

University of Groningen

Characterization of the Gene Cluster Involved in Isoprene Metabolism in *Rhodococcus* sp. Strain AD45

van Hylckama Vlieg, Johan E.T.; Leemhuis, Hans; Lutje Spelberg, Jeffrey H.; Janssen, Dick B.

Published in:
Journal of Bacteriology

DOI:
[10.1128/JB.182.7.1956-1963.2000](https://doi.org/10.1128/JB.182.7.1956-1963.2000)

IMPORTANT NOTE: You are advised to consult the publisher's version (publisher's PDF) if you wish to cite from it. Please check the document version below.

Document Version
Publisher's PDF, also known as Version of record

Publication date:
2000

[Link to publication in University of Groningen/UMCG research database](#)

Citation for published version (APA):

van Hylckama Vlieg, J. E. T., Leemhuis, H., Lutje Spelberg, J. H., & Janssen, D. B. (2000). Characterization of the Gene Cluster Involved in Isoprene Metabolism in *Rhodococcus* sp. Strain AD45. *Journal of Bacteriology*, 182(7), 1956-1963. <https://doi.org/10.1128/JB.182.7.1956-1963.2000>

Copyright

Other than for strictly personal use, it is not permitted to download or to forward/distribute the text or part of it without the consent of the author(s) and/or copyright holder(s), unless the work is under an open content license (like Creative Commons).

The publication may also be distributed here under the terms of Article 25fa of the Dutch Copyright Act, indicated by the "Taverne" license. More information can be found on the University of Groningen website: <https://www.rug.nl/library/open-access/self-archiving-pure/taverne-amendment>.

Take-down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

Downloaded from the University of Groningen/UMCG research database (Pure): <http://www.rug.nl/research/portal>. For technical reasons the number of authors shown on this cover page is limited to 10 maximum.

Characterization of the Gene Cluster Involved in Isoprene Metabolism in *Rhodococcus* sp. Strain AD45

JOHAN E. T. VAN HYLCKAMA Vlieg, HANS LEEMHUIS, JEFFREY H. LUTJE SPELBERG,
AND DICK B. JANSSEN*

Department of Biochemistry, Groningen Biomolecular Sciences and Biotechnology Institute,
University of Groningen, NL-9747 AG Groningen, The Netherlands

Received 29 September 1999/Accepted 30 December 1999

The genes involved in isoprene (2-methyl-1,3-butadiene) utilization in *Rhodococcus* sp. strain AD45 were cloned and characterized. Sequence analysis of an 8.5-kb DNA fragment showed the presence of 10 genes of which 2 encoded enzymes which were previously found to be involved in isoprene degradation: a glutathione *S*-transferase with activity towards 1,2-epoxy-2-methyl-3-butene (*isoI*) and a 1-hydroxy-2-glutathionyl-2-methyl-3-butene dehydrogenase (*isoH*). Furthermore, a gene encoding a second glutathione *S*-transferase was identified (*isoJ*). The *isoJ* gene was overexpressed in *Escherichia coli* and was found to have activity with 1-chloro-2,4-dinitrobenzene and 3,4-dichloro-1-nitrobenzene but not with 1,2-epoxy-2-methyl-3-butene. Downstream of *isoJ*, six genes (*isoABCDEF*) were found; these genes encoded a putative alkene monooxygenase that showed high similarity to components of the alkene monooxygenase from *Xanthobacter* sp. strain Py2 and other multicomponent monooxygenases. The deduced amino acid sequence encoded by an additional gene (*isoG*) showed significant similarity with that of α -methylacyl-coenzyme A racemase. The results are in agreement with a catabolic route for isoprene involving epoxidation by a monooxygenase, conjugation to glutathione, and oxidation of the hydroxyl group to a carboxylate. Metabolism may proceed by fatty acid oxidation after removal of glutathione by a still-unknown mechanism.

2-Methyl-1,3-butadiene (isoprene) is a volatile compound that is emitted in large quantities from plants, especially under thermal stress conditions. The annual global emission from plants is estimated to be 500 million tons, which is similar to the global emission of methane (35). In addition, isoprene emission from several bacteria has been detected (50). Isoprene plays an important role in atmospheric chemistry since it is involved in the generation of ozone and carbon monoxide (42). Despite this, little work on the microbial degradation of isoprene has been done. Cleveland and Yavitt (8) showed that microorganisms consume isoprene even when it is present at trace level concentrations and that soil microorganisms may provide a significant biological sink for atmospheric isoprene. Information about the physiology of isoprene degradation is scarce, however. van Ginkel et al. (44) have shown that degradation in a *Nocardia* sp. starts with a monooxygenase which oxidizes isoprene to 1,2-epoxy-2-methyl-3-butene and 1,2,3,4-diepoxybutane. Ewers and Knackmuss (13) reported the presence of a glutathione-dependent activity towards 1,2-epoxy-2-methyl-3-butene in cell extracts of an isoprene-utilizing *Rhodococcus* sp., but the enzyme catalyzing this reaction and the products formed were not further characterized. Moreover, no work on the genetics of isoprene metabolism has been published.

In our laboratory, *Rhodococcus* sp. strain AD45 was isolated for its capability to use isoprene as the sole source of carbon and energy. The pathway of isoprene degradation in strain AD45 starts with oxidation by a monooxygenase to yield 1,2-epoxy-2-methyl-3-butene (45). A glutathione *S*-transferase cat-

alyzes the nucleophilic attack of glutathione at the sterically most-hindered carbon atom. The reaction product, 1-hydroxy-2-glutathionyl-2-methyl-3-butene (HGMB), is then oxidized in two consecutive steps to 2-glutathionyl-2-methyl-3-butenic acid (GMBA) by a NAD⁺-dependent dehydrogenase that is highly specific for HGMB. The glutathione *S*-transferase and the dehydrogenase have been purified to homogeneity and characterized (46).

Strain AD45 is also capable of oxidizing chlorinated ethenes to the corresponding chlorinated epoxyethanes (45). These epoxides and their decomposition products cause the toxicity that is associated with the oxidative cometabolic degradation of chlorinated ethenes in other organisms (47). Since this toxicity is the main limiting factor for the application of monooxygenase-expressing organisms for dichloroethene removal, it is desirable to obtain insight in the pathways that lead to biological detoxification of these reactive transformation products. The glutathione *S*-transferase of strain AD45 converts *cis*-1,2-dichloroepoxyethane without the formation of toxic chlorinated reaction products and thus may effectively detoxify chlorinated ethene epoxides (45).

Currently, no information on the genetics of isoprene metabolism is available, and the metabolic pathway is still not completely known. In the present study we report the complete nucleotide sequence and polypeptide analysis of an 8,456-bp region encoding the putative isoprene monooxygenase, the glutathione *S*-transferase with activity towards epoxides, a second glutathione *S*-transferase, the HGMB-specific dehydrogenase, and a putative racemase. Furthermore, we have overexpressed both glutathione *S*-transferases, and data on their activities with 1,2-epoxy-2-methyl-3-butene, 1,2-dichloro-4-nitrobenzene (DCNB), and 1-chloro-2,4-dinitrobenzene (CDNB) are reported. The implications for the catabolic pathway of isoprene will be discussed.

* Corresponding author. Mailing address: Department of Biochemistry, Groningen Biomolecular Sciences and Technology Institute, University of Groningen, Nijenborgh 4, NL-9747 AG Groningen, The Netherlands. Phone: 31-50-3634209. Fax: 31-50-3634165. E-mail: d.b.janssen@chem.rug.nl.

MATERIALS AND METHODS

Bacterial strains and media. *Rhodococcus* sp. strain AD45 was grown on isoprene in batch culture or continuous culture using a mineral medium supplemented with 20 mg of yeast extract liter⁻¹ as described before (45) or maintained at 30°C on 0.8% nutrient broth agar plates. For the isolation of genomic DNA, the organism was cultivated in 0.8% nutrient broth medium. *Escherichia coli* strains HB101, JM101, and TOP10F' (Invitrogen, Carlsbad, Calif.), which were used for cloning purposes, were grown at 37°C in Luria-Bertani medium. For plasmid selection the appropriate antibiotic was added at the following concentrations: 50 µg ml⁻¹ for kanamycin, 50 µg ml⁻¹ for ampicillin, and 12.5 µg ml⁻¹ for tetracycline. *E. coli* BL21(DE3) grown at 17°C was used for high-level expression of both glutathione S-transferases of strain AD45.

