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## CLINICAL IMPLICATIONS OF MOLECULAR IMAGING RESEARCH

# Current and Emerging Radiotracers in Molecular Cardiovascular Imaging

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**ABSTRACT:** Cardiovascular imaging has rapidly advanced over the past decades. Traditional imaging techniques such as echocardiography, computed tomography, and cardiovascular magnetic resonance are essential for assessing the structural and functional aspects of the cardiovascular system but often fall short in providing direct insights into disease activity. This gap is increasingly being bridged by molecular nuclear imaging techniques, including positron emission tomography and single-photon emission computed tomography, which enable the visualization of disease processes at the molecular and cellular levels. This review highlights the role of cardiovascular molecular imaging, emphasizing its current and potential applications in diagnosing and managing cardiovascular disease. With advancements in positron emission tomography scanners, novel radiotracers, and sophisticated imaging software, molecular imaging is set to play an essential role in precision medicine by enhancing our understanding of disease mechanisms, accelerating the development of targeted therapies, and facilitating personalized patient care.

**Key Words:** cardiovascular diseases ■ myocardial perfusion imaging ■ precision medicine ■ radiopharmaceuticals ■ sarcoidosis

Cardiovascular imaging has evolved at an accelerated pace over the past 2 decades adding a range of advanced imaging modalities to our armamentarium of investigational tools. In particular, noninvasive imaging has solved many of the issues around motion correction and spatial resolution, and imaging modalities, such as computed tomography (CT) and cardiovascular magnetic resonance (CMR), are now performed as part of routine care across the world. Similar approaches can be applied to molecular nuclear imaging techniques. They too are now being used to investigate patients in clinical practice, with a range of exciting research approaches on the horizon that might further contribute to patient care.

Traditional imaging techniques such as echocardiography, CT, and CMR are invaluable for assessing structural and functional aspects of the cardiovascular system but provide limited and indirect information about disease activity. This diagnostic gap is bridged by molecular

imaging techniques including positron emission tomography (PET) and single-photon emission computed tomography (SPECT) that can measure disease activity as it is occurring in the body. This article will review molecular nuclear imaging in humans, outlining the most promising techniques and exploring their current and potential applications in patients with cardiovascular disease (Figure 1). The focus will be on the application of these molecular imaging techniques to study disease activity while acknowledging that the vast majority of current clinical PET and SPECT imaging focuses on myocardial perfusion imaging. The latter will not be discussed further here.

## CARDIOVASCULAR MOLECULAR IMAGING

Molecular imaging involves the visualization, characterization, and measurement of biological processes at the

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## Nonstandard Abbreviations and Acronyms

<b>[<sup>11</sup>C]mHED</b>	carbon-11 meta-hydroxyephedrine
<b>[<sup>123</sup>I]-MIBG</b>	[ <sup>123</sup> I]-meta-iodobenzylguanidine
<b>[<sup>18</sup>F]FDG</b>	<sup>18</sup> F-fluorodeoxyglucose
<b>CMR</b>	cardiovascular magnetic resonance
<b>CT</b>	computed tomography
<b>CXCR4</b>	chemokine receptor type 4
<b>DOTATATE</b>	DOTA-D-Phe-Tyr3-octreotate
<b>FAP</b>	fibroblast activation protein
<b>FAPI</b>	fibroblast activation protein-specific inhibitor
<b>Na[<sup>18</sup>F]F</b>	<sup>18</sup> F-sodium fluoride
<b>PAREPET</b>	Prediction of Arrhythmic Events With Positron Emission Tomography
<b>PET</b>	positron emission tomography
<b>PIB</b>	Pittsburgh compound B
<b>SPECT</b>	single-photon emission computed tomography
<b>SSTR</b>	somatostatin receptor
<b>TSPO</b>	translocator protein

molecular and cellular levels. This technique provides crucial insights into the cellular and molecular mechanisms underlying various cardiovascular conditions. In today's era of precision medicine, molecular imaging is anticipated to play an increasingly vital role in establishing clinical diagnoses, distinguishing between active and inactive disease states, accelerating the development of novel drug therapies, and facilitating a personalized approach where the right medication is targeted to the right patient at the right time and the correct dose. Indeed, molecular imaging has already become an integral part of routine clinical cardiovascular practice. For example, <sup>18</sup>F-fluorodeoxyglucose ([<sup>18</sup>F]FDG) PET imaging is recommended by international guidelines for the evaluation of different cardiovascular disease stages, such as the assessment of myocardial viability, in patients with suspected endocarditis, cardiac sarcoidosis, and large vessel vasculitis.<sup>1–3</sup> However, with the advancements in PET scanners, image analysis software, and the development of novel radiopharmaceuticals, these advanced imaging techniques have the potential to play an even more significant in both clinical and research settings.

An ideal radiotracer should exhibit high specificity for its molecular target of interest (eg, an enzyme, receptor, or transport protein) that is linked to a specific cell population or disease state with minimal uptake in other tissues. The tracer should also be stable, resisting enzymatic degradation and maintaining integrity during imaging. Other desirable qualities include fast uptake into the tissues of interest and favorable kinetics such as rapid clearance to ensure a high signal-to-noise ratio.<sup>4</sup> Several

novel tracers display excellent imaging properties, allowing the in vivo assessment of many of the key pathological processes involved in cardiovascular diseases, including inflammation, infection, calcification, fibrosis, and thrombosis. For widespread clinical adoption, these imaging tests will likely need to be clearly associated with specific changes in patient management.

