rev.Pathol.* 2009;4(1553-4014):435-459.
- (20) Saetta M, Turato G, Timens W, Jeffery PK. Pathology of chronic obstructive pulmonary disease. *Eur.Respir.Mon.* 2006;11(39):159-176.
- (21) Rutgers SR, Postma DS, Ten Hacken NH, Kauffman HF, van Der Mark TW, Koeter GH, et al. Ongoing airway inflammation in patients with COPD who do not currently smoke. *Thorax* 2000 01;55(0040-6376; 1):12-18.
- (22) Willemsse BW, Ten Hacken NH, Rutgers B, Lesman-Leegte IG, Postma DS, Timens W. Effect of 1-year smoking cessation on airway inflammation in COPD and asymptomatic smokers. *Eur Respir J* 2005 11;26(0903-1936; 5):835-845.
- (23) Lapperre TS, Postma DS, Gosman MM, Snoeck-Stroband JB, Ten Hacken NH, Hiemstra PS, et al. Relation between duration of smoking cessation and bronchial inflammation in COPD. *Thorax* 2006 02;61(0040-6376; 2):115-121.
- (24) Postma DS, Timens W. Remodeling in asthma and chronic obstructive pulmonary disease. *Proc Am Thorac Soc* 2006 07;3(1546-3222; 5):434-439.
- (25) Berg K, Wright JL. The Pathology of Chronic Obstructive Pulmonary Disease: Progress in the 20th and 21st Centuries. *Arch Pathol Lab Med* 2016 Dec;140(12):1423-1428.

- (26) O'Shaughnessy TC, Ansari TW, Barnes NC, Jeffery PK. Inflammation in bronchial biopsies of subjects with chronic bronchitis: inverse relationship of CD8+ T lymphocytes with FEV1. *Am J Respir Crit Care Med* 1997 03;155(1073-449; 1073-449; 3):852-857.
- (27) Saetta M, Di SA, Turato G, Facchini FM, Corbino L, Mapp CE, et al. CD8+ T-lymphocytes in peripheral airways of smokers with chronic obstructive pulmonary disease. *Am J Respir Crit Care Med* 1998 03;157(1073-449; 1073-449; 3):822-826.
- (28) Zhu J, Qiu YS, Majumdar S, Gamble E, Matin D, Turato G, et al. Exacerbations of Bronchitis: bronchial eosinophilia and gene expression for interleukin-4, interleukin-5, and eosinophil chemoattractants. *Am J Respir Crit Care Med* 2001 Jul 1;164(1):109-116.
- (29) Gosman MM, Willemse BW, Jansen DF, Lapperre TS, van Schadewijk A, Hiemstra PS, et al. Increased number of B-cells in bronchial biopsies in COPD. *Eur Respir J* 2006 01;27(0903-1936; 1):60-64.
- (30) Hogg JC, Chu F, Utokaparch S, Woods R, Elliott WM, Buzatu L, et al. The nature of small-airway obstruction in chronic obstructive pulmonary disease. *N Engl J Med* 2004 06/24;350(1533-4406; 26):2645-2653.
- (31) van der Strate BW, Postma DS, Brandsma CA, Melgert BN, Luinge MA, Geerlings M, et al. Cigarette smoke-induced emphysema: A role for the B cell? *Am J Respir Crit Care Med* 2006 04/01;173(1073-449; 7):751-758.
- (32) Brusselle GG, Demoor T, Bracke KR, Brandsma CA, Timens W. Lymphoid follicles in (very) severe COPD: beneficial or harmful? *Eur Respir J* 2009 07;34(1399-3003; 1):219-230.
- (33) Brandsma CA, Hylkema MN, van der Strate BW, Slebos DJ, Luinge MA, Geerlings M, et al. Heme oxygenase-1 prevents smoke induced B-cell infiltrates: a role for regulatory T cells? *Respir Res.* 2008;9(1465-993):17.
- (34) Brandsma CA, Kerstjens HA, Geerlings M, Kerkhof M, Hylkema MN, Postma DS, et al. The search for autoantibodies against elastin, collagen and decorin in COPD. *Eur Respir J* 2011 05;37(1399-3003; 0903-1936; 5):1289-1292.
- (35) Brandsma CA, Kerstjens HA, van Geffen WH, Geerlings M, Postma DS, Hylkema MN, et al. Differential switching to IgG and IgA in active smoking COPD patients and healthy controls. *Eur Respir J* 2012 08;40(1399-3003; 0903-1936; 2):313-321.
