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Macrophage-mediated phagocytosis of bacteria adhering on biomaterial surfaces

Da Silva Domingues, Joana

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CHAPTER II

Macrophage response to staphylococcal biofilms on crosslinked poly(ethylene) glycol polymer coatings and common biomaterials *in vitro*

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ABSTRACT

Biomaterial-associated infections (BAI) are serious clinical complications that threaten the longevity of implanted devices and lead to high morbidity and mortality. Poly(ethylene)glycol (PEG) coatings have been studied as a strategy to reduce the incidence of BAI by reducing protein deposition that promotes pathogen adhesion and growth on device surfaces. Despite their effectiveness to reduce protein adsorption and a hundred-fold reduction in bacterial adhesion, PEG-based coatings still facilitate weak bacterial adhesion that can form an initial basis for biofilms. Here, we describe a methodology enabling direct, quantitative and detailed qualitative *in situ* observation of macrophage morphology, migration and phagocytosis of bacteria. *In vitro* interaction of macrophages with *Staphylococcus epidermidis* 3399, adhering to commercial, crosslinked PEG-based coatings (OptiChem®), was compared with fluorinated ethylene propylene, silicone rubber and glass. Adhesion, phagocytosis, and migration were studied real-time in a parallel-plate flow chamber. Macrophages cultured on OptiChem® coatings showed enhanced migration and phagocytosis of bacteria compared to common biomaterials. Bacterial clearance per macrophage on both inert and reactive OptiChem® coatings were about three times higher than on the common biomaterials studied, corresponding with up to 70% reduction in bacterial numbers on OptiChem®, whereas on the biomaterials less than 40% bacterial reduction was obtained. These findings show that bacterial clearance from cross-linked PEG-based coatings by macrophages is more effective than from common biomaterials, possibly resulting from weak adhesion of bacteria on OptiChem®. Moreover, macrophages exhibit higher mobility on OptiChem® retaining an improved capability to clear bacteria from larger areas than from other common biomaterials, where they appear more immobilized.

INTRODUCTION

Placement of indwelling medical devices into the human body to support and restore function has become common practice in modern medicine with reasonable overall success rates. In 2006, for instance, nearly 800,000 primary total hip and knee arthroplasties were performed in the United States alone [1]. Biomaterial-associated infections (BAI) and adverse interactions between the indwelling device and the surrounding tissues and cells are, however, factors that threaten the device's functionality and longevity [2]. BAI, although of relatively low incidence, represents a serious complication of extensive significance, with related high morbidity and mortality rates, as well as with high associated health care costs. Despite advances in surgical techniques, perioperative contamination remains the most common route for the infection of biomaterial devices [3]. Microorganisms, usually sourced from the patient's skin, adhere to the implant surface, colonize it and rapidly form biofilms [4]. Generally, microorganisms embedded in biofilms are much less susceptible to antimicrobial treatments [5] and host immune mechanisms than planktonic organisms and hence, infection usually persists until the device is removed.

BAI pathogenesis depends on many factors, such as implant site, device type and the patient's general health status, but also on the interaction between the biomaterial surface, the host's immune system and the infecting pathogen [1,6]. Following biomaterial implantation, tissue trauma and injury trigger a cascade of physiological events that activate the immune system [7]. Neutrophils and monocytes / macrophages are the major host inflammatory cell populations that arrive within minutes to hours at the implant site [7,8]. In contrast to neutrophils that may disappear within hours, macrophages increase in numbers over time and

remain at the implant surface for several weeks depending on the severity of the injury [8]. Macrophages orchestrate the host inflammation process and eventual foreign body reactions, but are also an important active component in pathogen clearance. During bacterial infection *in vivo*, macrophages adhere to the infected tissue and detect bacteria via cell surface receptors [7]. After recognition and pathogen attachment, macrophages engulf bacteria (phagocytosis) and activate cellular functions such as proliferation, secretion of proteins and cytokines, and respiratory bursts to destroy phagocytosed bacteria and recruit other cells from the adaptive immune system, as for example T and B lymphocytes [9]. However, the presence of a biomaterial affects the immune system's response to bacterial infection, and the biomaterial surface chemistry can stimulate [7] or reduce macrophage adhesion, phagocytic activity and migration [10,11]. Therefore, macrophage-biomaterial-bacteria interactions are crucial factors influencing pathogenesis of BAI [10].