## GLUCOSE METABOLISM/[<sup>18</sup>F]FDG

### Myocardial Hibernation

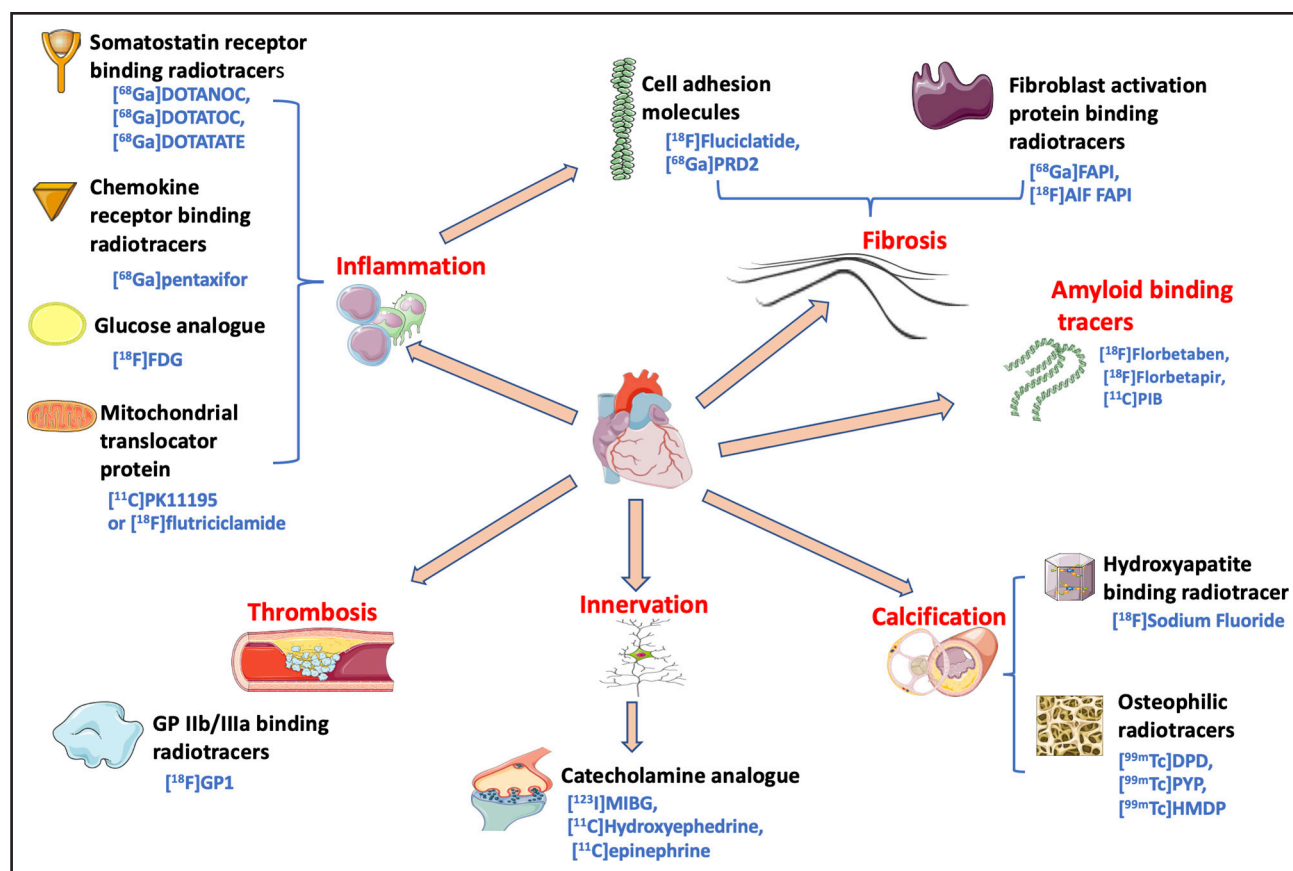
One of the most common indications for molecular imaging in clinical practice is the assessment of myocardial hibernation. PET is considered the gold standard for myocardial hibernation assessments and is based on the glucose analog [<sup>18</sup>F]FDG. This tracer is transported into viable myocytes by GLUT1 and GLUT4 transporters, where it is subsequently phosphorylated to [<sup>18</sup>F]FDG-6-phosphate by hexokinase,<sup>5</sup> which cannot undergo further metabolism. The tracer is, thereby, trapped within viable myocardial cells. Myocardial cells exposed to repeated or chronic ischemia switch their metabolism from lipids to sugar consumption. [<sup>18</sup>F]FDG is, therefore, particularly well suited to identify hibernating myocardium. A mismatch in perfusion-metabolism data denotes hibernating myocardium with a high likelihood of functional recovery following revascularization. The extent of perfusion-metabolism mismatch on PET is a good predictor of myocardial recovery after coronary revascularization.<sup>6</sup> While the STICH [Surgical Treatment for Ischemic Heart Failure] trial questioned the role of viability imaging, an important limitation is that it did not include [<sup>18</sup>F]FDG PET assessments to quantify the extent of hibernating myocardium. The PARR2 [Positron Emission Tomography and Recovery Following Revascularization] trial demonstrated significant benefits in patients where PET recommendations were adhered to and in patients without recent angiography.<sup>7</sup> More studies, and in specific randomized clinical trials, are now required to further test the hypothesis that [<sup>18</sup>F]FDG PET viability imaging improves clinical outcomes.

### Inflammation and Infection

Inflammation is a key pathological process underlying many cardiac conditions, including atherosclerosis, large vessel vasculitis, myocarditis, sarcoidosis, and infective conditions such as endocarditis. Inflammation is, therefore, a key target for molecular imaging with a range of different tracers having been developed for that purpose.

