



## University of Groningen

## Novel views on endotyping asthma, its remission, and COPD

Carpaij, Orestes

DOI:

10.33612/diss.136744640

IMPORTANT NOTE: You are advised to consult the publisher's version (publisher's PDF) if you wish to cite from it. Please check the document version below.

Document Version Publisher's PDF, also known as Version of record

Publication date: 2020

Link to publication in University of Groningen/UMCG research database

Citation for published version (APA):

Carpaij, O. (2020). *Novel views on endotyping asthma, its remission, and COPD*. [Thesis fully internal (DIV), University of Groningen]. University of Groningen. https://doi.org/10.33612/diss.136744640

Copyright

Other than for strictly personal use, it is not permitted to download or to forward/distribute the text or part of it without the consent of the author(s) and/or copyright holder(s), unless the work is under an open content license (like Creative Commons).

The publication may also be distributed here under the terms of Article 25fa of the Dutch Copyright Act, indicated by the "Taverne" license. More information can be found on the University of Groningen website: https://www.rug.nl/library/open-access/self-archiving-pure/taverneamendment.

Take-down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

Downloaded from the University of Groningen/UMCG research database (Pure): http://www.rug.nl/research/portal. For technical reasons the number of authors shown on this cover page is limited to 10 maximum.

Download date: 19-03-2025



# Chapter 1

General introduction



#### 1.1 Asthma

Asthma is the most common inflammatory disease of the lung, affecting around 300 million individuals worldwide, with still increasing prevalence [I]. Due to the high incidence, the chronicity of the condition, and morbidity of the patients, the disease imposes a significant burden on the health care system [I]. Asthma causes symptoms such as wheezing, shortness of breath, chest tightness and cough, that fluctuate in severity over time [I]. These symptoms are associated with the level of airflow obstruction, which in turn is caused by bronchoconstriction (i.e. contraction of smooth muscle bundles around the airways), increased mucus production, and airway wall thickening (due to e.g. influx of inflammatory cells and airway wall remodeling) [I].

The word asthma comes from the Greek  $\alpha\sigma\theta\mu\alpha$ , literally meaning "hard breathing" or "death rattle" [2]. Homer used this term to describe the extreme breathlessness of Hector after being nearly defeated by Ajax, during the battle of Troy [3]. The various connotations of asthma and its symptoms highlight the fact that it is not one, homogenous condition [4]. Indeed, the previously described causes of airflow obstruction also suggest multiple underlying pathophysiological processes [5]. Although widely recognized as a heterogeneous disease, the diagnosis and assessment of asthma are predominantly focussed on the clinical expression of the disease, such as symptoms and loss of pulmonary function. The severity of airflow limitation, as measured by the reduction in forced expiratory volume in one second (FEV<sub>1</sub>) and its variability over time, after salbutamol, or after a bronchoconstrictive agent, provides the most useful information to the physician with respect to diagnosis and treatment options [1,6].

Based on demographics and clinical features, asthmatics can be clustered into "phenotypes". Phenotypes are defined as 'observable properties of an organism produced by the interaction of its genes and the environment' [7–9]. Examples of asthma phenotypes are shown in table I [9].

**Table 1:** asthma phenotypes in relation to clinical and pathological characteristics

Phenotype	Natural history	Clinical and physiological features	Pathobiology and biomarkers
Early-onset allergic	Early onset; mild to severe	Allergic symptoms and other diseases	Eosinophils; specific IgE; TH2 cytokines; thick subepithelial basement membrane
Late-onset	Adult onset; often severe	Sinusitis; less allergic	Corticosteroid-refractory eosinophilia; IL-5
Exercise-induced	-	Mild; intermittent with exercise	Mast-cell activation; TH2 cytokines; cysteinyl leukotrienes
Obesity-related	Adult onset	Women are primarily affected; very symptomatic; airway hyperresponsiveness less clear	Lack of TH2 biomarkers; oxidative stress
Smoking-related	-	Low FEV <sub>1</sub> ; more air trapping	Sputum neutrophilia; TH17 pathways; IL-8

Adapted with (Wenzel SE, et al. Nat Med. 2012 May 4;18(5):716-25)

Decades ago, these phenotypes, which were the only way to describe any disease, largely lacked clinical consequences [4,10]. The introduction of 'endotypes' provides caregivers with more insight into who will and will not respond to therapy [8]. Endotypes are subtypes of a phenotype, defined by a distinct pathogenic mechanism [5,8,10,11]. Approaches to define endotypes in asthma can, among others, be based on features of airway wall remodeling [11–13], eosinophilic inflammation in different lung compartments [11,13], changes in the microbiome [14], volatile compartment composition in exhaled breath [15], gene-expression signatures (e.g. single-cell RNAsequencing) [16,17], proteomics (i.e. set of proteins expressed in a sample), and other -omic modalities [8] (figure 1). The description of asthma endotypes enables us to integrate clinical features, established laboratory parameters, and novel biomarkers, in order to elucidate the multi-layered heterogeneity of asthma. Endotyping started around 1958, when Brown et al. showed that sputum eosinophils predict corticosteroid responsiveness in asthma [18]. Thereafter, different endotypes were determined, such as eosinophilic and non-eosinophilic asthma [5,9,10,20]. Nowadays, distinct pathogenic pathways (examples listed in table 1) are recognised, which were mainly identified in experimental animal models. The attribution of discrete pathogenic mechanisms to specific endotypes of asthma allows tailored interventions, such as anti-IgE [21], antiChapter 1 General introduction

IL-4/-IL-13 [22], anti-IL-13 [23], and anti-IL5 [24]. However, some endotypes, like non-eosinophilic asthma, lack a clear pathogenic mechanism and/or viable therapeutic options. A better understanding of the underlying biological mechanisms driving these endotypes should unveil new treatment targets.