- (36) Lee SH, Goswami S, Grudo A, Song LZ, Bandi V, Goodnight-White S, et al. Antielastin autoimmunity in tobacco smoking-induced emphysema. *Nat Med* 2007 05;13(1078-8956; 5):567-569.
- (37) Greene CM, Low TB, O'Neill SJ, McElvaney NG. Anti-proline-glycine-proline or antielastin autoantibodies are not evident in chronic inflammatory lung disease. *Am J Respir Crit Care Med* 2010 Jan 1;181(1):31-35.
- (38) Jeffery PK. Comparison of the structural and inflammatory features of COPD and asthma. Giles F. Filley Lecture. *Chest* 2000 05;117(0012-3692; 0012-3692; 5):251S-260S.
- (39) Lapperre TS, Sont JK, van Schadewijk A, Gosman MM, Postma DS, Bajema IM, et al. Smoking cessation and bronchial epithelial remodelling in COPD: a cross-sectional study. *Respir Res* 2007 11/26;8(1465-993; 1):85.
- (40) Saetta M, Turato G, Maestrelli P, Mapp CE, Fabbri LM. Cellular and structural bases of chronic obstructive pulmonary disease. *Am J Respir Crit Care Med* 2001 May;163(6):1304-1309.
- (41) Jeffery PK. Morphology of the airway wall in asthma and in chronic obstructive pulmonary disease. *Am Rev Respir Dis* 1991 May;143(5 Pt 1):1152-8; discussion 1161.
- (42) Kranenburg AR, Willems-Widyastuti A, Moori WJ, Sterk PJ, Alagappan VK, de Boer WI, et al. Enhanced bronchial expression of extracellular matrix proteins in chronic obstructive pulmonary disease. *Am J Clin Pathol* 2006 Nov;126(5):725-735.
- (43) Liesker JJ, Hacken NH, Zeinstra-Smith M, Rutgers SR, Postma DS, Timens W. Reticular basement membrane in asthma and COPD: Similar thickness, yet different composition. *Int.J.Chron.Obstruct.Pulmon.Dis.* 2009;4(1176-9106; 1):127-135.
- (44) Kunz LI, Strebis J, Budulac SE, Lapperre TS, Sterk PJ, Postma DS, et al. Inhaled steroids modulate extracellular matrix composition in bronchial biopsies of COPD patients: a randomized, controlled trial. *Plos One* 2013;8(1932-6203; 1932-6203; 5):e63430.
- (45) van Straaten JF, Coers W, Noordhoek JA, Huitema S, Flipsen JT, Kauffman HF, et al. Proteoglycan changes in the extracellular matrix of lung tissue from patients with pulmonary emphysema. *Mod Pathol* 1999 07;12(0893-3952; 7):697-705.
- (46) Zandvoort A, Postma DS, Jonker MR, Noordhoek JA, Vos JT, van der Geld YM, et al. Altered expression of the Smad signalling pathway: implications for COPD pathogenesis. *Eur Respir J* 2006 09;28(0903-1936; 3):533-541.

- (47) De Boer WI, van Schadewijk A, Sont JK, Sharma HS, Stolk J, Hiemstra PS, et al. Transforming growth factor beta1 and recruitment of macrophages and mast cells in airways in chronic obstructive pulmonary disease. *Am J Respir Crit Care Med* 1998 12;158(1073-449; 6):1951-1957.
- (48) van der Geld YM, Van Straaten JFM, Postma DS, Timens W. Role of proteoglycans in development and pathogenesis of emphysema. In: Garg HG, Roughley PJ, Hales CA, editors. *Proteoglycans in Lung Disease* New York: Marcel Dekker, Inc; 2002. p. 241-267.
- (49) Gubbiotti MA, Vallet SD, Ricard-Blum S, Iozzo RV. Decorin interacting network: A comprehensive analysis of decorin-binding partners and their versatile functions. *Matrix Biol* 2016 Sep;55:7-21.
- (50) McDonough JE, Yuan R, Suzuki M, Seyednejad N, Elliott WM, Sanchez PG, et al. Small-airway obstruction and emphysema in chronic obstructive pulmonary disease. *N Engl J Med* 2011 10/27;365(1533-4406; 0028-4793; 17):1567-1575.
- (51) Hogg JC, McDonough JE, Suzuki M. Small airway obstruction in COPD: new insights based on micro-CT imaging and MRI imaging. *Chest* 2013 05;143(1931-3543; 0012-3692; 5):1436-1443.