## <sup>18</sup>F-FLUORODEOXYGLUCOSE

[<sup>18</sup>F]FDG PET is also the most well-established radiotracer to image cardiovascular inflammation on the basis that activated macrophages and other inflammatory



**Figure 1. Established radiotracers for clinical use targeting key pathological processes underpinning cardiovascular diseases, including inflammation, fibrosis, amyloid deposits, calcification, innervation, and thrombosis.**

$[^{99\text{m}}\text{Tc}]$  indicates  $^{99\text{m}}\text{technetium}$ ;  $^{125}\text{I}$ ,  $^{125}\text{iodine}$ ;  $^{18}\text{F}$ ,  $^{18}\text{fluorine}$ ;  $^{68}\text{Ga}$ ,  $^{68}\text{gallium}$ ; DPD, 3,3-diphosphono-1,2-propanodicarboxylic acid; FAPI, fibroblast activation protein-specific inhibitor; FDG, fluorodeoxyglucose; GP1, glycoprotein 1; HMDP, hydroxymethylene diphosphonate; MIBG, meta-iodobenzylguanidine; PIB, Pittsburgh compound B; and PYP, pyrophosphate.

cells have higher glucose requirements than surrounding cardiovascular tissue. While, in viability assessments, physiological  $[^{18}\text{F}]\text{FDG}$  uptake by viable myocytes is encouraged, in studies of inflammation, it should be suppressed so that signals related to inflammation can be more readily visualized.

## Endocarditis

Despite advances in clinical investigations, the diagnosis of prosthetic valve endocarditis remains challenging. Echocardiography and blood cultures are inconclusive in 14% to 30% of episodes, especially in the early stages of disease.<sup>8</sup>  $[^{18}\text{F}]\text{FDG}$  PET-CT can detect the increased metabolic activity due to immune cell accumulation at sites of prosthetic valve infection<sup>1</sup> even in the absence of structural changes such as vegetation or valve incompetence. Overall,  $[^{18}\text{F}]\text{FDG}$  PET has a sensitivity of 81% to 91% and a specificity of 73% to 95% for the detection of prosthetic valve endocarditis and provides improved diagnostic certainty and detection of extracardiac sites of infection.<sup>1</sup> This technique has an established clinical role in the diagnosis of implantable device infections with

a sensitivity of 89% and specificity of 86%<sup>9</sup> and to guide decisions about the need for device removal, often a complex and high-risk procedure. However, evidence for  $[^{18}\text{F}]\text{FDG}$  PET in native valve endocarditis is more limited, demonstrating lower sensitivity compared with prosthetic valve endocarditis.<sup>10</sup> More specific molecular imaging probes to image native valve endocarditis are currently being developed and assessed (Table S1).

## Atherosclerosis and Vasculitis

$[^{18}\text{F}]\text{FDG}$  PET imaging has been widely researched in atherosclerosis as a marker of vascular inflammation, particularly in the aorta and carotid arteries.  $[^{18}\text{F}]\text{FDG}$  uptake is associated with plaque macrophage accumulation and hypoxic plaque conditions.<sup>11</sup> Its ability to image vulnerable plaques in humans was first demonstrated >20 years ago with higher uptake of  $[^{18}\text{F}]\text{FDG}$  in patients with symptomatic carotid plaques, as opposed to the asymptomatic contralateral side and healthy arteries.<sup>12</sup>  $[^{18}\text{F}]\text{FDG}$  has since been validated as a surrogate marker of plaque macrophage content and hypoxia in humans by several independent studies and in different vascular

beds.<sup>13</sup> [<sup>18</sup>F]FDG PET has been used as an end point in trials assessing the efficacy of drugs targeting atherosclerotic inflammation in the carotid arteries and ascending thoracic aorta with the results mirroring those of large clinical end point studies.<sup>14</sup> Further work is required to assess whether [<sup>18</sup>F]FDG PET imaging can improve patient risk stratification and can help direct the use of novel anti-inflammatory agents. [<sup>18</sup>F]FDG PET imaging is also being increasingly used to aid the diagnosis and management of rarer aortic conditions such as large vessel vasculitis and prosthetic vascular graft infections.<sup>15</sup>

## Sarcoidosis

Sarcoidosis is a multisystem granulomatous disease of unknown cause that can affect any organ system including the heart. Cardiac involvement is characterized by focal regions of inflammation (noncaseating granulomas) that is often asymptomatic but can also lead to aortic valve conduction disease, ventricular arrhythmias, heart failure, and sudden cardiac death. Indeed, cardiac sarcoidosis accounts for 13% to 25% of sarcoid-related deaths.<sup>16</sup> Accurate diagnosis of cardiac involvement is important to initiate appropriate treatment and prevent complications due to the sarcoid disease process. The diagnosis of cardiac sarcoidosis remains challenging. Isolated cardiac sarcoidosis can occur, and echocardiography has poor sensitivity, meaning that it cannot be used to rule out the disease. Guidelines, therefore, recommend [<sup>18</sup>F]FDG alongside echocardiography and CMR in the diagnostic pathway.<sup>17</sup> Increased myocardial [<sup>18</sup>F]FDG uptake is observed in regions of active myocardial sarcoidosis and can be used to direct and monitor the use of anti-inflammatory therapies (Figure 2).<sup>18,19</sup> A meta-analysis of 7 studies involving 164 patients, assessing [<sup>18</sup>F]FDG PET in the diagnosis of cardiac sarcoidosis, reported 89% sensitivity and 78% specificity,<sup>20</sup> with other studies also demonstrating the powerful prognostic information provided by this approach and its ability to track response to therapy.<sup>21</sup> However, robust quantitative analysis methods and specific imaging phenotypes predictive of adverse outcomes are currently lacking and would enable a better understanding of long-term outcomes and the selection of patients who may benefit from early aggressive strategies. Investigation of more specific inflammatory tracers is also required.