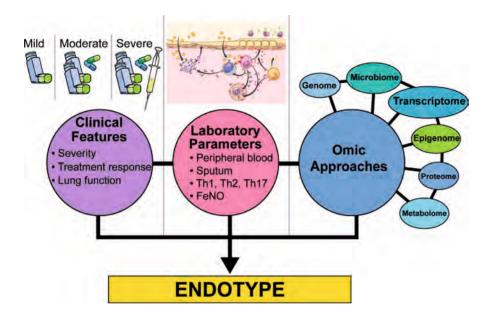


Figure 1: Approaches to endotype asthma.

Reprinted with permission (Tyler SR, et al. J Allergy Clin Immunol. 2019 Jul;144(1):13-23).

Airway inflammation is a prominent feature of asthma. Evidence of elevated type 2-helper T cell (TH2)-associated responses like increased eosinophil numbers in blood, sputum or biopsies or elevated exhaled NO levels, can be found in more than 80% of children and 35 – 50% of adults with asthma [16,25–28]. TH2-driven inflammation is seen in the majority of early-onset allergic, eosinophilic late-onset, and exercise-induced asthmatics [9]. TH2 pathways can be addressed by targeted therapy (e.g. inhaled corticosteroid, anti-IL-5, and anti-IL-13 treatment) and offer opportunities to be linked as a biomarker for therapy response [23,29–31]. One proposed biomarker for TH2 inflammation is periostin, an IL-5 and IL-13 inducible extracellular matrix protein secreted by structural cells of the airways such as basal epithelial cells [17,32]. The role of periostin in asthma and TH2-driven inflammatory responses is an area of active research [33]. The gene encoding periostin is part of a transcriptional TH2 signature in sputum,

that has been successfully used as a biomarker for the response to corticosteroids [16], and has been put forward as a biomarker for anti-IL13 responsiveness in asthmatics [23]. Notwithstanding, currently available literature does not sufficiently support the use of serum periostin levels as a biomarker in clinical practice.

A relatively unexplored, difficult to treat phenotype, is asthma with obesity [34]. Both asthma and obesity are based on observable properties (i.e. phenotypes) and asthma with obesity can thus be defined as a mix of phenotypes, yet the underlying interaction between these diseases is likely to induce a discrete asthma endotype [35,36]. For instance, a previous study found no difference in asthma severity between obese and non-obese asthma patients, but the asthma patients with obesity were characterized by an increased number of neutrophils in the blood and sputum [37]. This is of interest since neutrophilic inflammation has been described as a clinical feature associated with specific asthma endotypes and might reflect a specific pathogenic mechanism (e.g. TH17 inflammation) amendable to tailored therapy [38]. Moreover, while it is known that patients with obesity have a higher risk for developing asthma compared to the general population [36], the risk factors causing this relation might be related to insulin resistance or other components of the metabolic syndrome, rather than to the higher body mass index per se [35,39]. Understanding the unique mechanisms of the asthma-obesity syndrome could potentially reveal new therapeutic options for this phenotype-combination.

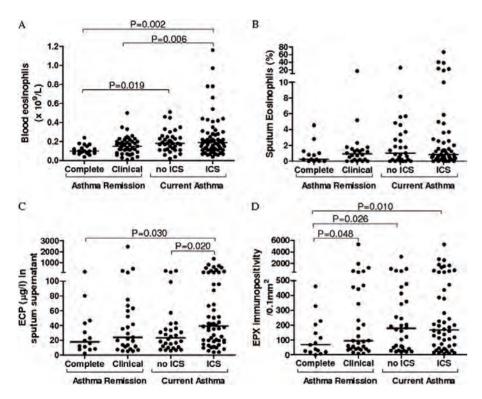
#### 1.2 Asthma remission

Even though asthma is a chronic respiratory disease for which a curative intervention is not available, it has been reported that asthma patients can go into spontaneous remission [40]. Patients in asthma remission are no longer burdened by symptoms, and no longer require any asthma medication. Notwithstanding the lack of symptoms and medication use, patients in asthma remission might still have (asymptomatic) bronchial hyperresponsiveness, low lung function, and ongoing airway inflammation [13]. This situation is referred to as clinical remission. Interestingly, a subset of asthma remission patients has normal lung function and absence of bronchial hyperresponsiveness. This situation is referred to as complete asthma remission [40]. In other words, individuals in complete asthma remission, have been diagnosed with asthma in the past, but

IO II

Chapter I General introduction

would now be categorized as respiratory healthy by clinicians. Figure 2 shows that individuals with clinical asthma remission have similar blood eosinophil levels as patients with asthma without ICS treatment, and comparable eosinophilic peroxidase immunopositivity in bronchial biopsies compared to ICS-using and –naïve asthmatics. Individuals with complete asthma remission had the lowest degree of eosinophilic inflammation. In agreement with this, complete asthma remission might be the disease state closest to cured asthma.



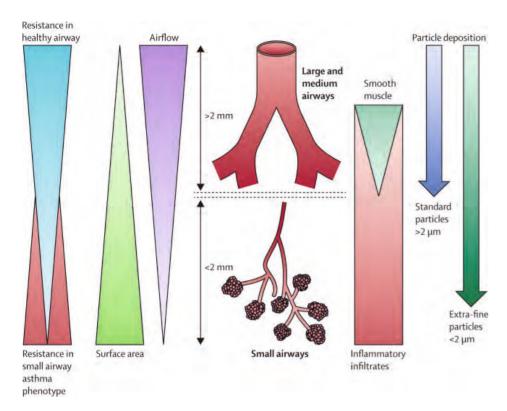
**Figure 2:** eosinophilic inflammation in inhaled corticosteroid (ICS) naïve and ICS-using asthmatics, clinical- and complete asthma remission subjects. **A:** absolute blood eosinophils, **B:** percentage of sputum eosinophils, **C:** eosinophil cationic protein (ECP) in sputum supernatant, **D:** eosinophilic peroxidase (EPX) immunopositivity (pixels) in bronchial biopsies P values of EPX immunopositivity are after correction for age, sex, and smoking using multiple regression analysis. Horizontal bars represent median values. Reprinted with permission (Broekema M et al. Am J Respir Crit Care Med. 2011 Feb 1;183(3):310-6).