DNA preparation and manipulation. Genomic DNA of strain AD45 was isolated from an exponentially growing nutrient broth culture (100 ml) as described before (45).

Small-scale preparation of plasmid DNA (<10 kb) from 1.5-ml cultures of *E. coli* was performed using the High Pure plasmid isolation kit (Boehringer, Mannheim, Germany) according to the recommendations of the manufacturer. Small-scale isolation of DNA of large plasmids (>10 kb) was performed using the alkaline lysis method (31) with two modifications. The lysozyme step was omitted, and ethanol instead of isopropanol was used for DNA precipitation.

Large-scale preparation of plasmid DNA from 100-ml cultures of *E. coli* was performed using the Plasmid Midi kit (Qiagen, Hilden, Germany) according to the recommendations of the manufacturer. DNA of pLAFR3 used for the construction of the gene library was isolated by the alkaline lysis method and purified by ultracentrifugation using a CsCl gradient as described by Sambrook et al. (31).

PCR products were purified using a Qiaquick PCR purification kit (Qiagen). DNA restriction, ligations, and dephosphorylations were performed according to protocols described by Sambrook et al. (31).

CNBr cleavage and N-terminal sequencing. Two milligrams of purified glutathione S-transferase (46) was digested with CNBr as described before (19). Peptides were separated by reversed-phase high-pressure liquid chromatography (19), and the N-terminal amino acid sequence of one of the peptides was determined as described before (46). The following sequence was obtained: RVGLDAFRARILDGFNGQG(HoSer). The carboxy-terminal residue was tentatively identified as a homoserine (HoSer), which is the expected product of methionine after CNBr cleavage.

PCR amplification and overexpression of the *isoI* and *isoJ* genes. Degenerate primers for amplification of fragments of the glutathione S-transferase gene were designed on the basis of peptide sequences of the purified glutathione S-transferase (46). Forward primer PGT1, 5'-AAACCATGGT(T/A/C)ACZGTZTAX-CGZTAXGTZCCZGCGZTGG-3' (X = C or T; Y = G or A; Z = G, A, C, or T), was based on the N-terminal amino acid sequence of the protein (MITVYGYV-PAWGPDISPYVTKVXNYTFTGI). Reverse primer PGT3, 5'-AAACCATGGCATZCCXTGZCCYTYAAZCCYTCZAG(A/G/T)AT-3', was based on the N-terminal amino acid sequence of a peptide isolated after CNBr cleavage (see above). For overexpression of the *isoI* and *isoJ* genes the plasmids pGEFIsoI and pGEFIsoJ were constructed. In these plasmids the *isoI* and *isoJ* genes were cloned in the *NcoI* site behind the T7 promoter in the expression vector pGEF+ (32). The *isoI* gene was amplified with forward primer PFI, 5'-AACATACCATGGTACCGTTTA-3', and reverse primer PRI, 5'-ATGCTCAAGATCCATGGTTTCGGACT-3' (*NcoI* sites are underlined; the start codon is in boldface; substituted nucleotides are in italics). The *isoJ* gene was amplified with forward primer PFJ, 5'-ATCGTTCATGGTTGACTTCTA-3', and reverse primer PRJ, 5'-AAGAATCGGCCATGGCGCTCACTTTTGACA-3'. The *isoABCDEF* operon was amplified with forward primer PFM2, 5'-TGCTATTGACACGGACGATTGGTACGACA-3', and reverse primer PRM2, 5'-GATGGGTCCTTCCGGGATTCGGGGGTAGTTC-3'. The genes were cloned behind the *araBAD* promoter in the expression vector pBAD-TOPO (Invitrogen), and protein expression was induced according to the recommendations of the manufacturer.

PCR amplification was performed as described by Innis and Gelfand (18). Each 50-µl reaction mixture contained 5 µl of 10× DNA polymerase buffer, 50 µM deoxynucleoside triphosphates, 1 to 2 U of GoldStar DNA polymerase (Eurogentec, Seraing, Belgium) or *Pwo* DNA polymerase (Boehringer Mannheim B.V., Almere, The Netherlands), 20 pmol of each primer (synthesized by Eurosequence BV, Groningen, The Netherlands), and 10 to 100 ng of template DNA. Amplification of the ISO1 fragment with PGT1 and PGT3 primers was performed by denaturation at 94°C for 5 min followed by "touchdown" PCR that consisted of 14 cycles of denaturation at 94°C for 1 min, hybridization starting at 60°C for 1 min (2°C decrease every second cycle to a touchdown at 48°C), and elongation at 72°C for 1 min (11). Amplification was completed by 25 additional cycles during which the hybridization temperature was 48°C. Amplification of the complete *isoI* and *isoJ* genes was performed by 30 cycles of denaturation at 94°C for 1 min, hybridization at 55°C for 1 min, and elongation at 72°C for 1 min.

A partial purification of IsoJ from *E. coli* BL21(DE3) was carried out by anion exchange chromatography as described by van Hylckama Vlieg et al. (46).

Preparation of the gene library. Sixty micrograms of genomic DNA of strain AD45 was partially digested with *Sau3A* to yield an average fragment size of approximately 23 kb. The digest was dephosphorylated and ligated into the *Bam*HI site of the pLAFR3 cosmid cloning vector (39). The ligation mixture was

packaged with the DNA packaging kit (Boehringer, Mannheim, Germany) and transduced to *E. coli* HB101 according to the recommendations of the manufacturer.

Colonies of the gene library were tested for monooxygenase activity by the indole assay as described by O'Connor et al. (27) or by spraying colonies with 10 mM indole in diethylether. As a control, isoprene-grown colonies of *Rhodococcus* sp. strain AD45 were used.

DNA-DNA hybridization experiments. For Southern blotting experiments, DNA was digested with the appropriate restriction enzyme and separated by agarose gel electrophoresis. The gel was incubated in 0.25 M HCl to nick the DNA before it was transferred to a nylon membrane (Boehringer) by the capillary flow method (31). The DNA was fixed to the membrane by illumination with UV radiation for 3 min.

For colony blotting, colonies grown overnight were transferred to a nitrocellulose membrane (Schleicher and Schuell, Dassel, Germany). After the membrane was dried in air, it was incubated for 5 min in four consecutive steps on Whatman no. 1 filters that were saturated with 10% sodium dodecyl sulfate (SDS), 0.5 N NaOH with 1.5 M NaCl, 1 M Tris-HCl with 1.5 M NaCl (pH 7.5), or 2× SSC (1× SSC is 0.15 M NaCl plus 0.015 M sodium citrate). The DNA was fixed to the membrane by incubation at 80°C for 30 min.

DNA probes were prepared by incorporation of digoxigenin-dUTP with the DIG DNA-labeling kit (Boehringer). The membrane carrying the DNA was prehybridized by incubation at 68°C for 1 h with 5× SSC containing 0.1% *N*-lauroylsarcosine, 0.02% SDS, and 1% blocking agent (Boehringer). Hybridization was performed by overnight incubation at 68°C in this buffer to which 20 ng of denatured probe ml⁻¹ was added, and subsequently the membrane was washed twice in 0.1× SSC containing 0.1% SDS at 68°C. Detection of the hybridized probe was done with anti-digoxigenin-alkaline phosphatase Fab fragments (Boehringer) following the manufacturer's recommendations.

DNA sequencing and analysis. Cycle sequencing (26) was performed on double-stranded DNA using the Thermo Sequenase cycle sequencing kit (Amersham BV, Roosendaal, The Netherlands) with 7-deaza-dGTP and 5'Cy5 fluorescent primers. Sequence reactions were run on the ALF-Express automatic sequencing machine (Pharmacia, Uppsala, Sweden) at the BioMedical Technology Centre (Academic Hospital, Groningen, The Netherlands).