## Limitations of [<sup>18</sup>F]FDG

Although now widely used in clinical practice, [<sup>18</sup>F]FDG has important limitations as a radiotracer for cardiovascular applications. First and foremost, it is a glucose analog that lacks specificity, as most human cells metabolize glucose for ATP synthesis. Physiological FDG uptake, therefore, occurs throughout the body including the myocardium. This can hamper

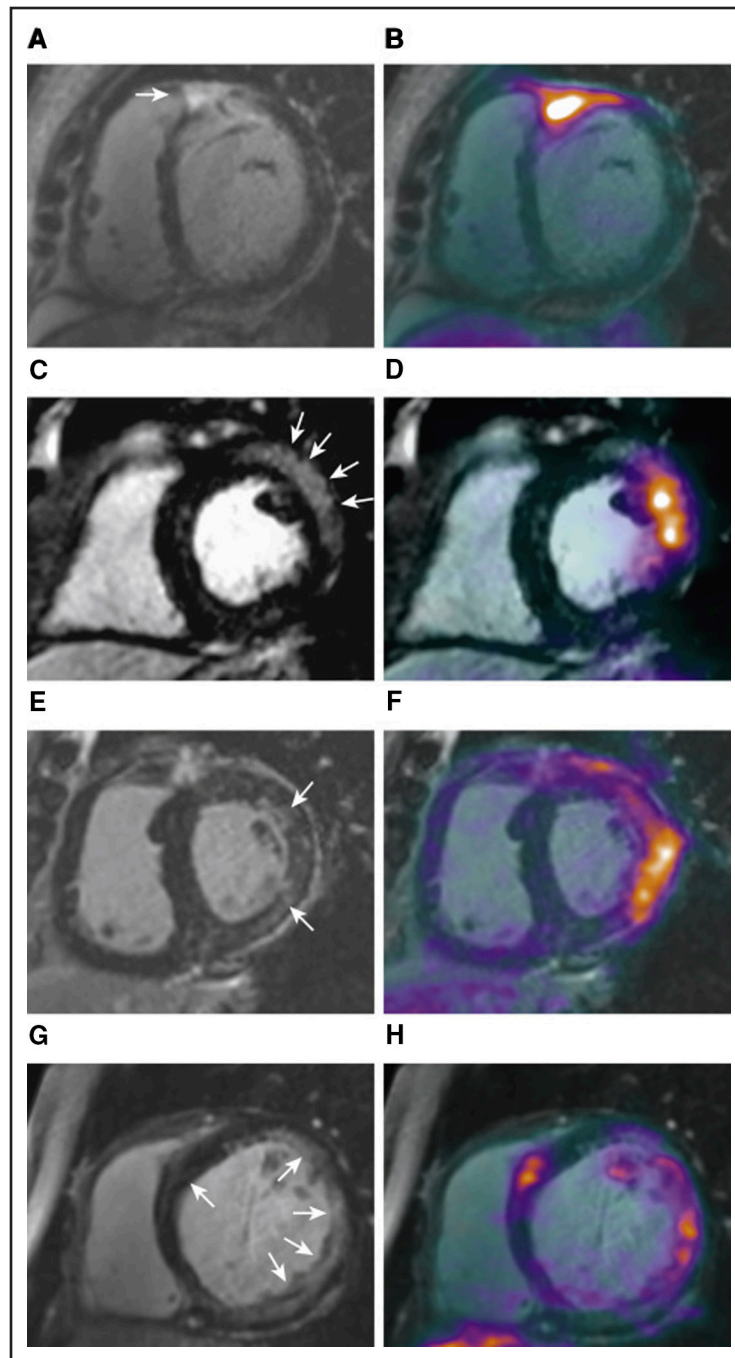
the interpretation of cardiac inflammation, with high-intensity physiological [<sup>18</sup>F]FDG uptake obscuring any underlying inflammatory signal. While physiological [<sup>18</sup>F]FDG uptake can be suppressed by dietary restrictions (eg, a 12-hour fast with avoidance of carbohydrates for 24 hours) and intravenous heparin administration, these measures are not always successful with diffuse or patchy myocardial [<sup>18</sup>F]FDG uptake remaining, potentially causing diagnostic difficulties.<sup>22</sup> Second, noninfected prosthetic heart valves frequently display homogenous [<sup>18</sup>F]FDG uptake in the perivalvular area on PET/CT, particularly in the early stages following surgery, and can demonstrate high-intensity uptake when different forms of surgical glue are used as part of the operative strategy.<sup>9</sup> Interpretation of [<sup>18</sup>F]FDG PET/CT in suspected prosthetic valve endocarditis can, therefore, be challenging with similar issues also arising in suspected cardiac device infections. On this basis, there is a major interest in developing more specific tracers for infection imaging.

## SOMATOSTATIN RECEPTOR RADIOTRACERS

Activated inflammatory cells have abundant SSTRs (somatostatin receptors) on their surface compared with normal cardiomyocytes. As such, there is little background uptake of somatostatin receptor radiotracers such as <sup>68</sup>Ga-DOTA-Nal-octreotide, <sup>68</sup>Ga-DOTA-D-Phe-Tyr3-octreotate (DOTATATE), and <sup>68</sup>Ga-DOTA-D-Phe-Tyr-octreotide in the heart.<sup>23</sup> SSTR-targeted radiotracers have now been investigated in myocarditis, sarcoidosis, coronary atherosclerosis, and large vessel vasculitis.