To date, the molecular and cellular mechanisms of asthma remission are not known. Understanding its biology is an important research goal as it could potentially reveal novel biological targets that can be addressed therapeutically to induce complete asthma remission and cure the disease. As such, eliciting complete asthma remission in patients with persistent disease would be, together with asthma prevention, the ultimate therapeutic goal. Nevertheless, the vast majority of studies focus on clinical asthma remission. But as shown in figure 2, clinical remission still demonstrates features that are similar to persistent asthma. Therefore, it can be hypothesized that the study of complete asthma remission has a higher chance to reveal biological pathways with the capacity to treat asthma or even induce asthma remission.

## 1.3 Small airways dysfunction

Asthma affects the entire bronchial tree, including the small airways [41,42]. Small airways disease is important since these airways are estimated to comprise 80 - 90% of the total airway surface area [43,44]. The small airways, defined as those with an internal diameter ≤2mm [45], significantly contribute to the airway resistance in patients with obstructive pulmonary disease [41]. Figure 3 illustrates the hypothesis of small airways physiology; in the healthy population, airway resistance gradually decreases from large to small airways due to an exponential increase of the surface area. In conditions with small airways disease such as asthma this is not the case, as inflammatory infiltrates partially block the most distal airways [46,47], causing less peripheral airflow, consequently decreasing gas diffusion in the alveoli [42]. And while the surface of the lumen and airway walls has shrunk, it is postulated that small airways disease reduces both particle deposition and exhalation [42]. Moreover, airway wall remodeling can increase its stiffness, thereby affecting the elasticity and resistance of the small airways [48].

I2



**Figure 3:** small airway asthma phenotype. Reprinted with permission (Lipworth B, et al. Lancet Respir Med 2014;2: 497–506).

There are various tools to assess small airways dysfunction [42,49]; these are presented in table 2. The different physiological tests can identify different aspects of the small airways [41], which are increasingly altered per GINA severity stage [41]. However, there is no gold standard for many of these tests, and some are difficult to perform for the severely obstructed patient. For clinical practice, it is important that these measurements delineate different types of small airways disease, which differ in treatment options (e.g. use of ultra-fine inhaled corticosteroids [50]) or prognosis [51].

Table 2: the assessment of small airways

Tool	Outcome	Measures
Spirometry	Dynamic volumes and flow	FEF <sub>2575%</sub>
Whole-body plethysmography	Lung volumes and hyperinflation/ air trapping and resistance	Raw, functional residual capacity, ratio of residual volume to total lung capacity
Multiple Breath Nitrogen Washout (MBNW)	Airtrapping and ventilation heterogeneity	Functional residual capacity, ratio of closing volume to vital capacity, ratio of residual volume to total lung capacity, $S_{\rm acin}$ , $S_{\rm cond}$
Impulse oscillometry (IOS)	Airway resistance and reactance	R <sub>5</sub> -R <sub>20</sub> , reactance area under curve, reactance at 5 Hz, resonant frequency
Exhaled fractional nitric oxide (FeNO)	Airway inflammation	Alveolar and bronchial nitric oxide fractions
Imaging	Air trapping and regional distribution	High-resolution CT parametric response mapping, gamma scintigraphy, PET, MRI, OCT
Bronchoscopy	Airway resistance and inflammation	Wedged airway resistance, transbronchial biopsy, bronchoalveolar lavage
Induced sputum sample investigation	Airway inflammation	Cell and cytokine profile

 $\mathbf{FEF}_{25,75\%}$ : forced mid-expiratory flow between 25% and 75% offorced vital capacity.  $\mathbf{S}_{acin}$ : acinar (diffusion) dependent ventilation heterogeneity.  $\mathbf{S}_{cond}$ : conductive (convection) dependent ventilation heterogeneity.  $\mathbf{R}_{\mathbf{S}} - \mathbf{R}_{20}$ : peripheral airways resistance as difference between measurements at 5 Hz and 20 Hz.  $\mathbf{Raw}$ : total airway resistance,  $\mathbf{OCT}$ : optical coherence tomography. Reprinted with permission (Lipworth B et al. Lancet Respir Med 2014;2: 497–506).

# 1.4 COPD

Chronic Obstructive Pulmonary Disease (COPD) is the third leading cause of death worldwide, accounting for approximately three million deaths annually (i.e. 6% of all deaths globally), and its prevalence is still rising [52]. COPD is characterized by respiratory symptoms and persistent airflow obstruction (i.e. FEV<sub>1</sub>/FVC ratio <70%) due to airway and alveolar abnormalities (e.g. obstructive bronchiolitis, emphysema, chronic mucus hypersecretion) caused by significant exposure to noxious particles or gases [53]. It is widely accepted that smoking is the main risk factor for COPD, yet only 20 – 30% of smokers will ultimately develop the disease [54]. The classification of disease severity is usually based on exacerbation frequency and the severity of airflow obstruction, but it does not accurately capture the heterogeneity of COPD.