Poly(ethylene) glycol (PEG) has been promoted as an infection-resistant biomaterial coating due to low protein adsorption and hundred-fold reductions in adhering bacteria with respect to common biomaterials [12-15]. Nonetheless, this performance does not prevent the formation of weakly adhering, more mature biofilms [14,16]. Currently, it is unknown how macrophages deal with the few bacteria adhering to PEG-based coatings. Therefore, the aim of this study was to investigate the *in vitro* response of macrophages to *Staphylococcus epidermidis* adhering to a fully characterized commercial, multi-component crosslinked PEG-based polymer coating (OptiChem®, Accelr8 Technology Corp., Denver, CO, USA) and to compare this with the response of the macrophages on common biomaterials such as fluorinated ethylene propylene (FEP) and silicone rubber. Glass was included since it acted as the substratum for the polymer coating and is a hydrophilic material,

unlike FEP and silicone rubber. For convenience however, glass will be mentioned in this manuscript under the denominator “common biomaterials”, as opposed to PEG-based polymer coatings.

MATERIALS AND METHODS

Substrata

OptiChem®-coated glass slides (Accelr8 Technology, now commercially available as Schott-Nexterion™ Slide H) were supplied by Accelr8 Technology Corporation. OptiChem® is a multi-component, cross-linked transparent and robust polymer coating, having PEG as its active component. The surface coating has an amine-reactive (i.e. an NHS active ester) terminal chemical functionality to allow specific immobilization of biomolecules. The NHS chemistry can also be deactivated to provide a surface with very low, nonspecific binding of biological molecules [17-19]. Extensive surface chemistry and analytical details regarding the coating and its bio-immobilization properties have been published [17-19].

OptiChem® was applied on optical-grade glass slides by spin coating and curing. Slides were stored at -20°C until use. Half of the coated slides were deactivated by quenching the NHS surface groups (“inert Optichem®”) using hydroxyethylamine [18]. The remaining slides were used in its NHS-reactive form, denoted here as “reactive OptiChem®”. Macrophage activity was also evaluated on FEP, silicone rubber and glass. FEP, silicone rubber and glass slides were cleaned in 2% RBS 35 detergent solution (Omnilabo International BV, Breda, The Netherlands) under sonication and rinsed with demineralized water, submerged in methanol, and washed with demineralized water again. All samples were sterilized in 70% ethanol for 10

min and rinsed with sterile, demineralized water and finally with sterile phosphate buffered saline (PBS, 10 mM potassium phosphate, 150 mM NaCl, pH 6.8).

Bacterial strain and growth conditions

S. epidermidis 3399 is a clinical isolate from the skin and was used because skin-derived organisms like *S. epidermidis* are often involved in perioperative contamination of biomaterial implant surfaces. The staphylococcus was first grown aerobically overnight at 37°C on blood agar plates from a frozen stock. The plates were kept at 4°C, never longer than 2 weeks. One colony was used to inoculate 10 ml of tryptone soya broth (TSB, OXOID, Basingstoke, England), which was incubated for 24 h at 37°C and used to inoculate a second culture in 200 ml TSB. Bacteria were harvested after overnight growth by centrifugation (5 min at 5,000 g at 10°C) and washed twice with sterile PBS. Bacteria were resuspended in sterile PBS to a concentration of 3×10^8 bacteria/ml.