Increased <sup>68</sup>Ga-DOTA-D-Phe-Tyr-octreotide and [<sup>68</sup>Ga]DOTATATE activity is seen in patients with acute pericarditis/myocarditis or acute myocardial infarction, demonstrating good agreement with the pattern of injury on CMR.<sup>24</sup> [<sup>68</sup>Ga]DOTATATE PET has also been validated as a marker of atherosclerotic inflammation, with increased [<sup>68</sup>Ga]DOTATATE activity observed in culprit coronary and carotid plaques following acute myocardial infarction and stroke, respectively, with more favorable imaging characteristics compared with [<sup>18</sup>F]FDG.<sup>25</sup> Similarly, higher [<sup>64</sup>Cu]DOTATATE uptake is observed in symptomatic versus contralateral carotid plaques ( $P<0.001$ ) in patients with stroke, while aortic [<sup>68</sup>Ga]DOTATATE uptake correlates with the calcific plaque burden and [<sup>18</sup>F]FDG activity ( $r=0.64$ ;  $P<0.01$ ).<sup>26</sup> Finally, few studies have investigated the utility of SSTR-PET-CT imaging to diagnose cardiac sarcoid, again reporting good concordance with CMR.<sup>27</sup> Similarly, <sup>68</sup>Ga-DOTA-D-Phe-Tyr-octreotide can identify active granulomatosis and help with treatment evaluation in cardiac sarcoidosis.<sup>27</sup>





**Figure 2. Positron emission tomography (PET)/magnetic resonance (MR) imaging in cardiac sarcoidosis.**

(A) Subepicardial (near transmural) late gadolinium enhancement (LGE) in the basal anteroseptum extending in to the right ventricular free wall with increased fluorodeoxyglucose (FDG) uptake localizing to exactly the same region on fused CMR/PET (maximum standardized uptake value = 3.4; maximum tissue-to-background ratio = 2.3; maximum target-to-normal myocardium ratio = 2.0). (B) Subepicardial LGE in the basal anterolateral wall with increased FDG uptake co-localizing to exactly that region on CMR/PET. (C) Patchy midwall LGE in the anterolateral wall with matched increased FDG uptake on CMR/PET. (D) Multifocal LGE in the lateral wall with matched increased FDG uptake on CMR/PET. (E) Patchy midwall LGE in the anterolateral wall with (F) matched increased FDG uptake on CMR/PET. (G) Multifocal LGE in the lateral wall with (H) matched increased FDG uptake on CMR/PET. MR and PET images from 4 patients with active cardiac sarcoidosis in whom characteristic patterns of myocardial late gadolinium enhancement (left column) colocalize with increased  $^{18}\text{F}$ fluorine fluorodeoxyglucose uptake (fused images, right column). Reprinted from Dweck et al<sup>19</sup> with permission.

## CHEMOKINE RECEPTORS

Chemokines are involved in the recruitment of leucocytes, and CXCR4 (chemokine receptor type 4) is expressed on

several proinflammatory immune cell types, in particular, macrophages and T lymphocytes.<sup>28</sup> Therefore, CXCR4-directed PET tracers such as [ $^{68}\text{Ga}$ ]pentixafor are used to image inflammation in clinical research.

Increased CXCR4 expression and [<sup>68</sup>Ga]pentixafor uptake are observed in the infarct zone in patients with acute myocardial infarction and negatively correlate with scar volume and predict cardiac outcomes.<sup>29</sup> [<sup>68</sup>Ga]pentixafor has also been used to detect coronary inflammation in patients with acute myocardial infarction with increased [<sup>68</sup>Ga]pentixafor activity localizing to culprit lesions and outperforming the results of [<sup>18</sup>F]FDG.<sup>30</sup> Imaging-based guidance to optimize therapeutic timing of CXCR4 inhibition resulted in improved functional outcomes in mice,<sup>31</sup> but, although promising, clinical studies proving this concept are lacking and now required.

Recent studies have also shown the importance of CCR2-positive monocytes and macrophages as mediators of adverse remodeling in myocardial infarction, inflammation after heart transplantation,<sup>32</sup> and leukocyte trafficking to the site of arterial inflammation in abdominal aortic aneurysm.<sup>33</sup> However, previous work on CCR2 has mainly been limited to animal models, with clinical evaluation in patients now warranted.

## MITOCHONDRIAL TRANSLOCATOR PROTEIN

Mitochondrial TSPO (translocator protein) is a key outer mitochondrial membrane protein that is expressed by activated mononuclear cells and other inflammatory cells. The abundant expression of TSPO in macrophages has been utilized to image the immune response of the heart to inflammatory conditions such as myocarditis. Several PET radioligands for TSPO have been described including [<sup>11</sup>C]PK11195 and [<sup>18</sup>F]flutriclamide (<sup>18</sup>F-GE180). Increased [<sup>11</sup>C]PK11195 activity is observed in the culprit carotid plaques of patients with recent stroke.<sup>34</sup> Similarly, increased [<sup>11</sup>C]PK11195 uptake has also been demonstrated in patients with large vessel vasculitis, correlating with other markers of inflammation and demonstrating increased activity in symptomatic versus asymptomatic patients.<sup>35</sup>

Despite promising results from initial studies with TSPO radiotracers, these radiotracers have some major limitations including high nonspecific uptake and wide variability in TSPO binding caused by a single nucleotide polymorphism in the TSPO gene, which leads to patients with high, mixed, and low affinity binding potential.<sup>36</sup> Interpretation of TSPO activity, therefore, requires genetic assessment for this polymorphism, greatly increasing the complexity and expenses of TSPO imaging. A new selective PET radiotracer for imaging TSPO, [<sup>18</sup>F]LW223 appears to overcome this issue although human studies are lacking.<sup>37</sup>

## INFECTION-SPECIFIC RADIOPHARMACEUTICALS

While the radiotracers discussed above are useful for detecting inflammation, there is still a need for