- (52) Barnes PJ. Chronic obstructive pulmonary disease. *N Engl J Med* 2000 Jul 27;343(4):269-280.
- (53) Saetta M. Airway inflammation in chronic obstructive pulmonary disease. *Am J Respir Crit Care Med* 1999 11;160(1073-449; 1073-449; 5):S17-S20.
- (54) Saetta M, Baraldo S, Corbino L, Turato G, Braccioni F, Rea F, et al. CD8+ve cells in the lungs of smokers with chronic obstructive pulmonary disease. *Am J Respir Crit Care Med* 1999 08;160(1073-449; 1073-449; 2):711-717.
- (55) Eidelman D, Saetta MP, Ghezzi H, Wang NS, Hoidal JR, King M, et al. Cellularity of the alveolar walls in smokers and its relation to alveolar destruction. Functional implications. *Am Rev Respir Dis* 1990 06;141(0003-0805; 0003-0805; 6):1547-1552.
- (56) Demedts IK, Demoor T, Bracke KR, Joos GF, Brusselle GG. Role of apoptosis in the pathogenesis of COPD and pulmonary emphysema. *Respir Res* 2006 Mar 30;7:53.
- (57) Voelkel NF, Cool CD. Pulmonary vascular involvement in chronic obstructive pulmonary disease. *Eur Respir J Suppl* 2003 Nov;46:28s-32s.
- (58) Wright JL, Petty T, Thurlbeck WM. Analysis of the structure of the muscular pulmonary arteries in patients with pulmonary hypertension and COPD: National Institutes of Health nocturnal oxygen therapy trial. *Lung* 1992;170(2):109-124.
- (59) Wright JL, Levy RD, Churg A. Pulmonary hypertension in chronic obstructive pulmonary disease: current theories of pathogenesis and their implications for treatment. *Thorax* 2005 Jul;60(7):605-609.
- (60) Barbera JA, Riverola A, Roca J, Ramirez J, Wagner PD, Ros D, et al. Pulmonary vascular abnormalities and ventilation-perfusion relationships in mild chronic obstructive pulmonary disease. *Am J Respir Crit Care Med* 1994 Feb;149(2 Pt 1):423-429.
- (61) Peinado VI, Pizarro S, Barbera JA. Pulmonary vascular involvement in COPD. *Chest* 2008 Oct;134(4):808-814.
- (62) Budhiraja R, Tuder RM, Hassoun PM. Endothelial dysfunction in pulmonary hypertension. *Circulation* 2004 Jan 20;109(2):159-165.
- (63) Verbeken EK, Cauberghs M, Mertens I, Clement J, Lauweryns JM, Van de Woestijne KP. The senile lung. Comparison with normal and emphysematous lungs. 1. Structural aspects. *Chest* 1992 03;101(0012-3692; 0012-3692; 3):793-799.
- (64) Verbeken EK, Cauberghs M, Mertens I, Clement J, Lauweryns JM, Van de Woestijne KP. The senile lung. Comparison with normal and emphysematous lungs. 2. Functional aspects. *Chest* 1992 03;101(0012-3692; 0012-3692; 3):800-809.
- (65) Kirkwood TB. Evolution of ageing. *Nature* 1977 Nov 24;270(5635):301-304.
- (66) Kirkwood TB. Understanding the odd science of aging. *Cell* 2005 Feb 25;120(4):437-447.
- (67) Hurst JR, Vestbo J, Anzueto A, Locantore N, Mullerova H, Tal-Singer R, et al. Susceptibility to exacerbation in chronic obstructive pulmonary disease. *N Engl J Med* 2010 Sep 16;363(12):1128-1138.
- (68) Miravitlles M, Calle M, Soler-Cataluna JJ. Clinical phenotypes of COPD: identification, definition and implications for guidelines. *Arch Bronconeumol* 2012 Mar;48(3):86-98.
- (69) Silverman EK, Chapman HA, Drazen JM, Weiss ST, Rosner B, Campbell EJ, et al. Genetic epidemiology of severe, early-onset chronic obstructive pulmonary disease. Risk to relatives for airflow obstruction and chronic bronchitis. *Am J Respir Crit Care Med* 1998 Jun;157(6 Pt 1):1770-1778.

- (70) Bafadhel M, McKenna S, Terry S, Mistry V, Reid C, Haldar P, et al. Acute exacerbations of chronic obstructive pulmonary disease: identification of biologic clusters and their biomarkers. *Am J Respir Crit Care Med* 2011 Sep 15;184(6):662-671.