Cell culture conditions

J774A.1 mouse macrophages were grown in tissue culture polystyrene (TCPS) flasks (Greiner Bio-One, Frickenhausen, Germany), and maintained in Dulbecco's modified Eagle's medium (DMEM) supplemented with 4.5g/l D-glucose, pyruvate, and 10% fetal bovine serum (DMEM + 10% FBS) at 37°C in a humidified atmosphere of 5% CO₂. Cells were passaged every four days at 70 – 80% confluency by scraping. The cells were passaged up to a maximum of seven times.

J774A.1 morphology, migration and phagocytic activity

Macrophage morphology, migration and phagocytic activity on the crosslinked PEG-based coatings and the different common biomaterials were assessed using real-time *in situ* image analysis in a parallel-plate flow chamber with a CCD camera

(Basler Ahrensburg, Germany) mounted on a phase-contrast microscope (Olympus BH-2; Olympus, Tokyo, Japan) (for a detailed description of the system, see Busscher *et al.* [20]). Assays were performed on the bottom plate of the flow chamber containing the substrata under study. The system was first filled with sterile PBS to remove air bubbles from the tubing and chamber, and perfused for 30 min with a laminar flow of 1.5 ml/min, corresponding to a shear rate of 11 s^{-1} . Then, flow was switched to bacterial suspension in PBS that circulated at the same flow rate until the density of adhering bacteria had reached on average 4.2×10^5 bacteria/cm² on all substrata, as evaluated real-time with the image analysis system. Subsequently, the suspension was switched once more to sterile PBS to remove unbound bacteria from the system, which did not remove any adhering bacteria as could be experimentally established by virtue of the use of real-time imaging, since the flow rate stayed the same and no air-liquid interface was involved. The flow chamber was warmed up to 37°C. Then, a macrophage suspension consisting of 7.5×10^5 cells/ml in DMEM + 10% FBS was introduced into the system. Once the entire volume of buffer inside the chamber was replaced by the cell suspension, flow was stopped. Images were collected throughout the assay for 120 min at 1 min intervals. Phagocytic activity was determined by comparing the number of bacteria adhering per cm² on the substrata at different time intervals. In addition, the difference between the initial numbers of bacteria adhering to the substratum prior to exposure to macrophages and the final bacterial density after 120 min exposure to the macrophages was calculated to determine the number of bacteria ingested per adherent macrophage. Bacterial growth during 120 min phagocytic activity was minimal in DMEM + 10% FBS and therefore neglected in these calculations.

RESULTS

Macrophage morphology

Phase-contrast images of cultured J774A.1 murine macrophages interacting with bacteria adhering to crosslinked PEG-based coatings and the different common biomaterials in DMEM + 10% FBS are shown in Figure 1. Macrophages adhering to common biomaterials maintain a more spherical shape throughout the experiment, while those interacting with inert and reactive OptiChem® acquire a more elongated form increasing the contact area with the surface.

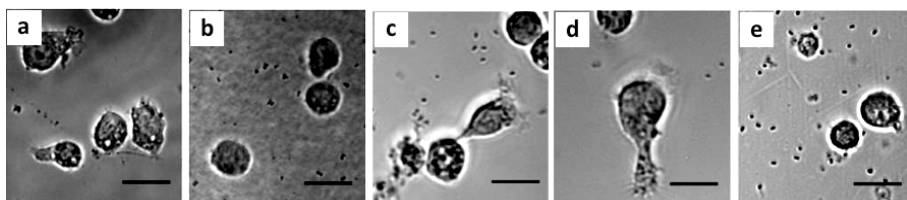


Figure 1. Phase-contrast microscopic images showing the different morphologies of macrophages. Macrophages adhering to different cross-linked PEG-based coatings and common biomaterials in the presence of *S. epidermidis* 3399 in serum-containing culture media in the flow chamber. (a) FEP, (b) Silicone rubber, (c) Inert OptiChem®, (d) Reactive OptiChem®, and (e) Glass. The bar denotes 20 μm .