The BLAST program (3) was used to search DDBJ, EMBL, and GenBank databases for proteins that had sequence similarity. Multiple sequence alignments were made with ClustalW, version 1.7 (43) using a gap penalty of 6 and a gap opening penalty of 20. Other parameters were default, and, if necessary, minor adjustments were made manually. Pairwise similarities were calculated with the ALIGN program, which uses the BLOSSUM 50 matrix (17).

Enzyme assays. Glutathione S-transferase activity with epoxides was determined with online gas chromatographic monitoring of substrate depletion as described previously (46). Enantiomers of 1,2-epoxy-2-methyl-3-butene were separated with chiral gas chromatography as described by Wistuba et al. (51). Separation of enantiomers of other chiral compounds was carried out as described before (22). Activities towards DCNB and CDNB were assayed at 30°C in a 50 mM Tris-HCl buffer containing 1 mM substrate (16). Activities were determined at pH 7.0 or 8.5 with CDNB and at pH 7.0 with DCNB since non-enzyme-catalyzed reaction rates were too high for an accurate assay of specific activities at pH 8.0. Substrate was added from a 20 mM stock solution in ethanol. The formation of glutathione conjugates was assayed by monitoring the increase of the absorbance at 340 nm in the presence of 2 mM glutathione for DCNB or at 345 nm in the presence of 10 mM glutathione for CDNB. Activities were calculated using extinction coefficients of 8.5 and 9.6 mM⁻¹ cm⁻¹ for DCNB and CDNB conjugates, respectively (16). Activities were corrected for nonenzymatic reaction rates and for background glutathione S-transferase activity of *E. coli* using extracts of *E. coli* BL21(DE3)pGEF+.

Nucleotide sequence accession number. The nucleotide sequence described in this article was deposited at the EMBL, DDBJ, and GenBank databases under accession no. AJ249207.

RESULTS AND DISCUSSION

Cloning of the genes involved in isoprene metabolism. In order to clone a fragment of the glutathione S-transferase of *Rhodococcus* sp. strain AD45 (46) the degenerate primers PGT1 and PGT3 were designed on the basis of the N terminus and an internal peptide obtained after CNBr digestion. With these primers a 471-bp fragment, designated ISO1, was amplified from genomic DNA of strain AD45. The PCR product was cloned and sequenced, and comparison of the deduced peptide sequence with the N-terminal sequence of the protein and with the sequence of the isolated peptide confirmed that the PCR product represented part of the gene encoding IsoI.

A gene library of *Rhodococcus* sp. strain AD45 in the cosmid pLAFR3 was constructed. Approximately 6,000 independent clones were obtained when the library was transduced to *E. coli*

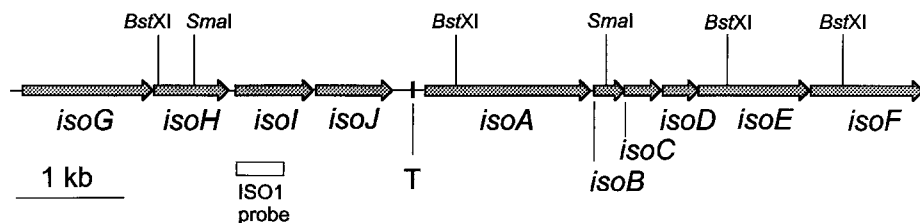


FIG. 1. Schematic representation of the 8,456-bp DNA region of *Rhodococcus* sp. strain AD45 carrying the *isoABCDEFGHIJ* ORFs and a putative *rho*-independent transcription terminator (T).

HB101. An analysis of 28 clones showed that 12 contained plasmids with inserts. The library was screened for the presence of DNA that hybridized with the ISO1 probe by colony blotting. Out of 1,200 insert-containing recombinants tested, three clones that contained plasmids, designated pHL1, pHL8, and pHL9, that hybridized with the ISO1 probe were identified. The clones did not utilize isoprene as a growth substrate. In extracts of cells grown on Luria-Bertani medium no activity towards 1,2-epoxy-2-methyl-3-butene could be detected, indicating that the glutathione *S*-transferase was not functionally expressed. The gene library was also screened for the presence of monooxygenase activity by the indole assay, but no active clones were identified, even when plates were incubated overnight with 0.1 mM isoprene in the gas phase for induction.

Nucleotide sequence analysis. Starting from the ISO1 sequence, an 8,456-bp fragment was sequenced from plasmids pHL8 and pHL9 (Fig. 1). The overall G+C content of the sequenced fragment was 58.9%, which is low compared to the high G+C content typically found for rhodococci, 67 to 72% (14). A sequence similarity search with the BLAST program in various protein and nucleotide databases helped to identify 10 open reading frames (ORFs) that all encoded (putative) proteins that showed similarity with entries in the databases (Fig. 1; Table 1). All ORFs were encoded on the same strand and were preceded by plausible ribosome binding sites. One ORF (*isoH*) starts with a GTG codon, which frequently occurs in *Rhodococcus* mRNA translation (21). A G+C-rich region of almost perfect dyad symmetry (nucleotides 3685 to 3704), followed by an A+T-rich region, is located downstream of *isoJ* (Fig. 1). This structure is characteristic of a *rho*-independent transcription terminator (29).

The ORF that encodes the glutathione *S*-transferase with activity for epoxides was designated *isoI*. The 471-bp ISO1 sequence is identical to that of a fragment of the *isoI* gene, and the molecular mass of the *isoI*-encoded deduced protein (Table 1) is in close agreement with the experimentally determined value of 27 kDa (46). Downstream of the *isoI* gene an ORF designated *isoJ* was found (Table 1).

Upstream of *isoI* an ORF designated *isoH* was found, and it was concluded that this ORF encodes the dehydrogenase with activity for HGMB, since the deduced amino acid sequence of residues 2 to 23 is identical to the N-terminal sequence of the dehydrogenase purified from strain AD45 (46). This indicates that in the mature protein the initiator methionine residue is cleaved off.

Upstream of *isoH* an ORF designated *isoG* was found. A few ATG codons were available as starting points of translation. The start codon that results in the deduced protein that aligns best with similar proteins and that is preceded by a plausible ribosome binding site is included in Table 1.

A gene cluster of six closely spaced ORFs was found downstream of *isoJ*. The ORFs were designated *isoABCDEF* on the

basis of the high sequence similarity with the *tmoABCDEF* gene cluster encoding the toluene 4-monooxygenase of *Pseudomonas mendocina* KR1 and the recently published *aam-ABCDEF* alkene monooxygenase gene cluster of *Xanthobacter* sp. strain Py2 (52–55). The first 7 bp of *isoC* overlapped with the 3' end of *isoB*, and the TGA stop codon of *isoD* overlapped with the presumed ATG start codon of *isoE*.

The flanking regions of the 8,456-bp fragment were further analyzed by single-strand sequencing. Upstream of *isoG*, sequencing a 912-bp fragment revealed the presence of an ORF similar to those encoding γ -glutamylcysteine synthetases, proteins that catalyze the first step in glutathione biosynthesis. Sequence analysis of a 1,892-bp fragment downstream of *iso-ABCDEF* showed the presence of an ORF similar to those encoding aldehyde dehydrogenases and an ORF similar to those encoding glutathione synthetases. Glutathione synthetases couple glycine to γ -glutamylcysteine yielding glutathione. Hence, on a 11,260-bp DNA fragment in strain AD45, the *isoABCDEFGHIJ* genes, a gene encoding a putative aldehyde dehydrogenase, and two genes required for glutathione biosynthesis are present.