- (71) Santos S, Marin A, Serra-Batlles J, de la Rosa D, Solanes I, Pomares X, et al. Treatment of patients with COPD and recurrent exacerbations: the role of infection and inflammation. *Int J Chron Obstruct Pulmon Dis* 2016 Mar 11;11:515-525.
- (72) Montserrat-Capdevila J, Godoy P, Marsal JR, Barbe F, Galvan L. Risk of exacerbation in chronic obstructive pulmonary disease: a primary care retrospective cohort study. *BMC Fam Pract* 2015 Dec 8;16:173-015-0387-6.
- (73) Hunter LC, Lee RJ, Butcher I, Weir CJ, Fischbacher CM, McAllister D, et al. Patient characteristics associated with risk of first hospital admission and readmission for acute exacerbation of chronic obstructive pulmonary disease (COPD) following primary care COPD diagnosis: a cohort study using linked electronic patient records. *BMJ Open* 2016 Jan 22;6(1):e009121-2015-009121.
- (74) Meiners S, Eickelberg O, Konigshoff M. Hallmarks of the ageing lung. *Eur Respir J* 2015 03;45(1399-3003; 0903-1936; 3):807-827.
- (75) Aoshiba K, Zhou F, Tsuji T, Nagai A. DNA damage as a molecular link in the pathogenesis of COPD in smokers. *Eur Respir J* 2012 Jun;39(6):1368-1376.
- (76) Birch J, Anderson RK, Correia-Melo C, Jurk D, Hewitt G, Marques FM, et al. DNA damage response at telomeres contributes to lung aging and chronic obstructive pulmonary disease. *Am J Physiol Lung Cell Mol Physiol* 2015 Nov 15;309(10):L1124-37.
- (77) Caramori G, Adcock IM, Casolari P, Ito K, Jazrawi E, Tsaprouni L, et al. Unbalanced oxidant-induced DNA damage and repair in COPD: a link towards lung cancer. *Thorax* 2011 Jun;66(6):521-527.
- (78) Ahmad T, Sundar IK, Tormos AM, Lerner CA, Gerloff J, Yao H, et al. Shelterin Telomere Protection Protein 1 Reduction Causes Telomere Attrition and Cellular Senescence via Sirtuin 1 Deacetylase in Chronic Obstructive Pulmonary Disease. *Am J Respir Cell Mol Biol* 2017 Jan;56(1):38-49.
- (79) Takasaka N, Araya J, Hara H, Ito S, Kobayashi K, Kurita Y, et al. Autophagy induction by SIRT6 through attenuation of insulin-like growth factor signaling is involved in the regulation of human bronchial epithelial cell senescence. *J Immunol* 2014 Feb 1;192(3):958-968.
- (80) Rutten EP, Gopal P, Wouters EF, Franssen FM, Hageman GJ, Vanfleteren LE, et al. Various Mechanistic Pathways Representing the Aging Process Are Altered in COPD. *Chest* 2016 Jan;149(1):53-61.
- (81) Savale L, Chaouat A, Bastuji-Garin S, Marcos E, Boyer L, Maitre B, et al. Shortened telomeres in circulating leukocytes of patients with chronic obstructive pulmonary disease. *Am J Respir Crit Care Med* 2009 Apr 1;179(7):566-571.
- (82) Houben JM, Mercken EM, Ketelslegers HB, Bast A, Wouters EF, Hageman GJ, et al. Telomere shortening in chronic obstructive pulmonary disease. *Respir Med* 2009 Feb;103(2):230-236.
- (83) Lee J, Sandford AJ, Connett JE, Yan J, Mui T, Li Y, et al. The relationship between telomere length and mortality in chronic obstructive pulmonary disease (COPD). *PLoS One* 2012;7(4):e35567.
- (84) Amsellem V, Gary-Bobo G, Marcos E, Maitre B, Chaar V, Validire P, et al. Telomere dysfunction causes sustained inflammation in chronic obstructive pulmonary disease. *Am J Respir Crit Care Med* 2011 Dec 15;184(12):1358-1366.
- (85) Nouredine H, Gary-Bobo G, Alifano M, Marcos E, Saker M, Vienney N, et al. Pulmonary artery smooth muscle cell senescence is a pathogenic mechanism for pulmonary hypertension in chronic lung disease. *Circ Res* 2011 Aug 19;109(5):543-553.
- (86) Tsuji T, Aoshiba K, Nagai A. Alveolar cell senescence in patients with pulmonary emphysema. *Am J Respir Crit Care Med* 2006 Oct 15;174(8):886-893.