Macrophage migration and phagocytic activity

Macrophage activity was assessed microscopically in real-time. The number of macrophages adhering per cm^2 on each substratum is presented in Table 1. J774A.1 cell migration and phagocytosis of bacteria on glass, inert and reactive OptiChem® in the presence of adhering staphylococci are shown in Figure 2.

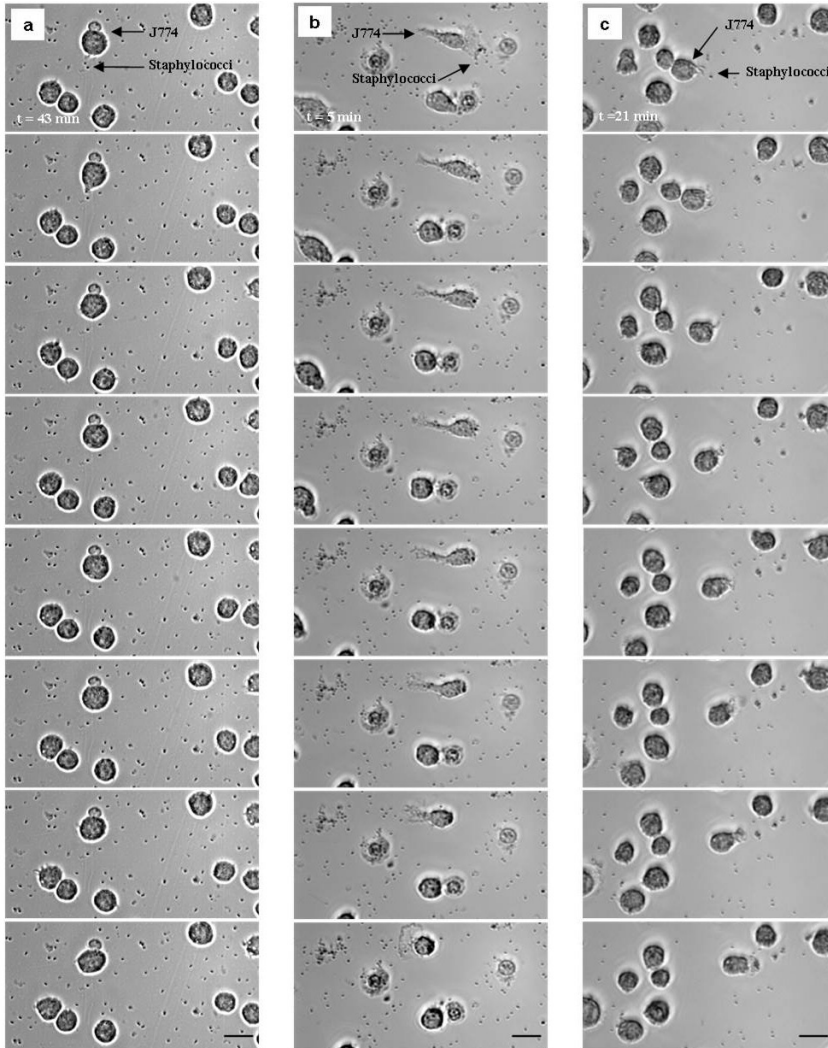


Figure 2. Time-lapse light micrographs of the migration and phagocytosis of *S. epidermidis* 3399 by murine macrophages. Phagocytosis was followed in serum-containing culture media in the flow chamber on (a) glass, (b) inert OptiChem®, and (c) reactive OptiChem®. The interval between the micrographs is 2 min, increasing from top to bottom. “t” denotes the time of exposure to macrophages. The bar denotes 20 µm. See supplementary information for video time-lapse files of macrophage real-time migration and phagocytosis.

Macrophages adhering to common biomaterials are immobilized to the substratum and their migration is restricted to a few μms . Consequently, macrophages only phagocytose bacteria attached in their close surroundings via the projection of pseudopodia. In contrast, macrophages adhering on inert and reactive OptiChem® coatings are more mobile, migrating relatively freely over the substratum towards adherent staphylococci. Time-lapse videos of macrophage migration and phagocytosis of *S. epidermidis* on the different substrata are available in the supporting information.