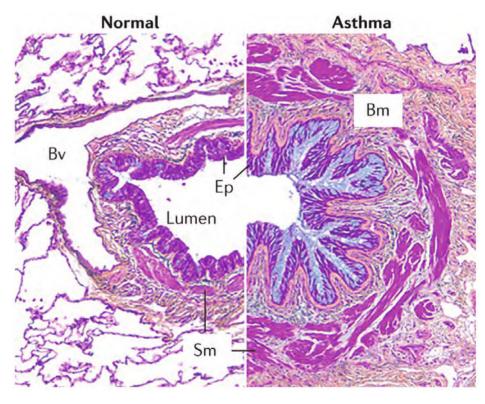
Chapter I General introduction

Further clinical characterization, such as measuring blood and sputum inflammatory cell counts, alpha-I-antitrypsine levels (i.e. genetic predisposition), residual volume, diffusion capacity of carbon monoxide, and bronchial hyperresponsiveness, as well as imaging of the lung using inspiration-expiration CT-scans, is required to define COPD endotypes. The classification of COPD patients into specific endotypes enables clinicians to improve treatment of COPD patients by prescribing a more personalized therapy [53,55]. For instance, COPD patients with elevated sputum eosinophils need to be treated with inhaled corticosteroids [55]. Other treatment options for COPD endotypes – so called 'treatable traits' – are: lung volume reduction therapy for COPD patients with hyperinflation and heterogeneous emphysema [56], phosphodiesterase-4 inhibitors for patients with severe airflow limitation and symptoms of chronic bronchitis [57], and long-term low-dose macrolides for patients with airway bacterial colonisation quantified by sputum culture [55], or in case of frequent exacerbations.

COPD is a chronic inflammatory disease of the respiratory tract and lung parenchyma in response to chronic irritants such as cigarette smoke [53], and is associated with increased numbers of macrophages in the peripheral airways, lung parenchyma and pulmonary vessels, and with increased numbers of activated neutrophils and lymphocytes [53]. In a sub-population of COPD patients, there may also be increased levels of eosinophils [58–60], a TH2 driven inflammatory cell type. The latter inflammatory cell type is of clinical interest, for two reasons. First, eosinophilia in patients with COPD is associated with a higher frequency of exacerbations [61]. Second, numerous studies have shown that blood eosinophil counts predict the magnitude of the effect of inhaled corticosteroids [62-64]. Consequently, thresholds for blood eosinophils (i.e. <100 or >300 cells/µl) have been implemented in the GOLD guidelines for treatment of COPD [53]. Although Casanova et al. found that 43.8% of COPD patients have eosinophilia (i.e. >300 cells/μl) [60], individual predictions of response are still far from ideal, especially for patients with blood eosinophils between 100 and 300 cells/µl. As also mentioned for asthma, periostin has potential as a biomarker for TH2-driven inflammation and ICS responsiveness in asthma. Yet, its applicability as a biomarker for ICS responsiveness in COPD has been investigated to a limited extent only.

# 1.5 Airway wall remodeling

Asthma is characterised by structural changes of the airways, so-called airway wall remodeling [65]. Eventually, this results in airway wall thickening in proportion to disease severity and duration [66,67]. Factors involved in remodeling are: epithelial fragility, subepithelial collagen deposition including basement membrane thickening, smooth muscle hypertrophy, mucus metaplasia, angiogenesis and increased extracellular matrix (e.g. glyco-) proteins [68,69]. Figure 3 shows a cross-section of a normal compared to a remodeled airway wall, the latter from severe asthma. Airway remodeling is linked to clinical features such as lung function impairment, hyperresponsiveness, mucus hypersecretion, and air trapping [70–74].



**Figure 3:** cross section of a severe asthmatic airway (right) compared with a normal airway (left). Asthma involves mucosal inflammation that most frequently consists of activated eosinophils, mast cells and T lymphocytes within the context of a remodelled airway with mucous metaplasia, an increase in smooth muscle (**Sm**), fibrosis and angiogenesis. **Bm:** basement membrane, **Bv:** blood vessel, **Ep:** epithelium. Reprinted with permission (Holgate ST, et al. Nat Rev Dis Primers. 2015 Sep 10;1:15025).

General introduction

Ι

Quantifying parameters associated with airway wall remodeling is of interest for two reasons: first of all, early stages of pulmonary obstructive conditions could be detected by measuring (biomarkers of) sub-symptomatic airway wall remodeling; various studies have found that remodeling of the airways already occurs in early and mild stages of asthma [75,76], also preceding the development of asthma in at-risk children, younger than six years [77]. Second, the parameters of airway wall remodeling can then be correlated with types of inflammatory processes and with inhaled, systemic, or intervention therapy response [78,79]. This would ultimately allow clinicians to use these remodeling features as a biomarker for a certain endotype, providing stratification of treatment options for the patients.

Currently, two diagnostic tools are available: high-resolution CT and assessment of the airway wall by bronchial biopsy [80,81]. Unfortunately, measuring bronchial wall and lumen area using a CT-scan does not provide information on the cause of airway wall thickening. Inspection of the bronchial biopsy is the gold standard to determine airway remodeling, but is a burden to patients and is time-consuming due to processing and staining of the biopsy sections. Additionally, a biopsy is only a small segment, which is routinely generalized to reflect the status of the airway wall remodelling along the small airways. Thus, current tools to detect and quantify airway remodeling are limited, especially in vivo. Optical Coherence Tomography (OCT) is a novel imaging technique that produces infrared-refraction images, both cross-sectional and sagittal (up to 5cm). A few studies already investigated OCT intensity areas in the airway walls between patients with obstructive pulmonary diseases and healthy participants; Ding et al. saw smaller luminal areas and thicker airway wall areas in higher COPD GOLD stages, compared to their never-smoking or smoking control peers [82]. Another study successfully used OCT imaging to determine in vivo elastic airway wall properties in individuals with and without obstructive pulmonary disease [83]. In asthma, the mucosal- and epithelial thickness measured by OCT was higher than in allergic- and non-allergic non-asthmatic controls [84]. However, no literature is available with respect to what component in the airway wall causes the OCT to refract and reflect infrared light, or what asthma phenotypes have other OCT signals. Most importantly, a relationship between OCT imaging and biomarkers for airway wall remodelling has not been described to date. Although OCT tissue intensity has been linked to collagen

deposition in the ovaria and skin [85,86], this has not yet been investigated in the airways.