Copy number of the *isoI* gene. In order to determine the copy number of *isoI* in *Rhodococcus* sp. strain AD45, total DNA was digested with *Bam*HI, *Bst*XI, *Eco*RI, *Sac*I, *Sal*I, or *Sma*I, separated on an agarose gel, and hybridized with the ISO1 probe. Only one band hybridized with all six restriction enzymes that were tested. For two of the enzymes tested, *Bst*XI and *Sma*I, the sites flank the ISO1 sequence in the 8,456-bp genomic fragment. With these enzymes hybridizing fragments of 3 and 3.5 kb, respectively, were observed, which is in agreement with the sequence data (Fig. 1). The largest hybridizing fragment, a 10-kb fragment, was observed when the DNA was digested with *Sac*I. These data suggest that only one copy of the *isoI* gene is present in *Rhodococcus* sp. strain AD45, although the possibility that there are more copies of a large highly conserved DNA fragment (>10 kb) cannot be excluded.

Similarity of *isoI* and *isoJ* gene products with glutathione *S*-transferases. A sequence similarity search with the *isoI* and *isoJ* gene products showed that both proteins are similar to each other and to other glutathione *S*-transferases (Table 1). The presence of two (putative) glutathione *S*-transferase genes has also been observed in the 2,4,5-trichlorophenoxyacetic acid-degrading *Burkholderia cepacia* strain AC1100 (9) and in *Sphingomonas paucimobilis* strain SYK-6, which grows on lignin (23, 24). Mutagenesis and complementation studies have shown that these genes are not essential for metabolism of 2,4,5-trichlorophenoxyacetic acid, and no physiological function could be assigned to these genes (9).

Similarities of IsoJ to glutathione *S*-transferases in the databases were much higher than similarities of IsoI to glutathione *S*-transferases (Table 1). Proteins IsoI and IsoJ both displayed the highest degree of sequence similarity to other

TABLE 1. Genes and gene products of the *iso* locus and homologies to proteins in the databases

ORF	Position in sequence (nucleotides)	Gene product		Similar gene product	Organism	Identity (%) ^a	Accession no.
		Calculated molecular mass (Da)	Identification				
<i>isoG</i>	57–1274	43,450	Putative racemase	YfdE	<i>Escherichia coli</i>	31.6	AE000325
				Mcr	<i>Mus musculus</i>	30.5	AAB72146
				Fct	<i>Oxalobacter formigenes</i>	26.4	AAC45298
				CaiB	<i>Escherichia coli</i>	24.6	CAA47971
<i>isoH</i>	1291–1971	24,033	NAD-dependent HGMB-dehydrogenase	CsgA	<i>Myxococcus xanthus</i>	33.8	1587000
				XpIII	<i>Xanthobacter</i> sp. strain Py2	24.2	CAA56244
				2bhd	<i>Streptomyces exfoliatus</i>	22.2	1611236A
				XpIV	<i>Xanthobacter</i> sp. strain Py2	22.0	CAA56245
				LigD	<i>Sphingomonas paucimobilis</i> SYK-6	18.6	BAA02030
				Fax ^b	<i>Drosophila melanogaster</i>	27.0	AAB07806
<i>isoI</i>	2038–2754	27,096	Glutathione <i>S</i> -transferase active with epoxides	LigF	<i>Sphingomonas paucimobilis</i> SYK-6	24.3	BAA02031
				IsoJ	<i>Rhodococcus</i> sp. strain AD45	23.8	AJ249207
				ORF3	<i>Burkholderia cepacia</i> AC1100	22.3	AAC43335
				PcpC	<i>Flavobacterium</i> sp. strain ATCC 39723	21.1	Q03520
				GSTB1-1	<i>Proteus mirabilis</i>	20.5	P15214
				ORF4	<i>Burkholderia cepacia</i> AC1100	18.5	AAC43336
				YFCG	<i>Escherichia coli</i>	43.8	P39100
				ORF3	<i>Burkholderia cepacia</i> AC1100	41.9	AAC43335
<i>isoJ</i>	2789–3490	26,342	Glutathione <i>S</i> -transferase active with CDNB and DCNB	LigF	<i>Sphingomonas paucimobilis</i> SYK-6	27.9	BAA02031
				PcpC	<i>Flavobacterium</i> sp. strain ATCC 39723	24.5	Q03520
				GSTB1-1	<i>Proteus mirabilis</i>	24.3	P15214
				IsoI	<i>Rhodococcus</i> sp. strain AD45	23.8	AJ249207
				XamoA	<i>Xanthobacter</i> sp. strain Py2	67.9	AJ012090
				TouA	<i>Pseudomonas stutzeri</i> OX1	49.0	AJ005663
				BmoA	<i>Pseudomonas aeruginosa</i> JI104	48.4	D83068
				TmoA	<i>Pseudomonas mendocina</i> KR1	46.6	M65106
<i>isoA</i>	3796–5340	58,461	Oxygenase α -subunit	TbuA1	<i>Ralstonia pickettii</i> PK01	46.6	U04052
				MemA	<i>Methylococcus capsulatus</i> (Bath)	23.4	P22869
				AmoC	<i>Rhodococcus rhodochrous</i> B-276	23.3	D37875
				XamoB	<i>Xanthobacter</i> sp. strain Py2	55.3	AJ012090
				TouB	<i>Pseudomonas stutzeri</i> OX1	37.9	AJ005663
				TbuU	<i>Ralstonia pickettii</i> PK01	34.7	U04052
				TmoB	<i>Pseudomonas mendocina</i> KR1	32.6	M65106
				BmoB	<i>Pseudomonas aeruginosa</i> JI104	30.2	D83068
<i>isoB</i>	5376–5660	10,331	Oxygenase γ -subunit	BmoC	<i>Pseudomonas aeruginosa</i> JI104	44.4	D83068
				XamoC	<i>Xanthobacter</i> sp. strain Py2	43.4	AJ012090
				TmoC	<i>Pseudomonas mendocina</i> KR1	41.2	M65106
				TbuB	<i>Ralstonia pickettii</i> PK01	40.2	U04052
<i>isoC</i>	5653–5997	12,772	Ferredoxin	XamoD	<i>Xanthobacter</i> sp. strain Py2	51.8	AJ012090
				TbuV	<i>Ralstonia pickettii</i> PK01 ^b	38.4	U04052
				TmoD	<i>Pseudomonas mendocina</i> KR1	35.5	M65106
				TouD	<i>Pseudomonas stutzeri</i> OX1	33.3	AJ005663
<i>isoD</i>	6016–6348	12,380	Effector or coupling protein	XamoE	<i>Xanthobacter</i> sp. strain Py2	52.5	AJ012090
				TbuA2	<i>Ralstonia pickettii</i> PK01	40.2	U04052
				TouE	<i>Pseudomonas stutzeri</i> OX1	38.1	AJ005663
				TmoE	<i>Pseudomonas mendocina</i> KR1	34.5	M65106
<i>isoE</i>	6345–7373	38,510	Oxygenase β -subunit	XamoF	<i>Xanthobacter</i> sp. strain Py2	38.8	AJ012090
				TbuC	<i>Ralstonia pickettii</i> PK01	36.5	U04052
				TouF	<i>Pseudomonas stutzeri</i> OX1	34.4	AJ005663
				AmoD	<i>Rhodococcus rhodochrous</i> B-276	32.7	D37875
				MemC	<i>Methylococcus capsulatus</i> (Bath)	29.9	P22869
				TmoF	<i>Pseudomonas mendocina</i> KR1	29.1	M65106

^a The percentage of similarity was calculated by pairwise alignment using the program ALIGN with default parameterization.

^b Only residues 105 to 340 of the failed axon connection protein were included in calculation of sequence identity.

glutathione *S*-transferases in a region located in the N-terminal domain. This domain is involved in glutathione binding (4, 10, 48). Typically, glutathione *S*-transferases contain one or more tyrosine residues at the N terminus. These residues are also present in IsoI and IsoJ. In some glutathione *S*-transferases the N-terminal tyrosine decreases the pK_a of the glutathione thiol and thus enhances its nucleophilicity (4). In class theta glutathione *S*-transferases, to which many bacterial enzymes belong,

this role is performed by a serine that is located in the N-terminal part of the protein, whereas the tyrosine residues are not essential for catalysis (6, 49). Mutagenesis studies may reveal whether serine residues also play a critical role in glutathione conjugation catalyzed by IsoI or IsoJ.