Under sterile operating conditions, the number of bacteria-carrying particles that fall on an open wound varies between 10^2 and 10^5 per cm^2 [21-23]. In this study, the initial bacterial density on all substrata was $(4.2 \pm 0.9) \times 10^5$ per cm^2 before macrophages were added into the system. The number of bacteria on the surface can thus be considered reasonably close to a clinically relevant situation of perioperative contamination. After exposure to macrophages, the numbers of adhering staphylococci decreased significantly.

Table 1. Numbers of *S. epidermidis* remaining adherent on the surface per unit surface area after exposure to macrophages (N_{2h}) for 120 min, together with the numbers of macrophages per unit surface area and the number of staphylococci taken per macrophage for the five substrata involved in this study. The initial number of adhering staphylococci prior to exposure to macrophages was on average $4.2 \times 10^5 \text{ cm}^{-2}$, as determined during an experiment using real-time *in situ* observation. SD over six images per substratum surface.

Substratum	N_{2h} ($10^5/\text{cm}^2$)	Macrophages ($10^4/\text{cm}^2$)	Bacteria/macrophage
FEP	1.8 ± 0.2	6.3 ± 0.5	2.4 ± 0.4
Silicone rubber	3.0 ± 0.5	4.1 ± 0.8	1.3 ± 0.9
Inert OptiChem®	1.4 ± 0.3	4.3 ± 0.5	5.4 ± 1.1
Reactive OptiChem®	1.1 ± 0.3	5.6 ± 0.7	6.9 ± 1.0
Glass	4.1 ± 0.3	5.2 ± 0.8	2.8 ± 0.6

Figure 3 shows the percentage of bacteria left adhering on the surface as a function of exposure time to macrophages. Bacterial clearance per macrophage on inert and reactive OptiChem® coatings was on average three times higher than on the common biomaterials (see Table 1).

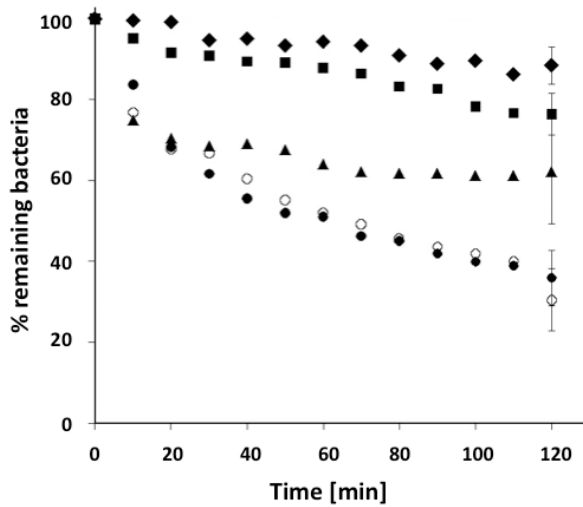


Figure 3. Percentage of adhering *S. epidermidis* 3399 remaining on the different surfaces after exposure to macrophages for 120 min. Experiments were done in serum-containing cell culture media with respect to their initial adherent density ($4.2 \times 10^5 \text{ cm}^{-2}$) on: FEP (▲), Silicone rubber (◆), Inert OptiChem® (○), Reactive OptiChem® (●), and Glass (■). Error bars represent the standard deviation over six images.