## 1.6 Outline of the thesis

The overall aim of this thesis was to provide an overview of what is known about the asthma-obesity and asthma remission phenotypes, add knowledge to these topics, and to introduce new methods to:

- Endotype asthma, by single-cell RNA-sequencing.
- Endotype COPD, by measuring serum periostin levels and performing transcriptomic clustering.
- Analyze airway wall remodeling, which occurs in both obstructive pulmonary diseases.

In **Chapter 2**, we provide an overview of the current literature about clinical and complete asthma remission. We discuss the definition, prevalence, clinical characteristics, inflammatory markers, histological signs and genotypes linked to these phenomena. Next to current knowledge, we highlight future studies that will enable further exploration of the pathophysiology of asthma remission.

In 1972-1976, children diagnosed with clinical asthma were extensively characterized and re-examined in young adulthood and late adulthood. In these latter visits, the subjects were divided in three groups: persistent asthma, clinical- and complete asthma remission. The aim of the study in **Chapter 3** was to determine whether asthma remission persisted during this long-term follow-up, and which childhood factors are associated with clinical and/or complete asthma remission during adulthood.

Chapter 4 focuses on the number of exhaled particles in asthma patients, asthma remission subjects, and healthy individuals, enrolled in the exploring Asthma ReMission by Single-cell TRansciptiONal sequencinG (ARMSTRONG) study. Small airway disease is thought to close distal airways, consequently blocking non-volatile particles to be exhaled. Analysis of Particles of Exhaled Air (PExA) is a novel tool that enables measurement of the exhaled particle mass, which are produced by opening- and closing

Chapter I General introduction

of the lining tract fluid in the small airways. Our hypothesis is that the PExA mass is reduced in asthmatics and individuals with clinical asthma remission, while subject with complete asthma remission and healthy controls exhale more PExA mass. In line with this, we hypothesize that PExA correlates well with already established small airways parameters.

Chapter 5 describes the differences of small airways disease and inflammation in asthmatics, in subjects in clinical or complete asthma remission, and in healthy individuals, who participated in the ARMSTRONG cohort. Although it is known that subjects with clinical remission still express a degree of lung function impairment, bronchial hyperresponsiveness and comparable levels of airway inflammation [13], no data is published addressing their small airways function and inflammatory levels, compared to both asthmatics and healthy controls. We hypothesize that the parameters linked to small airways dysfunction and airway inflammation are similar in asthmatics and subjects with clinical asthma remission. Second, we anticipate that complete asthma remission subjects regain their pulmonary function, showing comparable features like healthy controls.

In **Chapter 6**, we apply a recently published model that predicts asthma remission on our own Dutch asthma remission cohorts. Authors of the Childhood Asthma Management Program (CAMP) trial showed that a combination of clinical features yields more that 80% probability to achieve asthma remission in young adulthood [87].

In **Chapter 7**, we review the asthma-obesity relationship. We focus on the increased risk to develop asthma in overweight and obese individuals, the effect of physical inactivity on asthma, the link between obesity and airway inflammation, the mechanical effects of obesity on asthma, the anti-inflammatory therapy response in obese asthmatics, the improvements of asthma symptoms due to weight loss, and the overweight comorbidities affecting asthma.

In **Chapter 8,** we apply single-cell transcriptomics – a novel technique for asthma endotyping - on bronchial biopsies, bronchial brushes and nasal brushes extracted from asthmatics and healthy controls in the ARMSTRONG study, as well as lung parenchyma tissue from donor lungs. Two methods of single-cell RNA-sequencing are

used: SmartSeq2 analysis of FACS-sorted epithelial and CD4 T cells from airway wall and the analysis of total cell suspensions using the IOXGenomics Chromium platform for single cell RNA sequencing [88,89]. Based on the transcriptomic profile of each cell, a cellular landscape of upper- and lower airways, and lung parenchyma is charted, which enables us to identify proportions and transcriptional cell-typing of structural and inflammatory cells between locations and asthma versus healthy.

In **Chapter 9**, we assess whether airway remodeling can be assessed by optical coherence tomography (OCT). In this chapter, we align histological stainings of airway wall sections with paired ex-vivo OCT-images, both derived from five lobectomy specimens. The airway wall sections are stained for various extracellular matrix components, i.e. total collagen, collagen AI, Masson's Trichrome, elastin, and fibronectin. We correlate the area and intensity of both the stained airway wall and the OCT images.

In **Chapter 10**, we investigate whether serum periostin – a marker linked to TH2-driven inflammation and thereby a potential marker for ICS responsiveness in asthma – is elevated in COPD patients compared to healthy controls. Second, we assess if periostin levels are associated with cross-sectional and longitudinal characteristics, including levels of inflammatory cells in three compartments (i.e. blood, sputum, and bronchial biopsies) and amount of bronchial extracellular matrix components in COPD patients.

In Chapter 11, the potential to use gene-expression to further endotype conditions such as asthma and COPD is explored. In 2014, Baines et al., demonstrated that unbiased clustering based on gene-expression was able to distinguish three different phenotypes in asthma [90]. In this chapter, we use a COPD-associated gene signature from another cohort to cluster COPD patients based on their RNA sequencing data obtained from bronchial biopsies during the Groningen and Leiden Universities Corticosteroids in Obstructive Lung Disease (GLUCOLD) study [91,92]. Subsequently, we assess whether these clusters were clinically different cross-sectionally and longitudinally.