Two bacterial glutathione *S*-transferases, FosA (accession no. M85195) and FosB (accession no. Q03377), that convert epoxides have been described. The proteins are involved in

bacterial resistance to the antibiotic fosfomycin (1,2-epoxypropylphosphonic acid); they catalyze the conjugation of glutathione to the secondary carbon atom in the epoxide ring (5). Despite the analogy of the reaction catalyzed, IsoI has little sequence similarity to FosA and FosB, which is in agreement with the observation that IsoI does not require metal ions for activity (46).

Sequence similarity of IsoH with SDRs. A sequence similarity search with IsoH, which is the HGMB dehydrogenase, in various protein and DNA databases showed that this protein is similar to proteins of the short-chain dehydrogenase/reductase (SDR) family (20) including the XpIII and XpIV proteins of the epoxide carboxylase system of *Xanthobacter* sp. strain Py2 (2). Multiple sequence alignments with representative and well-studied members (accession no. P25529, 226912, and P00334) of this protein family showed that IsoH contains most of the highly conserved and functionally or structurally important residues (20, 41), which suggests that it has the same topology. The SDR protein family is a large group of enzymes that catalyze the oxidation or reduction of a broad range of substrates including primary alcohols, steroids, and sugars (20). They use NAD(H) or NADP(H) as a coenzyme, do not require metal ions for activity, are approximately 250 residues in length, and are active as monomers, dimers, or tetramers. Several proteins of this family have been subjected to detailed biochemical and structural studies.

The N-terminal parts of SDR proteins typically contain the Rossmann fold for cofactor binding. It is characterized by a GlyXXXGlyXGly motif. In IsoH the first two residues are present as Gly-12 and Gly-15, whereas an alanine residue is located at position 17. SDR proteins contain three highly conserved residues, Ser, Tyr, and Lys, that play a critical role in catalysis (20, 41). These residues are also conserved in IsoH as Ser-132, Tyr-144, and Lys-147. The C-terminal domains of SDR proteins are involved in substrate binding. They are highly variable, which may reflect the adaptation to diverse substrates. Since IsoH catalyzes the oxidation of a glutathione conjugate, it may be expected that substrate binding involves specific interactions with the glutathione moiety of the substrate. Therefore, we looked for similarity with other glutathione binding enzymes, but no glutathione binding site could be identified.

Similarity of isoABCDEF gene products to monooxygenases. The putative isoABCDEF gene products all have a high degree of similarity with peptides of the four-component monooxygenases which are involved in the oxidation of alkenes and aromatic compounds (Table 1). Moreover, the gene orders in the similar clusters are identical. For all gene products except IsoD the highest sequence identity was observed with the alkene monooxygenase of the propene utilizer *Xanthobacter* sp. strain Py2, whose sequence was recently published by Zhou et al. (55). In fact, many of the features of the isoprene monooxygenase gene cluster and the deduced proteins are very similar to those described for the alkene monooxygenase of strain Py2. Lower similarity was observed with polypeptides of three-component monooxygenases, such as methane monooxygenases of *Methylococcus capsulatus* (Bath) (38) and the alkene monooxygenase of *Rhodococcus rhodochrous* (previously *Nocardia corallina*) B-276 (30).

Detailed biochemical and biophysical analysis of toluene 4-monooxygenase of *P. mendocina* KR1 and alkene monooxygenase of *Xanthobacter* sp. strain Py2 (28, 37) has shown that these proteins consist of four dissociable components. This suggests the following model for isoprene monooxygenase of *Rhodococcus* sp. strain AD45. Oxidation of the substrate occurs at a di-iron ($\alpha\beta\gamma$)₂ oxygenase component consisting of the

proteins encoded by isoA, isoB, and isoE. Electrons needed for O₂ activation are provided by an NADH reductase, which is a di-iron disulfur flavoprotein encoded by the isoF gene. A Rieske-type ferredoxin, encoded by isoC, mediates electron transfer from the reductase to the oxygenase. The fourth component is a regulatory or effector protein, encoded by isoD, that may influence reaction rates or product distribution.

The isoprene monooxygenase of *Rhodococcus* sp. strain AD45 is induced in the presence of isoprene and catalyzes the oxidation of aliphatic and aromatic alkenes to the corresponding epoxides. Oxidation of indole by the isoprene monooxygenase in these cells resulted in the rapid formation of indigo. With several alkenes the enzyme produced predominantly one enantiomer of the corresponding epoxides such as (R)-1,2-epoxy-2-methyl-3-butene (enantiomeric excess = 95%), (R)-styrene oxide (87%), and (R)-p-chlorostyrene (59%), indicating that the enzyme may be useful for the biocatalytic production of optically active epoxides. Cells of strain AD45 grown on nutrient broth did not oxidize indole, which suggests that the isoprene monooxygenase is responsible for indigo formation.

The *E. coli* HB101 clones containing plasmid pHL8 or pHL9, both of which carry the isoABCDEF gene cluster, were not active with isoprene or indole (see above). From this we concluded that the isoprene monooxygenase is not functionally expressed in *E. coli*. Similarly, the alkene monooxygenase gene cluster of *Xanthobacter* sp. strain Py2 is also not expressed in *E. coli* (54, 55). When the plasmids pHL8 and pHL9 were transferred by triparental mating to *Xanthobacter*, *Pseudomonas*, or *Burkholderia* spp., the resulting transconjugants were not active with indole or isoprene, indicating that transcription or translation signals were also not recognized in these organisms. To further investigate the possibility of functional expression of the isoprene monooxygenase, the isoABCDEF gene cluster was cloned behind the araBAD promoter in the expression vector pBAD-TOPO. No active protein was produced, probably because one or several of the ribosome binding sites preceding the isoBCDEF, which significantly deviate from the *E. coli* consensus sequence, are poorly recognized in *E. coli* or because of improper protein folding. Complementation of mutants of strain AD45 defective in monooxygenase production may be necessary to obtain final proof that the isoABCDEF gene cluster encodes the isoprene monooxygenase.

Similarity searches with IsoG. Database searches with the isoG-encoded protein showed that it is similar to racemases, carnitine dehydratases (12), and formyl-coenzyme A (CoA) transferases (Table 1). The functionally characterized protein to which IsoG is most similar is α -methylacyl-CoA racemase of *Mus musculus*. This enzyme catalyzes the racemization of an α -methyl group in α -methylacyl-CoA esters that thus become available for enzymes of the β -oxidation pathway, which may only accommodate one enantiomer (33, 34). Furthermore, IsoG is similar to carnitine dehydratase of *E. coli* and the formyl-CoA transferase of *Oxalobacter formigenes* (Table 1). The latter enzyme is involved in the detoxification of oxalic acid, a toxic byproduct of the metabolism of virtually all life forms. It catalyzes the transfer of the CoA moiety from formyl-CoA to oxalic acid, which is subsequently decarboxylated (36).

Overexpression and activity of IsoI and IsoJ. The genes isoI and isoJ were amplified from the cosmid clone pHL8 and placed under the control of a T7 promoter in the expression vector pGEF+ (32). Due to the introduction of the NcoI restriction site, the second residue, which is an isoleucine in both proteins, was replaced by a valine. The resulting plasmids were designated pGEFIsoI and pGEFIsoJ, respectively, and were introduced into *E. coli* BL21(DE3). IsoI and IsoJ were

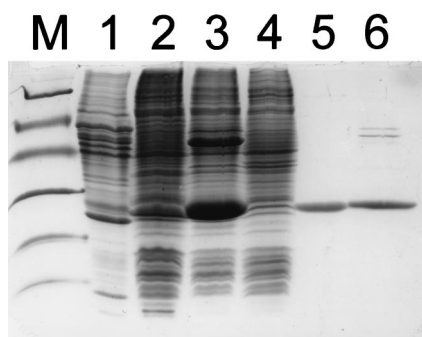


FIG. 2. SDS-PAGE of crude extracts of *E. coli* overexpressing IsoI or IsoJ, purified IsoI, and partially purified IsoJ. Lanes: M, marker proteins; 1, crude extract of *Rhodococcus* sp. strain AD45 grown on isoprene; 2, crude extract of *E. coli* BL21(DE3)(pGEFIsoI); 3, crude extract of *E. coli* BL21(DE3)(pGEFIsoJ); 4, crude extract of *E. coli* BL21(DE3); 5, IsoI purified from strain AD45 (46); 6, partially purified IsoJ from *E. coli* BL21(DE3)(pGEFIsoJ). Marker proteins: phosphorylase b (94 kDa), bovine serum albumin (67 kDa), ovalbumin (43 kDa), carbonic anhydrase (30 kDa), soybean trypsin inhibitor (20 kDa), and α -lactalbumin (14 kDa).