DISCUSSION

Macrophages are primary infiltrating immune system cells responding rapidly to wounding and implanted biomaterials, and are directly involved in the host inflammatory and foreign body response as well as in the defense against infectious pathogens. Macrophages adhere to device surfaces and remain at the implant-tissue interface for several days to realize their functions. Hence, the interaction between

macrophages and bacterially contaminated biomaterials is crucial in the development of BAI [8,10]. A mature biofilm is less likely to form if macrophages are able to remove and destroy microorganisms adhering on an implanted device. The response of macrophages to surfaces modified with PEG-based coatings has been assessed by others [24-26], but never on bacterially contaminated biomaterial surfaces as done here. Our study showed that macrophages phagocytosis of bacteria adhering on inert and reactive OptiChem® was similar for both coatings, but on average three times higher than on the common biomaterials included in this study. This difference and elevated phagocytic activity of macrophages to *S. epidermidis* adhering on cross-linked PEG-based coatings is attributed to an almost unlimited macrophage mobility on the PEG-based coating compared to common biomaterials. On Optichem®, macrophages reduced the numbers of adhering staphylococci by approximately 70% over a 2 h time period, as shown in Figure 3. There are no comparative data available in the literature to determine whether this is a high or low phagocytosis efficiency. In a recent study [27] phagocytosis of *S. epidermidis* and *Pseudomonas aeruginosa* on PEG-graft-polyacrylate (PEG-g-PA) copolymers has been studied. However, macrophages were allowed to adhere to the surface before bacteria were incorporated into the system, which is an entirely different model situation than our perioperative model.

The enhanced macrophage mobility and phagocytic activity on Optichem® coatings could result from weak cell-surface interactions between these cells and the PEG-based coatings, but also bacteria-surface interaction forces are important for effective phagocytosis. Adhesion forces between microorganisms and poly(ethylene) oxide (PEO) brush coatings have been assessed using atomic force microscopy [28] and found to be up to 10 times smaller for various *P. aeruginosa* strains on a PEO brush than on bare glass. Incremental increases in shear rate in a parallel-plate flow

chamber also indicated that the adhesion strength of *S. epidermidis* and *Staphylococcus aureus* is decreased on PEO-coated silicone rubber. More than 85% of these bacteria could be sheared off from the PEO brush coating whereas up to 10% of adherent bacteria could be stimulated to detach from hydrophobic silicone rubber [15]. Analogous to bacterial interactions with polymer brush coatings, macrophages adhering to OptiChem® coatings may be expected to experience weak adhesion forces as well, allowing them to move freely over the substratum towards adhering bacteria. Low adsorption of serum proteins on PEG-based surfaces, and specifically for OptiChem® coatings [18] produces poor cell adhesion [18,19]. Macrophage-surface interactions depend less on cell matrix-type adhesive proteins in contrast to other cell types [29], and macrophage surface mobility is increased without a substantial surface-adsorbed protein layer. This occurs on both the inactivated (inert) PEG surface as well as that retaining the NHS-reactive immobilizing chemistry. Weak interactions between adhering bacteria and OptiChem® coatings, as described above, may also help facilitate more efficient macrophage phagocytosis from these surfaces. This is an advantage, as phagocyte-mediated clearance of surface-adhered bacteria is more difficult for macrophages than their clearance of planktonic bacteria [30].

In vivo, the interaction between proteins, pathogens and the host defense cells at the biomaterial-tissue interface is a complex process where each may contribute to bacterial survival and persistence on biomaterials and in adjacent tissues [2,10]. Host defense functions are suggested to be affected in the presence of an infected biomaterial, for example, by diminishing host phagocytic activities [10,11,31]. We demonstrate that macrophages can phagocytise adhering bacteria more effectively on PEG-based coatings. Although macrophages are not the only cell type present at the interface *in vivo*, these results for macrophages are relevant, in that macrophages remain at the implanted biomaterial surface for longer periods of time than other

cells [8]. Also an important factor in the persistence of BAI is bacterial survival within macrophages once ingested. This intraphagocyte survival mechanism is both pathogen and substratum-dependent [10]. That such bacterial survival within macrophages is favored on OptiChem® coatings was not the focus of this study and should be elucidated.