## Ι

#### References

- GINA. Pocket Guide For Asthma Management and Prevention. Updated 2019. 2019.
- Liddell H. Liddell and Scott's an intermediate
   Greek-English lexicon. 7th editio. London:
   Oxford University Press 2000.
   Lancet 2017;390:980-7.
   Ito JT, Lourenço JD, Righetti RF, et
   al. Extracellular Matrix Component
- Netuveli G, Hurwitz B, Sheikh A. Lineages of language and the diagnosis of asthma. J R Soc Med 2007; Jan 100(1):19– 24.
- Gauthier M, Ray A, Wenzel SE. Evolving Concepts of Asthma. Am J Respir Crit Care Med 2015;192:660–8.
- 5. Lötvall J, Akdis CA, Bacharier LB, et al. Asthma endotypes: A new approach to classification of disease entities within the asthma syndrome. J Allergy Clin Immunol 2011;127:355–60.
- 6. Lange P, Halpin DM, O'Donnell DE, et al. Diagnosis, assessment, and phenotyping of COPD: beyond FEV<sub>1</sub>. Int J Chron Obstruct Pulmon Dis 2016;11 Spec Iss:3–12.
- Springfield M. Merriam-Webster's collegiate dictionary. 11th editi. Merriam-Webser Inc. 2008.
- 8. Tyler SR, Bunyavanich S. Leveraging -omics for asthma endotyping. J Allergy Clin Immunol 2019;144:13–23.
- Wenzel SE. Asthma phenotypes: the evolution from clinical to molecular approaches. Nat Med 2012;18:716–25.
- 10. Anderson GP. Endotyping asthma: new insights into key pathogenic mechanisms in a complex, heterogeneous disease. Lancet 2008;372:1107–19.

- II. Agustí A, Celli B, Faner R. What does endotyping mean for treatment in chronic obstructive pulmonary disease?

  Lancet 2017;390:980–7.
- al. Extracellular Matrix Component Remodeling in Respiratory Diseases: What Has Been Found in Clinical and Experimental Studies? Cells 2019;8:342.
- Broekema M, Timens W, Vonk JM, et al.
   Persisting remodeling and less airway
   wall eosinophil activation in complete
   remission of asthma. Am J Respir Crit Care
   Med 2011;183:310-6.
- 14. Kim B-S, Lee E, Lee M-J, et al. Different functional genes of upper airway microbiome associated with natural course of childhood asthma. Allergy 2018;73:644-52.
- 15. Brinkman P, Wagener AH, Hekking P-P, et al. Identification and prospective stability of electronic nose (eNose)—derived inflammatory phenotypes in patients with severe asthma. J Allergy Clin Immunol 2019;143:1811-1820.e7.
- 16. Woodruff PG, Modrek B, Choy DF, et al. T-helper Type 2-driven Inflammation Defines Major Subphenotypes of Asthma. Am J Respir Crit Care Med 2009;180:388-95.
- 7. Woodruff PG, Boushey HA, Dolganov GM, et al. Genome-wide profiling identifies epithelial cell genes associated with asthma and with treatment response to corticosteroids. Proc Natl Acad Sci 2007;104:15858–63.

- 18. Morrow Brown H. Treatment of chronic asthma with prednisolone; significance of eosinophils in the sputum. Lancet 1958;272:1245–7.
- 19. Bel EH. Clinical phenotypes of asthma. 27. Olivieri M, Zock J-P, Accordini S, et Curr Opin Pulm Med 2004;10:44–50. al. Risk factors for new-onset cat
- 20. Moore WC, Meyers DA, Wenzel SE, et al. Identification of Asthma Phenotypes Using Cluster Analysis in the Severe Asthma Research Program. Am J Respir Crit Care Med 2010;181:315–23.
- 21. Hanania NA, Wenzel S, Rosén K, et al. Exploring the effects of omalizumab in allergic asthma: an analysis of biomarkers in the EXTRA study. Am J Respir Crit Care Med 2013;187:804–11.
- 22. Wenzel S, Ford L, Pearlman D, et al. Dupilumab in Persistent Asthma with Elevated Eosinophil Levels. N Engl J Med 2013;368:2455–66.
- 23. Corren J, Lemanske RF, Hanania NA, et al. Lebrikizumab treatment in adults with asthma. N Engl J Med 2011;365:1088–98.
- 24. Castro M, Wenzel SE, Bleecker ER, et al. Benralizumab, an anti-interleukin 5 receptor α monoclonal antibody, versus placebo for uncontrolled eosinophilic asthma: a phase 2b randomised dose-ranging study. Lancet Respir Med 2014;2:879–90.
- 25. Sporik R, Holgate ST, Platts-Mills TA, et al. Exposure to house-dust mite allergen (Der p I) and the development of asthma in childhood. A prospective study. N Engl J Med 1990;323:502–7.

- 26. Permaul P, Hoffman E, Fu C, et al. Allergens in urban schools and homes of children with asthma. Pediatr Allergy Immunol 2012;23:543-9.
- 27. Olivieri M, Zock J-P, Accordini S, et al. Risk factors for new-onset cat sensitization among adults: a population-based international cohort study. J Allergy Clin Immunol 2012;129:420–5.
- 28. Amelink M, de Nijs SB, de Groot JC, et al. Three phenotypes of adult-onset asthma. Allergy 2013;68:674–80.
- 29. Walford HH, Doherty TA. Diagnosis and management of eosinophilic asthma: a US perspective. J Asthma Allergy 2014;7:53.
- 30. Brussino L, Heffler E, Bucca C, et al. Eosinophils Target Therapy for Severe Asthma: Critical Points. Biomed Res Int 2018;2018:1-6.
- 31. Pavord ID, Chanez P, Criner GJ, et al.