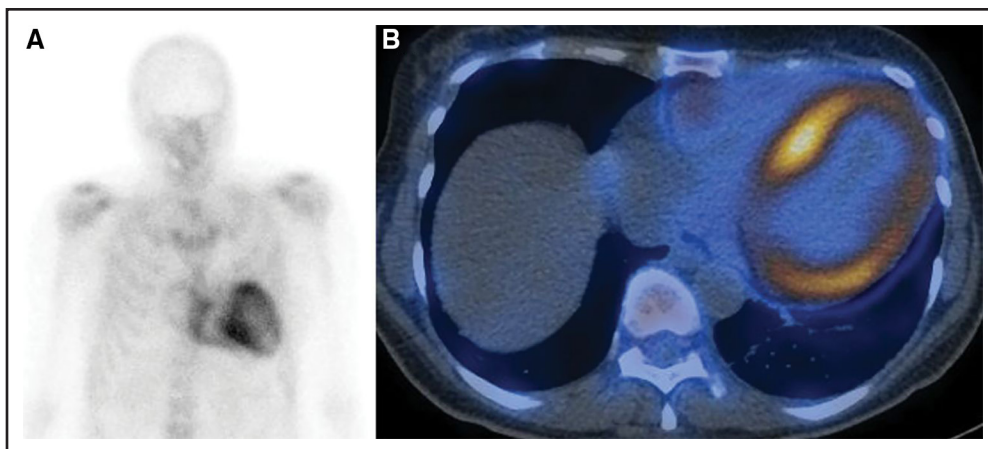
radioligands that can distinguish between infection and sterile inflammation. Examples of novel infection-specific radiotracers include <sup>89</sup>Zr-labelled anti-LTA mAb SAC55, which consists of a monoclonal antibody against *Staphylococcus aureus* labeled with zirconium-89, [<sup>68</sup>Ga]NOTA-ubiquitin, which consists of chelated ubiquitin (an antimicrobial peptide) labeled with gallium-68, and 2-<sup>18</sup>F-fluorodeoxysorbitol, which consists of sorbitol (a metabolic substrate for gram-negative bacteria) labeled with fluorine-18.<sup>38</sup> These radiotracers have shown promising results in infective conditions such as prosthetic joint and lung infections although they have not yet been tested in cardiovascular infections and their utility may depend on prior identification of the causative organism.

## Calcification

Calcification is a key healing response to cardiovascular injury and is a common pathological feature in atherosclerosis, valvular heart disease, and peripheral vascular disease. Assessing calcification activity provides a readout for vascular injury and disease activity across multiple disease states. Nuclear molecular imaging allows assessment of calcification activity with both SPECT and PET providing complementary information to the presence of established macroscopic calcification detected by CT.<sup>39</sup>

## BISPHOSPHONATE SPECT TRACERS

Transthyretin cardiac amyloidosis is a progressive, potentially fatal, infiltrative cardiomyopathy caused by extracellular deposition of transthyretin-derived insoluble amyloid fibrils in the myocardium. Transthyretin cardiac amyloidosis is more common than previously thought and treatable,<sup>40</sup> with early diagnosis and distinction from amyloid light chain cardiac amyloidosis key to improving patient outcomes. Bisphosphonate tracers including <sup>99m</sup>technetium 3,3-diphosphono-1,2-propanodicarboxylic acid, <sup>99m</sup>technetium pyrophosphate, and <sup>99m</sup>technetium hydroxymethylene diphosphonate demonstrate increased myocardial uptake in transthyretin cardiac amyloidosis with high sensitivity and specificity.<sup>41</sup> The value of bone scintigraphy in the diagnosis of cardiac amyloidosis was first demonstrated in the 1980s. Subsequently, the Perugini grading system was developed based on a simple 4-point visual scoring system of delayed (3-hour) planar images (grade 0, no cardiac uptake and normal bone uptake; grade 1, cardiac uptake that is less intense than bone signal; grade 2, cardiac uptake with intensity similar or greater than bone signal; and grade 3, cardiac uptake greater than bone uptake with much attenuated or absent bone signal; Figure 3).<sup>42,43</sup> Gillmore et al<sup>44</sup> analyzed bone scintigraphy and biochemical investigations from 1217 patients with suspected cardiac amyloidosis and confirmed that Perugini grade  $\geq 2$  myocardial



**Figure 3. Case example of  $^{99m}\text{Tc}$  3,3-diphosphono-1,2-propanodicarboxylic acid scintigraphy and single-photon emission computed tomography.**

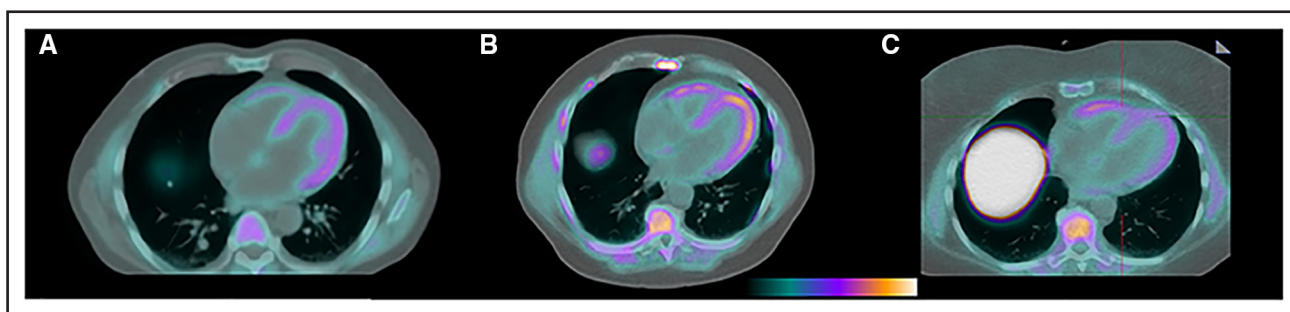
Examples of a whole-body anterior  $^{99m}\text{Tc}$  3,3-diphosphono-1,2-propanodicarboxylic acid scintigraphy (**A**) and single-photon emission computed tomography (**B**) showing abnormal uptake in a patient with cardiac transthyretin type amyloidosis consistent with Perugini grade 2. Reprinted from Martinez-Naharro et al<sup>43</sup> with permission. Copyright ©2018, Elsevier (the article is available under the Creative Commons CC-BY-NC-ND license).

uptake on bone scintigraphy in absence of monoclonal gammopathy was >99% sensitive and 86% specific for cardiac transthyretin amyloidosis amyloid. Several other studies have confirmed similar findings, and this combination is, therefore, sufficient to establish a diagnosis of transthyretin cardiac amyloidosis, without the need for endomyocardial biopsy.<sup>41</sup> The use of advanced SPECT/CT cameras can further increase the sensitivity of the exam.