CONCLUSIONS

We introduced a novel *in vitro* methodology to enable direct, quantitative and detailed qualitative *in situ* observations of macrophage adherent morphology, migration and engulfment of surface-resident bacteria. In the current study, we employed this methodology to compare macrophage clearance of adhering staphylococci from cross-linked PEG-based coatings and different common biomaterials, including glass. Substratum surfaces were first contaminated with bacteria prior to exposure to cultured macrophages in serum-based media to mimic perioperative bacterial contamination conditions. Macrophages on cross-linked PEG-based coatings exhibited enhanced cell mobility compared to the mobility on FEP, silicone rubber and glass, likely due to weak macrophage-surface interaction forces arising from strongly hydrated, low protein-adsorbing cross-linked PEG-based coatings. This greater intrinsic cell mobility and associated weak bacterial-surface adhesion forces facilitated higher phagocytosis on the PEG surfaces. Macrophage-mediated bacterial clearance was about three times more effective on the multi-component cross-linked PEG-based coatings (OptiChem®) than on the common biomaterials included in this study, irrespective whether the surface was the inactivated or reactive NHS-derivatized PEG-based coating.

REFERENCES

1. Del Pozo JL, Patel R (2009) Infection associated with prosthetic joints. *N Engl J Med* 361: 787-794.
2. Gristina AG (1987) Biomaterial-centered infection: microbial adhesion *versus* tissue integration. *Science* 237: 1588-1595.
3. Busscher HJ, Ploeg RJ, Van der Mei HC (2009) Snapshot: biofilms and biomaterials; mechanisms of medical device related infections. *Biomaterials* 25: 4247-4248.
4. Knobben BAS, Engelsma Y, Neut D, Van der Mei HC, Busscher HJ, *et al.* (2006) Intraoperative contamination influences wound discharge and periprosthetic infection. *Clin Orthop Relat Res* 452: 236-241.
5. Stewart PS, Costerton JW (2001) Antibiotic resistance of bacteria in biofilms. *Lancet* 358: 135-138.
6. Kwakman PH, Te Velde AA, Vandenbroucke-Grauls CM, Van Deventer SJ, Zaat SA (2006) Treatment and prevention of *Staphylococcus epidermidis* experimental biomaterial-associated infection by bactericidal peptide 2. *Antimicrob Agents Chemother* 50: 3977-3983.
7. Xia Z, Triffitt JT (2006) A review on macrophage responses to biomaterials. *Biomed Mater* 1: R1-9.
8. Anderson JM (2004) Inflammation, wound healing, and the foreign-body response. In: Ratner BD, Hoffman AS, Schoen FJ, Lemons JE, editors. *Biomaterials Science. An Introduction to Materials in Medicine*. Elsevier, San Diego, CA. pp 296-304.
9. Berton G, Lowell CA (1999) Integrin signalling in neutrophils and macrophages. *Cell Signal* 11: 621-635.
10. Boelens JJ, Dankert J, Murk JL, Weening JJ, Van der Poll T, *et al.* (2000) Biomaterial-associated persistence of *Staphylococcus epidermidis* in pericatheter macrophages. *J Infect Dis* 181: 1337-1349.
11. Henke PK, Bergamini TM, Brittian KR, Polk HC Jr (1997) Prostaglandin E2 modulates monocyte MHC-II (Ia) suppression in biomaterial infection. *J Surg Res* 69: 372-378.
12. Norde W, Gage D (2004) Interaction of bovine serum albumin and human blood plasma with PEO-tethered surfaces: influence of PEO chain length, grafting density, and temperature. *Langmuir* 20: 4162-4167.
13. Saldarriaga Fernández IC, Van der Mei HC, Lochhead MJ, Grainger DW, Busscher HJ (2007) The inhibition of the adhesion of clinically isolated bacterial strains on multi-component cross-linked poly(ethylene glycol)-based polymer coatings. *Biomaterials* 28: 4105-4112.
14. Nejadnik MR, Van der Mei HC, Norde W, Busscher HJ (2008a) Bacterial adhesion and growth on a polymer brush-coating. *Biomaterials* 29: 4117-4121.
15. Nejadnik MR, Van der Mei HC, Busscher HJ, Norde W (2008b) Determination of the shear force at the balance between bacterial attachment and detachment in weak-adherence systems, using a flow displacement chamber. *Appl Environ Microbiol* 74: 916-919.
16. Saldarriaga Fernández IC, Van der Mei HC, Metzger S, Grainger DW, Engelsman AF, *et al.* (2010) *In vitro* and *in vivo* comparisons of staphylococcal biofilm formation on a cross-linked poly(ethylene glycol)-based polymer coating. *Acta Biomater* 6: 1119-1124.
17. Mao G, Metzger S, Lochhead MJ (2005) *US patent* 6,844,028, 2005.