  Mepolizumab for Eosinophilic Chronic

  Obstructive Pulmonary Disease. N Engl J

  Med 2017;377:1613–29.
- 32. Conway SJ, Izuhara K, Kudo Y, et al. The role of periostin in tissue remodeling across health and disease. Cell Mol Life Sci 2014;71:1279–88.
- 33. Li W, Gao P, Zhi Y, et al. Periostin: its role in asthma and its potential as a diagnostic or therapeutic target. Respir Res 2015;16:57.
- 34. Peters-Golden M, Swern A, Bird SS, et al. Influence of body mass index on the response to asthma controller agents. Eur Respir J 2006;27:495–503.

- 35. Agrawal A, Mabalirajan U, Ahmad T, et al. Emerging Interface between Metabolic Syndrome and Asthma. Am J Respir Cell Mol Biol 2011;44:270-5.
- 36. Carpaij OA, van den Berge M. The 45. Weibel ER. Principles and methods for asthma-obesity relationship: underlying mechanisms and treatment implications. Curr Opin Pulm Med 2018;24:42-9.
- Telenga ED, Tideman SW, Kerstjens HAM, et al. Obesity in asthma: more neutrophilic inflammation as a possible explanation for a reduced treatment response. Allergy 2012;67:1060-8.
- 38. Newcomb DC, Peebles RS. Th17mediated inflammation in asthma. Curr Opin Immunol 2013;25:755-60.
- Thuesen BH, Husemoen LLN, Hersoug L-G, et al. Insulin resistance as a predictor of incident asthma- like symptoms in adults. Clin Exp Allergy 2009;39:700-7.
- 40. Vonk JM, Postma DS, Boezen HM, et al. Childhood factors associated with asthma remission after 30 year follow up. Thorax 2004;59:925-9.
- 41. Postma DS, Brightling C, Baldi S, et al. Exploring the relevance and extent of small airways dysfunction in asthma (ATLANTIS): baseline data from a prospective cohort study. Lancet Respir Med 2019;7:402-16.
- 42. Lipworth B, Manoharan A, Anderson W. Unlocking the quiet zone: the small airway asthma phenotype. Lancet Respir Med 2014;2:497-506. doi:10.1016/S2213-2600(14)70103-1
- 43. Bjermer L. The role of small airway disease in asthma. Curr Opin Pulm Med 2014;20:23-30.

- 44. Youngmans W. The mechanics of breathing. In: Best C, ed. The Physiological Basis of Medicale Practice. Baltimore: Williams & Wilkins 1061, 466-80.
- the morphometric study of the lung and other organs. Lab Invest 1963;12:131-55.
- 46. Kraft M, Pak J, Martin RJ, et al. Distal lung dysfunction at night in nocturnal asthma. Am J Respir Crit Care Med 2001;163:1551-6.
- 47. Zeidler MR, Goldin JG, Kleerup EC, et al. Small airways response to naturalistic cat allergen exposure in subjects with asthma. I Allergy Clin Immunol 2006;118:1075-81.
- 48. Nihlberg K, Andersson-Sjöland A, Tufvesson E. et al. Altered matrix production in the distal airways of individuals with asthma. Thorax 2010;65:670-6.
- 49. McNulty W, Usmani OS. Techniques of assessing small airways dysfunction. Eur Clin Respir J 2014;1:25898.
- 50. Gafar F, Boudewijn IM, Cox CA, et al. Predictors of clinical response to extrafine and non-extrafine particle inhaled corticosteroids in smokers and ex-smokers with asthma. Respir Res 2018;19:256.
- 51. van den Berge M, ten Hacken NHT, Cohen J, et al. Small Airway Disease in Asthma and COPD. Chest 2011;139:412-23.
- 52. WHO | Chronic obstructive pulmonary disease (COPD). WHO 2017.
- 53. Global Initiative for Chronic Obstructive Lung Disease. 2019.

- 54. Silverman EK, Chapman HA, Drazen JM, et al. Genetic epidemiology of severe, early-onset chronic obstructive pulmonary disease. Risk to relatives for airflow obstruction and chronic bronchitis. Am J Respir Crit Care Med 1998;157:1770-8.
- 55. Agusti A, Bel E, Thomas M, et al. Treatable traits: toward precision medicine of chronic airway diseases. Eur Respir J 2016;47:410-9.
- 56. Klooster K, ten Hacken NHT, Hartman JE, et al. Endobronchial Valves for Emphysema without Interlobar Collateral Ventilation. N Engl J Med 2015;373:2325-35.
- 57. Calverley PM, Rabe KF, Goehring U-M, et al. Roflumilast in symptomatic chronic obstructive pulmonary disease: two randomised clinical trials. Lancet 2009;374:685-94.
- 58. George L, Brightling CE. Eosinophilic airway inflammation: role in asthma and chronic obstructive pulmonary disease. Ther Adv Chronic Dis 2016;7:34-51. d
- 59. Singh D, Kolsum U, Brightling CE, et al. Eosinophilic inflammation in COPD: prevalence and clinical characteristics. Eur Respir J 2014;44:1697-700.
- 60. Casanova C, Celli BR, de-Torres JP, et al. Prevalence of persistent blood eosinophilia: relation to outcomes in patients with COPD. Eur Respir J 2017;50:1701162.