Other PET radiopharmaceuticals that bind amyloid deposits directly include [ $^{11}\text{C}$ ] Pittsburgh compound B (PIB), [ $^{18}\text{F}$ ]florbetapir (Figure 4),<sup>45</sup> and [ $^{18}\text{F}$ ]florbetaben. These have demonstrated promising results in clinical research studies as recently described in the expert consensus recommendations for multimodality imaging in cardiac amyloidosis.<sup>46</sup> [ $^{18}\text{F}$ ]florbetapir and [ $^{18}\text{F}$ ]florbetaben were able to distinguish amyloid light chain cardiac amyloidosis from transthyretin amyloidosis cardiac

amyloid or other mimicking conditions in clinical studies.<sup>47</sup> PET imaging can potentially provide incremental information on cardiac amyloid. In particular, it may play a role in the risk stratification of patients with cardiac amyloidosis, and several studies demonstrated poorer outcomes in patients with higher [ $^{11}\text{C}$ ]PIB accumulation in the myocardium.<sup>48</sup> Furthermore, to improve clinical outcomes, PET tracers have the potential to allow for early detection of preclinical amyloid deposits, which is lacking with current technologies. A dual-center study that examined PIB PET in a large cohort of patients with amyloidosis and controls found a high prevalence of PIB uptake among patients with amyloidosis without increased myocardial wall thickness, indicating the potential ability of PIB PET to detect early stages of cardiac amyloidosis.<sup>49</sup>

With currently available precursor protein-directed therapies, there is increasing interest in imaging methods



**Figure 4. Cardiac [ $^{18}\text{F}$ ]florbetapir uptake in patients with and without cardiac amyloidosis.**

**A**, [ $^{18}\text{F}$ ]florbetapir cardiac positron emission tomography/computed tomography images demonstrating identifiable myocardial uptake in a patient with cardiac amyloidosis confirmed by technetium pyrophosphate (mean myocardial SUV, 3.3), **(B)** a patient with light chain MGUS and potential for having cardiac light chain amyloidosis (mean myocardial SUV, 2.7), and **(C)** control patient with negative tenosynovial biopsy for amyloidosis at time of carpal tunnel release surgery (mean myocardial SUV, 2.0). Reprinted from Sperry et al<sup>45</sup> with permission. Copyright ©2021, Frontiers Media SA (the article is available under the Creative Commons CC-BY-NC-ND license). MGUS indicates monoclonal gammopathy of unknown significance; and SUV, standardised uptake values.



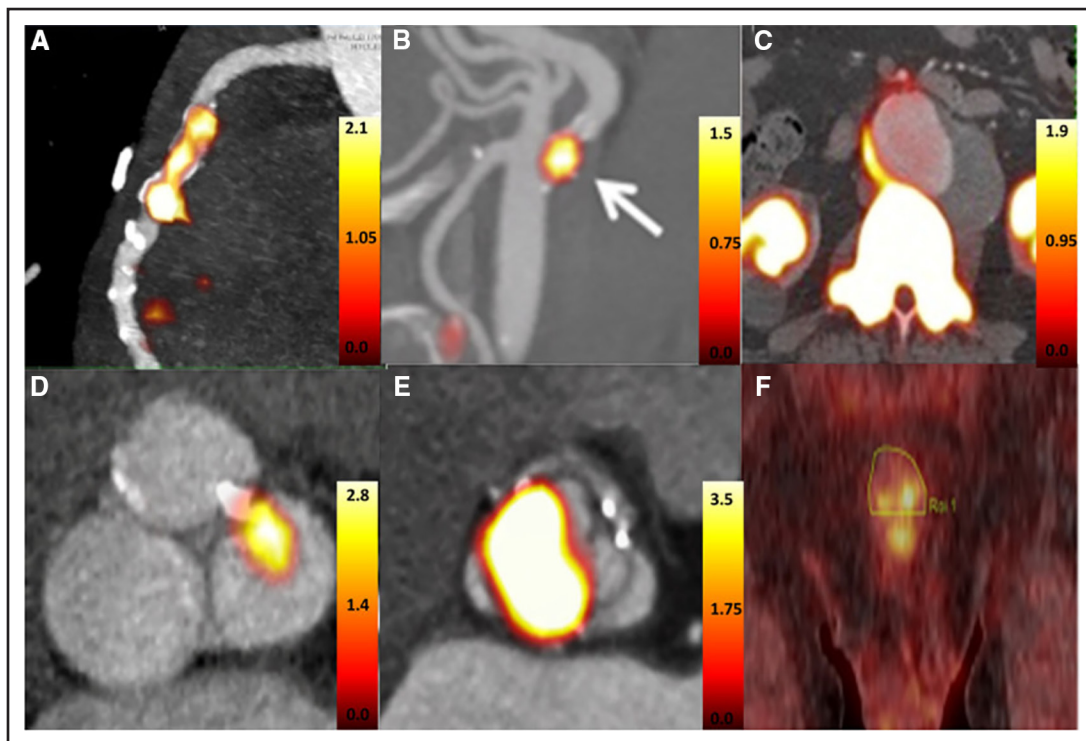
to accurately evaluate and monitor treatment response. Preliminary data showed a reduction in cardiac tracer uptake versus whole-body uptake on planar scintigraphy as early as 4 months after treatment initiation with the recombinant human anti-transthyretin amyloidosis monoclonal antibody NI006.<sup>50</sup> However, visual identification of changes on serial amyloid scans is extremely challenging, and more research on quantitative PET imaging with kinetic modeling methods to allow accurate monitoring of therapy and early identification of nonresponders is needed.<sup>51</sup> The ongoing I-CARE study is investigating whether <sup>18</sup>F-sodium fluoride (Na[<sup>18</sup>F]F) PET imaging can be used to track treatment responses in TTR cardiac amyloidosis and whether this offers incremental information to CMR approaches.