expressed in a soluble and active form at levels of 3 and 35% of the total soluble cellular protein content, respectively (Fig. 2). Microscopic analysis and SDS-polyacrylamide gel electrophoresis (PAGE) analysis of whole-cell lysates showed that inclusion bodies were formed in *E. coli* BL21(DE3)(pGEFIsoI) even when cells were grown at 17°C and when no isopropyl- β -D-thiogalactoside was used to induce expression of the T7 RNA polymerase. The molecular masses of the recombinant proteins were estimated to be 27 and 26 kDa, respectively, as judged by SDS-PAGE (Fig. 2). The extracts with recombinant IsoI and IsoJ were analyzed for activity with 1,2-epoxy-2-methyl-3-butene and with the typical glutathione *S*-transferase substrates CDNB and DCNB.

A glutathione-dependent activity with CDNB of 9.0 nmol min⁻¹ mg⁻¹ was detected at pH 7.0 with recombinant IsoJ. Using DCNB as a substrate, activities were 6.4 and 0.20 nmol min⁻¹ mg⁻¹ at pH 7.0 and 8.5, respectively. IsoI purified from *Rhodococcus* sp. strain AD45 (46) and recombinant IsoI were

not active with CDNB and DCNB. Hence, it was concluded that IsoJ has low but significant activity towards CDNB and DCNB whereas IsoI is not active with these typical glutathione *S*-transferase substrates. Optimal activity was observed at pH 6 to 7 (46), and a specific activity of 26 nmol min⁻¹ mg⁻¹ for CDNB at pH 7.0 was calculated assuming that recombinant IsoJ represented 35% of the total soluble protein (Fig. 2).

Recombinant IsoJ was not active with 1,2-epoxy-2-methyl-3-butene at pH 7.0 or 8.5. The highest specific activity with recombinant IsoI was observed at pH 8.5, and a specific activity of 6 to 10 μ mol min⁻¹ mg⁻¹ at pH 8.5 was calculated assuming that recombinant IsoI represented 3 to 5% of the total soluble protein (Fig. 2). The six- to sevenfold-lower specific activity of recombinant IsoI compared to the activity of IsoI that was purified from strain AD45 (46) may be caused by partial misfolding or the Ile-2-to-Val-2 substitution that was introduced in the cloning procedure.

Pathway of isoprene degradation and comparison with the pathway of propene degradation. The data presented in this study are consistent with a pathway for isoprene degradation that starts with the oxidation of the methyl-substituted double bond by a monooxygenase, which might be encoded by *isoABCDEF* (Fig. 3). This reaction yields 1,2-epoxy-2-methyl-3-butene, which is a substrate for IsoI. This enzyme catalyzes the glutathione-dependent opening of the epoxide ring, yielding the stable glutathione conjugate HGMB. Two consecutive oxidation steps by IsoH in which the hydroxyl is oxidized to a carboxylate then result in the formation of GMBA.

Propene metabolism in *Xanthobacter* sp. strain Py2 and *R. rhodochrous* strain B-276 also starts with oxidation by a monooxygenase yielding epoxypropane. Interestingly, the putative isoprene monooxygenase of *Rhodococcus* sp. strain AD45 is more similar to the alkene monooxygenase of *Xanthobacter* sp. strain Py2 than to the alkene monooxygenase of the phylogenetically more closely related propene utilizer *R. rhodochrous* B-276 (15, 30, 55). The latter is a three-component rather than a four-component monooxygenase. It consists of an ($\alpha\beta$)₂ oxygenase that is encoded by the *amoA* and *amoC* genes, a reductase that directly transfers electrons to the oxygenase that

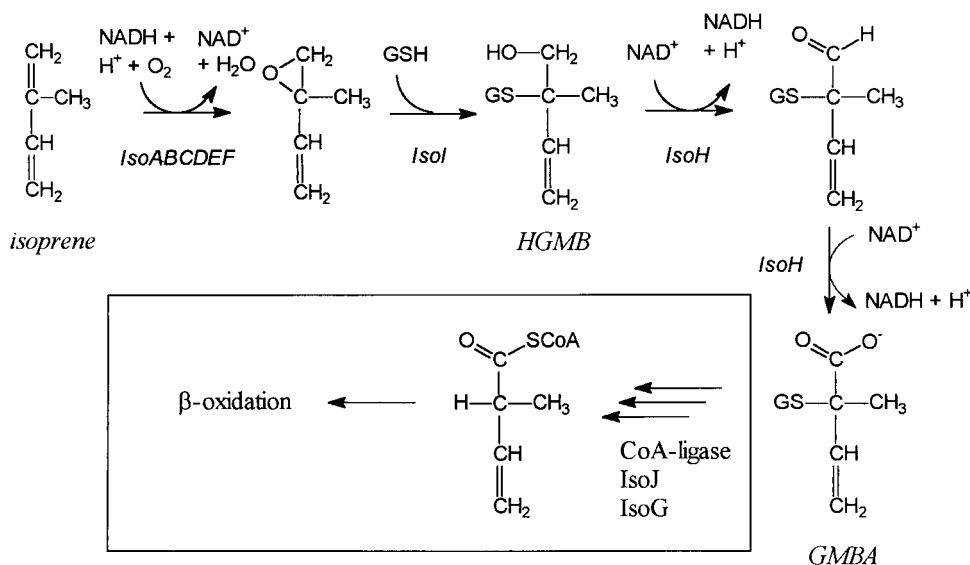


FIG. 3. Pathway of isoprene degradation in *Rhodococcus* sp. strain AD45. Box, proposed pathway for further GMBA metabolism (see text for details).

is encoded by *amoD*, and an effector protein that is encoded by *amoB* (15, 30).

In both propene utilizes the conversion of epoxypropene proceeds with the carboxylation of epoxypropene via a three-step pathway involving four different proteins to yield acetoacetate (1, 2). A detailed study of these proteins for *Xanthobacter* sp. strain Py2 revealed that an epoxyalkane-coenzyme M (CoM) transferase catalyzes the conjugation of CoM to epoxypropene to yield 2-hydroxypropyl-CoM. Hence, 2-hydroxyethers are the primary products of epoxide conversion in isoprene and propene metabolism. However, the enzymes catalyzing the formation of these compounds, XpI encoded by *orf1* in strain Py2 (CAA56241) (1, 2, 40) and IsoI in strain AD45, are not closely related. Furthermore, the next step in both pathways is the dehydrogenation of the 2-hydroxyethers by SDR proteins to the corresponding oxocompounds. Enzymes belonging to the SDR protein family, IsoH in strain AD45 and XpIII and XpIV in strain Py2, catalyze these reactions. Propene metabolism continues with the conversion of 2-ketopropyl-CoM to acetoacetate by an NADPH:2-ketopropyl-CoM oxidoreductase/carboxylase, whereas in isoprene metabolism the aldehyde in 1-oxo-2-glutathionyl-2-methyl-3-butene is oxidized to a carboxylate to yield GMBA.