- (87) Yang IV, Schwartz DA. Epigenetic control of gene expression in the lung. *Am J Respir Crit Care Med* 2011 May 15;183(10):1295-1301.
- (88) Horvath S. DNA methylation age of human tissues and cell types. *Genome Biol* 2013;14(10):R115.
- (89) Hannum G, Guinney J, Zhao L, Zhang L, Hughes G, Sada S, et al. Genome-wide methylation profiles reveal quantitative views of human aging rates. *Mol Cell* 2013 Jan 24;49(2):359-367.
- (90) Gao X, Jia M, Zhang Y, Breitling LP, Brenner H. DNA methylation changes of whole blood cells in response to active smoking exposure in adults: a systematic review of DNA methylation studies. *Clin Epigenetics* 2015 Oct 16;7:113-015-0148-3. eCollection 2015.

- (91) Machin M, Amaral AF, Wielscher M, Rezwan FI, Imboden M, Jarvelin MR, et al. Systematic review of lung function and COPD with peripheral blood DNA methylation in population based studies. *BMC Pulm Med* 2017 Mar 20;17(1):54-017-0397-3.
- (92) Ito K, Ito M, Elliott WM, Cosio B, Caramori G, Kon OM, et al. Decreased histone deacetylase activity in chronic obstructive pulmonary disease. *N Engl J Med* 2005 May 12;352(19):1967-1976.
- (93) Michan S, Sinclair D. Sirtuins in mammals: insights into their biological function. *Biochem J* 2007 May 15;404(1):1-13.
- (94) Baker JR, Vuppusetty C, Colley T, Papaioannou AI, Fenwick P, Donnelly L, et al. Oxidative stress dependent microRNA-34a activation via PI3Kalpha reduces the expression of sirtuin-1 and sirtuin-6 in epithelial cells. *Sci Rep* 2016 Oct 21;6:35871.
- (95) Yanagisawa S, Papaioannou AI, Papaporfyriou A, Baker J, Vuppusetty C, Loukides S, et al. Decreased serum sirtuin-1 in chronic obstructive pulmonary disease. *Chest* 2017 May 12.
- (96) Osei ET, Florez-Sampedro L, Timens W, Postma DS, Heijink IH, Brandsma CA. Unravelling the complexity of COPD by microRNAs: it's a small world after all. *Eur Respir J* 2015 09;46(1399-3003; 0903-1936; 3):807-818.
- (97) Christenson SA, Brandsma CA, Campbell JD, Knight DA, Pechkovsky DV, Hogg JC, et al. MiR-638 regulates gene expression networks associated with emphysematous lung destruction. *Genome Med* 2013 12/31;5(1756-994; 12):114.
- (98) Min T, Bodas M, Mazur S, Vij N. Critical role of proteostasis-imbalance in pathogenesis of COPD and severe emphysema. *J Mol Med (Berl)* 2011 Jun;89(6):577-593.
- (99) Chen ZH, Kim HP, Sciruba FC, Lee SJ, Feghali-Bostwick C, Stolz DB, et al. Egr-1 regulates autophagy in cigarette smoke-induced chronic obstructive pulmonary disease. *PLoS One* 2008 Oct 2;3(10):e3316.
- (100) Fujii S, Hara H, Araya J, Takasaka N, Kojima J, Ito S, et al. Insufficient autophagy promotes bronchial epithelial cell senescence in chronic obstructive pulmonary disease. *Oncoimmunology* 2012 Aug 1;1(5):630-641.
- (101) Mathew R, Pal Bhadra M, Bhadra U. Insulin/insulin-like growth factor-1 signalling (IIS) based regulation of lifespan across species. *Biogerontology* 2017 Feb;18(1):35-53.
- (102) Carmona JJ, Michan S. Biology of Healthy Aging and Longevity. *Rev Invest Clin* 2016 Jan-Feb;68(1):7-16.
- (103) Mitani A, Ito K, Vuppusetty C, Barnes PJ, Mercado N. Restoration of Corticosteroid Sensitivity in Chronic Obstructive Pulmonary Disease by Inhibition of Mammalian Target of Rapamycin. *Am J Respir Crit Care Med* 2016 Jan 15;193(2):143-153.
- (104) Chand HS, Harris JF, Mebratu Y, Chen Y, Wright PS, Randell SH, et al. Intracellular insulin-like growth factor-1 induces Bcl-2 expression in airway epithelial cells. *J Immunol* 2012 May 1;188(9):4581-4589.