18. Harbers GM, Emoto K, Greef C, Metzger SW, Woodward HN, et al. (2007) Functionalized poly(ethylene glycol)-based bioassay surface chemistry that facilitates bio-immobilization and inhibits nonspecific protein, bacterial, and mammalian cell adhesion. *Chem Mater* 19: 4405–4414.
19. Takahashi H, Emoto K, Dubey M, Castner DG, Grainger DW (2008) Imaging surface immobilization chemistry: correlation with cell patterning on non-adhesive hydrogel thin films. *Adv Funct Mater* 18: 2079-2088.
20. Busscher HJ, Van der Mei HC (2006) Microbial adhesion in flow displacement systems. *Clin Microbiol Rev* 19: 127-141.
21. Fitzgerald RH (1979) Microbiologic environment of the conventional operating-room. *Colloids Surf B Biointerfaces* 19: 127-141.
22. Oakes JA, Wood AJJ (1986) Infections in surgery. *N Engl J* 315: 1129-1138.
23. Nelson CL (1979) Environmental bacteriology in the unidirectional (horizontal) operating room. *Arch Surg* 114: 778–782.
24. Lynn AD, Kyriakides TR, Bryant SJ (2010) Characterization of the *in vitro* macrophage response and *in vivo* host response to poly(ethylene glycol)-based hydrogels. *J Biomed Mater Res A* 93: 941-953.
25. Schmidt DR, Kao WJ (2007) Monocyte activation in response to polyethylene glycol hydrogels grafted with RGD and PHSRN separated by interpositional spacers of various lengths. *J Biomed Mater Res A* 83: 617-625.
26. Jenney CR, Anderson JM (1999) Effects of surface-coupled polyethylene oxide on human macrophage adhesion and foreign body giant cell formation *in vitro*. *J Biomed Mater Res* 44: 206-216.
27. Wagner VE, Bryers JD (2004) Poly(ethylene glycol)-polyacrylate copolymers modified to control adherent monocyte-macrophage physiology: interactions with attaching *Staphylococcus epidermidis* or *Pseudomonas aeruginosa* bacteria. *J Biomed Mater Res A* 69: 79-90.
28. Roosjen A, Busscher HJ, Norde W, Van der Mei HC (2006) Bacterial factors influencing adhesion of *Pseudomonas aeruginosa* strains to a poly(ethylene oxide) brush. *Microbiology* 152: 2673-2682.
29. Godek ML, Michel R, Chamberlain LM, Castner DG, Grainger DW (2009) Adsorbed serum albumin is permissive to macrophage attachment to perfluorocarbon polymer surfaces in culture. *J Biomed Mater Res A* 88: 503-519.
30. Leid JG (2009) Bacterial biofilms resist key host defenses. *Microbe* 4: 66-70.
31. Kaplan SS, Basford RE, Mora E, Jeong MH, Simmons RL (1992) Biomaterial-induced alterations of neutrophil superoxide production. *J Biomed Mater Res* 26: 1039-1051.

SUPPORTING INFORMATION

Time-lapse videos of J774A.1 macrophages migrating towards *Staphylococcus epidermidis* 3399 and their phagocytic activity on glass and OptiChem® coated glass are shown in Supporting Information on this journal's electronic publication resources page on their website:

<http://www.ecmjournal.org/journal/papers/vol021/vol021a06.php>.