It remains to be established how in strain AD45 the conversion of GMBA proceeds. Removal of the glutathione moiety is a key step. We did not detect glutathionyl removal, carboxylation, or any other activity with GMBA added to crude extracts of isoprene-grown cells of strain AD45 even when NADH or NADPH was added (46). Both in strains AD45 and Py2 (40) the genes encoding enzymes involved in epoxide conversion are clustered. The *isoH* gene, which encodes the HGMB dehydrogenase, is similar to the genes encoding XpIII and XpIV in *Xanthobacter* sp. strain Py2, whereas IsoI and the two gene products of the *iso* gene cluster, IsoG and IsoJ, for which no role in isoprene degradation has been established, do not have significant homology with genes of propene catabolism in strain Py2. This is in agreement with the fact that epoxide metabolism in strains AD45 and Py2 proceeds via different mechanisms. Interestingly, some glutathione *S*-transferases catalyze the elimination of glutathionyl groups in a reversed Michael reaction (7), but such a reaction may be unlikely with glutathione conjugates in isoprene metabolism since this would require the energetically unfavorable elimination of a proton from the unactivated methyl group. Furthermore, we did not observe conversion of GMBA with extracts of *E. coli* BL21(DE3)(pGEFIsoJ) (unpublished data), although active IsoJ enzyme was produced, as concluded from activity measurements with CDNB or DCNB as the substrate. Reductive removal with a second glutathione molecule, thus generating glutathione disulfide, seems more feasible, but with overproduced IsoJ or with extracts of isoprene-grown cells of strain AD45 no glutathionyl removal from GMBA in the presence of 5 mM glutathione was detected.

An alternative possibility is that IsoJ catalyzes the reductive glutathione removal from a downstream metabolite. This may be the CoA-thioester of GMBA, which could be formed by an unidentified ATP-dependent CoA ligase (Fig. 3). Such a pathway seems feasible since a reductive elimination of glutathione is greatly facilitated when the carboxyl group is replaced by a more-electron-withdrawing CoA-thioester. As described above, IsoG is similar to racemases that catalyze the racemization of an α -methyl group in methylacyl-CoA thioester. Hence, both diastereomers of the GMBA-CoA thioester would become available for further metabolism, which for α -methylacyl-CoA esters mainly occurs through the 2*S*-diastereomer (34). This supports the hypothesis that a CoA-thioester

is an intermediate in the isoprene degradation pathway where IsoG could have a function in racemization. These reactions would result in the formation of 2-methyl-3-butenyl-CoA, which after reduction of the double bond could be further degraded via β -oxidation as in the isoleucine degradation pathway (25).

ACKNOWLEDGMENTS

The work of J.E.T.H.V. was financed by grant IOP91204 from the Dutch IOP Environmental Biotechnology program and grant ENV5-CT95-0086 from the EU Environment and Climate Programme.

Jaap Kingma is acknowledged for performing CNBr digestion and peptide isolation.

REFERENCES

- Allen, J. R., D. D. Clark, J. G. Krum, and S. A. Ensign. 1999. A role for coenzyme M (2-mercaptoethanesulfonic acid) in a bacterial pathway of aliphatic epoxide carboxylation. *Proc. Natl. Acad. Sci. USA* **96**:8432–8447.
- Allen, J. R., and S. A. Ensign. 1999. Two short-chain dehydrogenases confer stereoselectivity for enantiomers of epoxypropene in the multiprotein epoxide carboxylating systems of *Xanthobacter* strain Py2 and *Nocardia corallina* B276. *Biochemistry* **38**:247–256.
- Altschul, S. F., T. L. Madden, A. A. Schäffer, J. Zhang, Z. Zhang, W. Miller, and D. J. Lipman. 1997. Gapped BLAST and PSI-BLAST: a new generation of protein database search programs. *Nucleic Acids Res.* **25**:3389–3402.
- Armstrong, R. N. 1997. Structure, catalytic mechanism, and evolution of the glutathione *S*-transferases. *Chem. Res. Toxicol.* **10**:2–18.
- Bernat, A. B., L. T. Laughlin, and R. N. Armstrong. 1997. Fosfomycin resistance protein (FosA) is a manganese metalloglutathione transferase related to glyoxalase 1 and the extradiol dioxygenases. *Biochemistry* **36**:3050–3055.
- Board, P. G., M. Coggan, M. C. J. Wilce, and M. W. Parker. 1995. Evidence for an essential serine residue in the active site of the theta class glutathione transferases. *Biochem. J.* **311**:247–250.
- Chen, J., and R. N. Armstrong. 1995. Stereoselective catalysis of a retro-Michael reaction by mu class glutathione transferases. Consequences for the internal distribution of products in the active site. *Chem. Res. Toxicol.* **8**:580–585.
- Cleveland, C. C., and J. B. Yavitt. 1998. Microbial consumption of atmospheric isoprene in a temperate forest soil. *Appl. Environ. Microbiol.* **64**:172–177.
- Daubaras, D. L., C. D. Hershberger, K. Kitano, and A. M. Chakrabarty. 1995. Sequence analysis of a gene cluster involved in metabolism of 2,4,5-trichlorophenoxyacetic acid by *Burkholderia cepacia* AC1100. *Appl. Environ. Microbiol.* **61**:1279–1289.
- Dirr, H., P. Reinemer, and R. Huber. 1994. X-ray crystal structures of cytosolic glutathione *S*-transferases. Implications for protein architecture, substrate recognition and catalytic function. *Eur. J. Biochem.* **220**:645–661.
- Don, R. H., P. T. Cox, B. J. Wainwright, K. Baker, and J. S. Mattick. 1991. "Touch-down" PCR to circumvent spurious priming during gene amplification. *Nucleic Acids Res.* **19**:4008.
- Eichler, K., W. H. Schunck, H. P. Kleber, and M. A. Mandrand-Berthelot. 1994. Cloning, nucleotide sequence, and expression of the *Escherichia coli* gene encoding carnitine dehydratase. *J. Bacteriol.* **176**:2970–2975.
- Ewers, J. W. C., and H.-J. Knackmuss. 1991. Biodegradation of chloroethenes using isoprene as a co-substrate, p. 77–83. *In* H. Verachtert and W. Verstraete (ed.), *Proceedings of the International Symposium on Environmental Biotechnology*. Royal Flemish Society of Engineers, Oostende, Belgium.
- Finnerty, W. R. 1992. The biology and genetics of the genus *Rhodococcus*. *Annu. Rev. Microbiol.* **46**:193–218.
- Gallagher, S. C., R. Cammack, and H. Dalton. 1997. Alkene monooxygenase from *Nocardia corallina* B-276 is a member of the class of dinuclear iron proteins capable of stereospecific epoxidation reactions. *Eur. J. Biochem.* **247**:635–641.
- Habig, W. H., M. J. Pabst, and W. B. Jakoby. 1974. Glutathione *S*-transferases. The first enzymatic step in mercapturic acid formation. *J. Biol. Chem.* **249**:7130–7139.
- Henikoff, J. G. 1992. Amino acid substitution matrices from protein blocks. *Proc. Natl. Acad. Sci. USA* **89**:10915–10919.
- Innis, M. A., and D. H. Gelfand. 1990. A guide to methods and amplification, p. 3–11. *In* M. A. Innis, D. H. Gelfand, J. J. Sninsky, and T. J. White (ed.), *PCR protocols*. Academic Press, San Diego, Calif.
- Jacobs, M. H., A. J. van den Wijngaard, M. Pentenga, and D. B. Janssen. 1991. Characterization of the epoxide hydrolase from an epichlorohydrin-degrading *Pseudomonas* sp. *Eur. J. Biochem.* **202**:1217–1222.
- Jörnvall, H., B. Persson, M. Krook, S. Atrian, R. González-Duarte, J. Jeffrey, and D. Ghosh. 1995. Short-chain dehydrogenases/reductases (SDR). *Biochemistry* **34**:6004–6013.