- (105) Rahman I, van Schadewijk AA, Crowther AJ, Hiemstra PS, Stolk J, MacNee W, et al. 4-Hydroxy-2-nonenal, a specific lipid peroxidation product, is elevated in lungs of patients with chronic obstructive pulmonary disease. *Am J Respir Crit Care Med* 2002 Aug 15;166(4):490-495.
- (106) Rajendrasozhan S, Yang SR, Kinnula VL, Rahman I. SIRT1, an antiinflammatory and antiaging protein, is decreased in lungs of patients with chronic obstructive pulmonary disease. *Am J Respir Crit Care Med* 2008 Apr 15;177(8):861-870.
- (107) Suzuki M, Betsuyaku T, Ito Y, Nagai K, Nasuhara Y, Kaga K, et al. Down-regulated NF-E2-related factor 2 in pulmonary macrophages of aged smokers and patients with chronic obstructive pulmonary disease. *Am J Respir Cell Mol Biol* 2008 Dec;39(6):673-682.
- (108) Mizumura K, Cloonan SM, Nakahira K, Bhashyam AR, Cervo M, Kitada T, et al. Mitophagy-dependent necroptosis contributes to the pathogenesis of COPD. *J Clin Invest* 2014 Sep;124(9):3987-4003.
- (109) Hara H, Araya J, Ito S, Kobayashi K, Takasaka N, Yoshii Y, et al. Mitochondrial fragmentation in cigarette smoke-induced bronchial epithelial cell senescence. *Am J Physiol Lung Cell Mol Physiol* 2013 Nov 15;305(10):L737-46.
- (110) John-Schuster G, Gunter S, Hager K, Conlon TM, Eickelberg O, Yildirim AO. Inflammaging increases susceptibility to cigarette smoke-induced COPD. *Oncotarget* 2016 May 24;7(21):30068-30083.
- (111) Li L, Wang Y, Gao W, Yuan C, Zhang S, Zhou H, et al. Klotho Reduction in Alveolar Macrophages Contributes to Cigarette Smoke Extract-induced Inflammation in Chronic Obstructive Pulmonary Disease. *J Biol Chem* 2015 Nov 13;290(46):27890-27900.
- (112) Gao W, Yuan C, Zhang J, Li L, Yu L, Wiegman CH, et al. Klotho expression is reduced in COPD airway epithelial cells: effects on inflammation and oxidant injury. *Clin Sci (Lond)* 2015 Dec;129(12):1011-1023.

- (113) Hsu AC, Dua K, Starkey MR, Haw TJ, Nair PM, Nichol K, et al. MicroRNA-125a and -b inhibit A20 and MAVS to promote inflammation and impair antiviral response in COPD. *JCI Insight* 2017 Apr 6;2(7):e90443.
- (114) Burton DG, Krizhanovsky V. Physiological and pathological consequences of cellular senescence. *Cell Mol Life Sci* 2014 Nov;71(22):4373-4386.
- (115) Muller KC, Welker L, Paasch K, Feindt B, Erpenbeck VJ, Hohlfeld JM, et al. Lung fibroblasts from patients with emphysema show markers of senescence in vitro. *Respir Res* 2006 Feb 21;7:32.
- (116) Hashimoto Y, Sugiura H, Togo S, Koarai A, Abe K, Yamada M, et al. 27-Hydroxycholesterol accelerates cellular senescence in human lung resident cells. *Am J Physiol Lung Cell Mol Physiol* 2016 Jun 1;310(11):L1028-41.
- (117) Nyunoya T, Monick MM, Klingelutz A, Yarovinsky TO, Cagley JR, Hunninghake GW. Cigarette smoke induces cellular senescence. *Am J Respir Cell Mol Biol* 2006 Dec;35(6):681-688.
- (118) D'Errico A, Scarani P, Colosimo E, Spina M, Grigioni WF, Mancini AM. Changes in the alveolar connective tissue of the ageing lung. An immunohistochemical study. *Virchows Arch A Pathol Anat Histopathol* 1989;415(2):137-144.
- (119) Frette C, Jacob MP, Wei SM, Bertrand JP, Laurent P, Kauffmann F, et al. Relationship of serum elastin peptide level to single breath transfer factor for carbon monoxide in French coal miners. *Thorax* 1997 Dec;52(12):1045-1050.
- (120) Jones RL, Noble PB, Elliot JG, James AL. Airway remodelling in COPD: It's not asthma! *Respirology* 2016 Nov;21(8):1347-1356.