- 61. Liesker IJW, Bathoorn E, Postma DS, et al. Sputum inflammation predicts exacerbations after cessation of inhaled corticosteroids in COPD. Respir Med 2011;105:1853-60.
- 62. Hogg JC, Chu F, Utokaparch S, et al. The Nature of Small-Airway Obstruction in Chronic Obstructive Pulmonary Disease. N Engl J Med 2004;350:2645-53.
- 63. Barnes PJ. Immunology of asthma and chronic obstructive pulmonary disease. Nat Rev Immunol 2008;8:183-92.
- 64. Siddiqui SH, Pavord ID, Barnes NC, et al. Blood eosinophils: a biomarker of COPD exacerbation reduction with inhaled corticosteroids. Int J Chron Obstruct Pulmon Dis 2018;13:3669-76.
- 65. Grainge CL, Lau LCK, Ward JA, et al. Effect of Bronchoconstriction on Airway Remodeling in Asthma. N Engl J Med 2011;364:2006-15.
- Barbers RG, Papanikolaou IC, Koss MN, et al. Near fatal asthma: clinical and airway biopsy characteristics. Pulm Med 2012;2012:820608.
- 67. Al-Muhsen S, Johnson JR, Hamid Q. Remodeling in asthma. J Allergy Clin Immunol 2011;128:451-62; quiz 463-4.
- 68. Beckett PA, Howarth PH. Pharmacotherapy and airway remodelling in asthma? Thorax 2003;58:163-74.
- 69. Holgate ST, Wenzel S, Postma DS, et al. Asthma. Nat Rev Dis Prim 2015;1:15025.
- 70. James AL, Wenzel S. Clinical relevance of airway remodelling in airway diseases. Eur Respir J 2007;30:134-55.

- 71. Boulet L-P, Laviolette M, Turcotte H, et al. Bronchial Subepithelial Fibrosis Correlates With Airway Responsiveness to Methacholine. Chest 1997;112:45–52.
- 72. Ward C, Pais M, Bish R, et al. Airway inflammation, basement membrane thickening and bronchial hyperresponsiveness in asthma. Thorax 2002;57:309–16.
- 73. Hartley RA, Barker BL, Newby C, et al. Relationship between lung function and quantitative computed tomographic parameters of airway remodeling, air trapping, and emphysema in patients with asthma and chronic obstructive pulmonary disease: A single-center study. J Allergy Clin Immunol 2016;137:1413-1422. e12.
- 74. Jeffery PK. Remodeling in Asthma and Chronic Obstructive Lung Disease. Am J Respir Crit Care Med 2001;164:S28–38.
- 75. Saglani S, Payne DN, Zhu J, et al. Early Detection of Airway Wall Remodeling and Eosinophilic Inflammation in Preschool Wheezers. Am J Respir Crit Care Med 2007;176:858–64.
- 76. St-Laurent J, Bergeron C, Pagé N, et al. Influence of smoking on airway inflammation and remodelling in asthma. Clin Exp Allergy 2008;38:1582-9.
- 77. Krogsgaard Chawes BL, Bønnelykke K, Bisgaard H. Elevated Eosinophil Protein X in Urine from Healthy Neonates Precedes Development of Atopy in the First 6 Years of Life. Am J Respir Crit Care Med 2011;184:656–61.

- 78. Kirby M, Ohtani K, Lopez Lisbona RM, et al. Bronchial thermoplasty in asthma: 2-year follow-up using optical coherence tomography. Eur Respir J 2015;46:859–62.
- 79. Goorsenberg AWM, d'Hooghe JNS, de Bruin DM, et al. Bronchial Thermoplasty-Induced Acute Airway Effects Assessed with Optical Coherence Tomography in Severe Asthma. Respiration 2018;96:564-70.
- 80. Dournes G, Marthan R, Berger P, et al.
  Bronchi wall and lumen volumes to
  assess airway remodeling in asthma by
  using CT: An innovative method? J Allergy
  Clin Immunol 2014;133:1777.
- 81. Niimi A, Matsumoto H, Takemura M, et al. Relationship of airway wall thickness to airway sensitivity and airway reactivity in asthma. Am J Respir Crit Care Med 2003;168:982–8.
- 82. Chen Y, Ding M, Guan WJ, et al. Validation of human small airway measurements using endobronchial optical coherence tomography. Respir Med 2015;109:1446–53.
- 83. Williamson JP, McLaughlin RA, Noffsinger WJ, et al. Elastic Properties of the Central Airways in Obstructive Lung Diseases Measured Using Anatomical Optical Coherence Tomography. Am J Respir Crit Care Med 2011;183:612–9.
- 84. Adams DC, Miller AJ, Applegate MB, et al. Quantitative assessment of airway remodelling and response to allergen in asthma. Respirology Published Online First: 7 March 2019.

- 85. Yang Y, Wang T, Biswal NC, et al. Optical scattering coefficient estimated by optical coherence tomography correlates with collagen content in ovarian tissue. J Biomed Opt 2011;16:090504.
- 86. Ud-Din S, Foden P, Stocking K, et al. Objective assessment of dermal fibrosis in cutaneous scarring: using optical coherence tomography, high frequency ultrasound and immunohisto-morphometry of human skin. Br J Dermatol 2019;:0–2.
- 87. Wang AL, Datta S, Weiss ST, et al.

  Remission of persistent childhood asthma: early predictors of adult outcomes. J Allergy Clin Immunol 2018;0.
- 88. Vieira Braga FA, Kar G, Berg M, et al. A cellular census of human lungs identifies novel cell states in health and in asthma.

  Nat Med 2010;25:1153-63.
- 89. Picelli S, Faridani OR, Björklund ÅK, et al. Full-length RNA-seq from single cells using Smart-seq2. Nat Protoc 2014;9:171–81.
- 90. Baines KJ, Simpson JL, Wood LG, et al. Sputum gene expression signature of 6 biomarkers discriminates asthma inflammatory phenotypes. J Allergy Clin Immunol 2014;133:997–1007.
- gi. Lapperre TS, Snoeck-Stroband JB, Gosman MME, et al. Effect of fluticasone with and without salmeterol on pulmonary outcomes in chronic obstructive pulmonary disease: a randomized trial. Ann Intern Med 2009;151:517-27.

92. van den Berge M, Steiling K, Timens W, et al. Airway gene expression in COPD is dynamic with inhaled corticosteroid treatment and reflects biological pathways associated with disease activity. Thorax 2014;69:14–23.