21. Larkin, M. J., R. deMot, L. A. Kulakov, and I. Nagy. 1998. Applied aspects of *Rhodococcus* genetics. *Antonie Leeuwenhoek* **74**:133–153.
22. Lutje Spelberg, J. H., R. Rink, R. Kellogg, and D. B. Janssen. 1998. Enantioselectivity of a recombinant epoxide hydrolase from *Agrobacterium radiobacter*. *Tetrahedron Asymmetry* **9**:459–482.
23. Masai, E., Y. Katayama, S. Kawai, S. Nishikawa, M. Yamasaki, and N. Morohoshi. 1991. Cloning and sequencing of the gene for a *Pseudomonas paucimobilis* enzyme that cleaves β -aryl ether. *J. Bacteriol.* **173**:7950–7955.
24. Masai, E., Y. Katayama, S. Kubota, S. Kawai, M. Yamasaki, and N. Morohoshi. 1993. A bacterial enzyme degrading the model lignin compound β -etherase is a member of the glutathione *S*-transferase superfamily. *FEBS Lett.* **323**:135–140.
25. Massey, L. K., J. R. Sokatch, and R. S. Conrad. 1976. Branched-chain amino acid catabolism in bacteria. *Bacteriol. Rev.* **40**:42–54.
26. Murray, V. 1989. Improved double-stranded DNA sequencing using the linear polymerase chain reaction. *Nucleic Acids Res.* **17**:8889.
27. O'Connor, K. E., A. D. Dobson, and S. Hartmans. 1997. Indigo formation by microorganisms expressing styrene monooxygenase activity. *Appl. Environ. Microbiol.* **63**:4287–4291.
28. Pikus, J. D., J. M. Studs, C. Achim, K. E. Kauffmann, E. Münck, R. J. Steffan, K. McClay, and G. Fox. 1996. Recombinant toluene-4-monooxygenase: catalytic and Mössbauer studies of the purified diiron and Rieske components of a four-protein complex. *Biochemistry* **35**:9106–9119.
29. Platt, T. 1986. Transcription termination and the regulation of gene expression. *Annu. Rev. Biochem.* **55**:339–372.
30. Saeki, H., and K. Furuhashi. 1994. Cloning and characterization of a *No-cardia corallina* B-276 gene cluster encoding alkene monooxygenase. *J. Ferment. Bioeng.* **78**:339–406.
31. Sambrook, J., E. F. Fritsch, and T. Maniatis. 1989. *Molecular cloning: a laboratory manual*, 2nd ed. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y.
32. Schanstra, J. P., R. Rink, F. Pries, and D. B. Janssen. 1993. Construction of an expression and site-directed mutagenesis system of haloalkane dehalogenase in *Escherichia coli*. *Protein Expr. Purif.* **4**:479–489.
33. Schmitz, W., R. Fingerhut, and E. Conzelmann. 1994. Purification and properties of α -methylacyl-CoA racemase from rat liver. *Eur. J. Biochem.* **222**:313–323.
34. Schmitz, W., H. M. Helander, J. K. Hiltunen, and E. Conzelmann. 1997. Molecular cloning of cDNA species for rat and mouse liver-methylacyl-CoA racemases. *Biochem. J.* **326**:883–889.
35. Sharkey, T. D. 1996. Isoprene synthesis by plants and animals. *Endeavour (Oxford)* **20**:74–78.
36. Sidhu, H., S. D. Ogden, H. Y. Lung, B. G. Luttge, A. L. Baetz, and A. B. Peck. 1997. DNA sequencing and expression of the formyl coenzyme A transferase gene, *frc*, from *Oxalobacter formigenes*. *J. Bacteriol.* **179**:3378–3381.
37. Small, F. J., and S. A. Ensign. 1997. Alkene monooxygenase from *Xanthobacter* strain Py2. Purification and characterization of a four-component system central to the bacterial metabolism of aliphatic alkenes. *J. Biol. Chem.* **272**:24913–24920.
38. Stainthorpe, A. C., V. Lees, G. P. Salmond, H. Dalton, and J. C. Murrell. 1990. The methane monooxygenase gene cluster of *Methylococcus capsulatus* (Bath). *Gene* **91**:27–34.
39. Staskawicz, B., D. Dahlbeck, N. Keen, and C. Napoli. 1987. Molecular characterization of cloned avirulence genes from race 0 and race 1 of *Pseudomonas syringae* pv. *glycinea*. *J. Bacteriol.* **169**:5789–5794.
40. Swaving, J., C. A. Weijers, A. J. van Ooyen, and J. A. M. de Bont. 1995. Complementation of *Xanthobacter* Py2 mutants defective in epoxyalkane degradation, and expression and nucleotide sequence of the complementing DNA fragment. *Microbiology* **141**:477–484.
41. Tanaka, N., T. Nonaka, T. Tanabe, T. Yoshimoto, D. Tsuru, and Y. Mitsui. 1996. Crystal structures of the binary and ternary complexes of 7 α -hydroxysteroid dehydrogenase from *Escherichia coli*. *Biochemistry* **35**:7715–7730.
42. Thompson, A. M. 1992. The oxidizing capacity of earth's atmosphere; probable past and future changes. *Science* **256**:1157–1165.
43. Thompson, J. D., D. G. Higgins, and T. J. Gibson. 1994. CLUSTAL W: improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position-specific gap penalties and weight matrix choice. *Nucleic Acids Res.* **22**:4673–4680.
44. van Ginkel, C. G., E. de Jong, J. W. R. Tilanus, and J. A. M. de Bont. 1987. Microbial oxidation of isoprene, a biogenic foliage volatile, and of 1,3-butadiene, an anthropogenic gas. *FEMS Microbiol. Ecol.* **45**:275–279.
45. van Hylckama Vlieg, J. E. T., J. Kingma, A. J. van den Wijngaard, and D. B. Janssen. 1998. A glutathione *S*-transferase with activity towards *cis*-1,2-dichloroepoxyethane is involved in isoprene utilization by *Rhodococcus* sp. strain AD45. *Appl. Environ. Microbiol.* **64**:2800–2805.
46. van Hylckama Vlieg, J. E. T., J. Kingma, W. Kruijzinga, and D. B. Janssen. 1999. Purification of a glutathione *S*-transferase and dehydrogenase involved in isoprene metabolism in *Rhodococcus* sp. strain AD45. *J. Bacteriol.* **181**:2094–2101.
47. van Hylckama Vlieg, J. E. T., W. de Koning, and D. B. Janssen. 1997. Effect of chlorinated ethene conversion on viability and activity of *Methylosinus trichosporium* OB3b. *Appl. Environ. Microbiol.* **63**:4961–4964.
48. Vuilleumier, S. 1997. Bacterial glutathione *S*-transferases: what are they good for? *J. Bacteriol.* **179**:1431–1441.
49. Vuilleumier, S., and T. Leisinger. 1996. Protein engineering studies of dichloromethane dehalogenase/glutathione *S*-transferase from *Methylophilus* sp. strain DM11. Ser12 but not Tyr6 is required for enzyme activity. *Eur. J. Biochem.* **239**:410–417.
50. Wagner, W. P., M. Nemecek, and R. Fall. 1999. Three distinct phases of isoprene formation during growth and sporulation of *Bacillus subtilis*. *J. Bacteriol.* **181**:4700–4703.
51. Wistuba, D., K. Weigand, and H. Peter. 1994. Stereoselectivity of in vitro isoprene metabolism. *Chem. Res. Toxicol.* **7**:336–343.
52. Yen, K. M., M. R. Karl, L. M. Blatt, M. J. Simon, R. B. Winter, P. R. Fausset, H. S. Lu, A. A. Harcourt, and K. K. Chen. 1991. Cloning and characterization of a *Pseudomonas mendocina* KR1 gene cluster encoding toluene-4-monooxygenase. *J. Bacteriol.* **173**:5315–5327.
53. Yen, K. M., and M. R. Karl. 1992. Identification of a new gene, *imoF*, in the *Pseudomonas mendocina* KR1 gene cluster encoding toluene-4-monooxygenase. *J. Bacteriol.* **174**:7253–7261.
54. Zhou, N.-Y., C. K. N. Chan Kwo Chion, and D. J. Leak. 1996. Cloning and expression of the genes encoding the propene monooxygenase from *Xanthobacter*, Py2. *Appl. Microbiol. Biotechnol.* **44**:582–588.
55. Zhou, N.-Y., C. K. N. Chan Kwo Chion, and D. J. Leak. 1999. The alkene monooxygenase from *Xanthobacter* strain Py2 is closely related to aromatic monooxygenases and catalyzes aromatic monohydroxylation of benzene, toluene, and phenol. *Appl. Environ. Microbiol.* **65**:1589–1595.