- (121) Tjin G, Xu P, Kable SH, Kable EP, Burgess JK. Quantification of collagen I in airway tissues using second harmonic generation. *J Biomed Opt* 2014 Mar;19(3):36005.
- (122) Black PN, Ching PS, Beaumont B, Ranasinghe S, Taylor G, Merrilees MJ. Changes in elastic fibres in the small airways and alveoli in COPD. *Eur Respir J* 2008 May;31(5):998-1004.
- (123) Navratilova Z, Kolek V, Petrek M. Matrix Metalloproteinases and Their Inhibitors in Chronic Obstructive Pulmonary Disease. *Arch Immunol Ther Exp (Warsz)* 2016 Jun;64(3):177-193.
- (124) Houghton AM, Quintero PA, Perkins DL, Kobayashi DK, Kelley DG, Marconcini LA, et al. Elastin fragments drive disease progression in a murine model of emphysema. *J Clin Invest* 2006 Mar;116(3):753-759.
- (125) Brandsma CA, van den Berge M, Postma DS, Jonker MR, Brouwer S, Pare PD, et al. A large lung gene expression study identifying fibulin-5 as a novel player in tissue repair in COPD. *Thorax* 2015 Jan;70(1):21-32.
- (126) Wagner DE, Bonenfant NR, Parsons CS, Sokocevic D, Brooks EM, Borg ZD, et al. Comparative decellularization and recellularization of normal versus emphysematous human lungs. *Biomaterials* 2014 Mar;35(10):3281-3297.
- (127) Chen X, Song X, Yue W, Chen D, Yu J, Yao Z, et al. Fibulin-5 inhibits Wnt/beta-catenin signaling in lung cancer. *Oncotarget* 2015 Jun 20;6(17):15022-15034.
- (128) Krimmer DI, Burgess JK, Wooi TK, Black JL, Oliver BG. Matrix proteins from smoke-exposed fibroblasts are pro-proliferative. *Am J Respir Cell Mol Biol* 2012 Jan;46(1):34-39.
- (129) Hallgren O, Nihlberg K, Dahlback M, Bjermer L, Eriksson LT, Erjefalt JS, et al. Altered fibroblast proteoglycan production in COPD. *Respir Res* 2010 May 11;11:55-9921-11-55.
- (130) Zandvoort A, Postma DS, Jonker MR, Noordhoek JA, Vos JT, Timens W. Smad gene expression in pulmonary fibroblasts: indications for defective ECM repair in COPD. *Respir Res* 2008 Dec 16;9:83-9921-9-83.
- (131) Larsson-Callerfelt AK, Hallgren O, Andersson-Sjoland A, Thiman L, Bjorklund J, Kron J, et al. Defective alterations in the collagen network to prostacyclin in COPD lung fibroblasts. *Respir Res* 2013 Feb 14;14:21-9921-14-21.
- (132) Crosby LM, Waters CM. Epithelial repair mechanisms in the lung. *Am J Physiol Lung Cell Mol Physiol* 2010 Jun;298(6):L715-31.
- (133) Kotton DN, Morrisey EE. Lung regeneration: mechanisms, applications and emerging stem cell populations. *Nat Med* 2014 Aug;20(8):822-832.
- (134) Miller AJ, Spence JR. In Vitro Models to Study Human Lung Development, Disease and Homeostasis. *Physiology (Bethesda)* 2017 May;32(3):246-260.
- (135) Staudt MR, Buro-Auriemma LJ, Walters MS, Salit J, Vincent T, Shaykhiev R, et al. Airway Basal stem/progenitor cells have diminished capacity to regenerate airway epithelium in chronic obstructive pulmonary disease. *Am J Respir Crit Care Med* 2014 Oct 15;190(8):955-958.
- (136) Barkauskas CE, Cronce MJ, Rackley CR, Bowie EJ, Keene DR, Stripp BR, et al. Type 2 alveolar cells are stem cells in adult lung. *J Clin Invest* 2013 Jul;123(7):3025-3036.

- (137) Fadini GP, Schiavon M, Cantini M, Baesso I, Facco M, Miorin M, et al. Circulating progenitor cells are reduced in patients with severe lung disease. *Stem Cells* 2006 Jul;24(7):1806-1813.
- (138) Palange P, Testa U, Huertas A, Calabro L, Antonucci R, Petrucci E, et al. Circulating haemopoietic and endothelial progenitor cells are decreased in COPD. *Eur Respir J* 2006 Mar;27(3):529-541.
- (139) Paschalaki KE, Starke RD, Hu Y, Mercado N, Margariti A, Gorgoulis VG, et al. Dysfunction of endothelial progenitor cells from smokers and chronic obstructive pulmonary disease patients due to increased DNA damage and senescence. *Stem Cells* 2013 Dec;31(12):2813-2826.
- (140) Tura-Ceide O, Lobo B, Paul T, Puig-Pey R, Coll-Bonfill N, Garcia-Lucio J, et al. Cigarette smoke challenges bone marrow mesenchymal stem cell capacities in guinea pig. *Respir Res* 2017 Mar 23;18(1):50-017-0530-0.
- (141) Boyette LB, Tuan RS. Adult Stem Cells and Diseases of Aging. *J Clin Med* 2014 Jan 21;3(1):88-134.
- (142) Rock JR, Randell SH, Hogan BL. Airway basal stem cells: a perspective on their roles in epithelial homeostasis and remodeling. *Dis Model Mech* 2010 Sep-Oct;3(9-10):545-556.
- (143) Tilley AE, Harvey BG, Heguy A, Hackett NR, Wang R, O'Connor TP, et al. Down-regulation of the notch pathway in human airway epithelium in association with smoking and chronic obstructive pulmonary disease. *Am J Respir Crit Care Med* 2009 Mar 15;179(6):457-466.
- (144) Kneidinger N, Yildirim AO, Callegari J, Takenaka S, Stein MM, Dumitrascu R, et al. Activation of the WNT/beta-catenin pathway attenuates experimental emphysema. *Am J Respir Crit Care Med* 2011 Mar 15;183(6):723-733.
- (145) Van Durme YM, Eijgelsheim M, Joos GF, Hofman A, Uitterlinden AG, Brusselle GG, et al. Hedgehog-interacting protein is a COPD susceptibility gene: the Rotterdam Study. *Eur Respir J* 2010 Jul;36(1):89-95.
- (146) Nusse R. Wnt signaling and stem cell control. *Cell Res* 2008 May;18(5):523-527.
- (147) Wang R, Ahmed J, Wang G, Hassan I, Strulovici-Barel Y, Hackett NR, et al. Down-regulation of the canonical Wnt beta-catenin pathway in the airway epithelium of healthy smokers and smokers with COPD. *PLoS One* 2011 Apr 7;6(4):e14793.
- (148) Jiang Z, Lao T, Qiu W, Polverino F, Gupta K, Guo F, et al. A Chronic Obstructive Pulmonary Disease Susceptibility Gene, FAM13A, Regulates Protein Stability of beta-Catenin. *Am J Respir Crit Care Med* 2016 Jul 15;194(2):185-197.
- (149) Uhl FE, Vierkotten S, Wagner DE, Burgstaller G, Costa R, Koch I, et al. Preclinical validation and imaging of Wnt-induced repair in human 3D lung tissue cultures. *Eur Respir J* 2015 Oct;46(4):1150-1166.
- (150) Webster RG. Immunity to influenza in the elderly. *Vaccine* 2000 Feb 25;18(16):1686-1689.
- (151) Budden KF, Gellatly SL, Wood DL, Cooper MA, Morrison M, Hugenholtz P, et al. Emerging pathogenic links between microbiota and the gut-lung axis. *Nat Rev Microbiol* 2017 Jan;15(1):55-63.
- (152) Mortaz E, Adcock IM, Ricciardolo FL, Varahram M, Jamaati H, Velayati AA, et al. Anti-Inflammatory Effects of *Lactobacillus Rahnosus* and *Bifidobacterium Breve* on Cigarette Smoke Activated Human Macrophages. *PLoS One* 2015 Aug 28;10(8):e0136455.
- (153) Weiss DJ, Casaburi R, Flannery R, LeRoux-Williams M, Tashkin DP. A placebo-controlled, randomized trial of mesenchymal stem cells in COPD. *Chest* 2013 Jun;143(6):1590-1598.
- (154) Gu W, Song L, Li XM, Wang D, Guo XJ, Xu WG. Mesenchymal stem cells alleviate airway inflammation and emphysema in COPD through down-regulation of cyclooxygenase-2 via p38 and ERK MAPK pathways. *Sci Rep* 2015 Mar 4;5:8733.
- (155) Ikonomou L, Freishtat RJ, Wagner DE, Panoskaltis-Mortari A, Weiss DJ. The Global Emergence of Unregulated Stem Cell Treatments for Respiratory Diseases. Professional Societies Need to Act. *Ann Am Thorac Soc* 2016 Aug;13(8):1205-1207.