### <sup>18</sup>F-SODIUM FLUORIDE

Na[<sup>18</sup>F]F is an established bone PET tracer that binds to hydroxyapatite, a key crystalline component of both bone and vascular calcification. The mechanism of Na[<sup>18</sup>F]F uptake in bone is well established. It diffuses via the capillary network into the bone extracellular fluid and then exchanges with hydroxyl groups on exposed regions of hydroxyapatite crystals on the surface of the calcium deposit to form fluorapatite. The intensity of Na[<sup>18</sup>F]F

uptake depends on the surface area of exposed hydroxyapatite available for binding, which is much higher in regions of newly developing powdery microcalcification than in areas of established macrocalcification (the type of calcium imaged with CT) where most of the hydroxyapatite signal is internalized and not available for binding. Na[<sup>18</sup>F]F PET has now been investigated across multiple different cardiovascular disease states, where it consistently provides an assessment of calcification activity and vascular injury, detecting newly developing microcalcification before it is visible on CT and predicting disease progression and clinical events (Figure 5).<sup>52</sup>

Coronary Na[<sup>18</sup>F]F uptake as a novel marker of calcification activity was first described in 2012. Participants with increased Na[<sup>18</sup>F]F activity had higher rates of prior cardiovascular events ( $P=0.016$ ), angina ( $P=0.023$ ), and higher Framingham risk scores ( $P=0.011$ ).<sup>53</sup> Subsequent studies demonstrated that intense Na[<sup>18</sup>F]F uptake localizes to the culprit coronary vessel and areas of recent plaque rupture in patients with acute myocardial infarction. In patients with stable disease, it localizes to plaques with multiple high-risk features on intravascular ultrasound and a range of other adverse plaque characteristics assessed by different techniques.<sup>54</sup> Doris et al<sup>55</sup> confirmed that Na[<sup>18</sup>F]F provides an assessment of disease activity in the coronary arteries, predicting



**Figure 5.** <sup>18</sup>Fluorine sodium fluoride (<sup>18</sup>F]NaF) uptake in different disease states.

**A**, Patient with recent myocardial infarction, **(B)** patient with recent transient ischemic attack and uptake in the culprit carotid artery, **(C)** patient with abdominal aortic aneurysm, **(D)** patient with mild aortic valve stenosis and corresponding [<sup>18</sup>F]NaF uptake in areas of leaflet calcification, **(E)** patient with aortic valve degeneration and corresponding <sup>18</sup>F-NaF uptake, and **(F)** patient with erectile dysfunction and high penile uptake. Reprinted from Tzolos et al<sup>52</sup> with permission. Copyright ©2020, Wolters Kluwer Health (The article is available under the Creative Commons CC-BY-NC-ND license).

subsequent disease progression at both the plaque and patient levels. Indeed, patients without coronary Na[<sup>18</sup>F]F activity at baseline did not demonstrate subsequent progression in their coronary CT calcium score, whereas patients with coronary Na[<sup>18</sup>F]F uptake did, despite similar baseline plaque burdens.<sup>55</sup> Kwiecinski et al<sup>56</sup> demonstrated that coronary Na[<sup>18</sup>F]F uptake also predicts subsequent clinical events. In a cohort of patients with advanced established coronary artery disease, coronary Na[<sup>18</sup>F]F activity emerged as the strongest predictor of fatal or nonfatal MI outperforming cardiovascular risk factors and CT assessments of luminal stenosis and atherosclerotic plaque burden.<sup>56</sup> More recently, results from the prospective multicenter PRE<sup>18</sup>FFIR [Prediction of Recurrent Events with 18F-Fluoride to Identify Ruptured and High-risk Coronary Artery Plaques in Patients with Myocardial Infarction] trial showed that coronary atherosclerotic plaque activity assessed with Na[<sup>18</sup>F]F PET predicts spontaneous recurrent atherosclerotic events in a large cohort (n=704) of patients with established multi-vessel coronary artery disease.<sup>57</sup> Based on these findings, Na[<sup>18</sup>F]F PET might be used to guide the application of more intensive lipid-lowering, anti-inflammatory or other advanced therapies in this patient cohort who are already on standard secondary prevention treatments. Nevertheless, further work is required to investigate how coronary plaque disease activity should be used in clinical practice and how this information might ultimately improve clinical decision-making. This will require randomized controlled trials investigating whether targeting therapy using plaque activity imaging assessments improves patient outcomes in a cost-effective way.

Na[<sup>18</sup>F]F uptake has also been widely investigated in heart valve disease. In aortic stenosis, intense Na[<sup>18</sup>F]F uptake is observed within the valve, identifying where new macrocalcific deposits will develop on CT and predicting subsequent disease progression as assessed by CT calcium scoring and echocardiography.<sup>58</sup> While the clinical use of Na[<sup>18</sup>F]F is likely to be limited by CT calcium scoring, which provides similar predictive and prognostic information, Na[<sup>18</sup>F]F is used increasingly in the research arena, in particular, to investigate the factors associated with calcification activity in aortic stenosis and as an end point in clinical trials of novel therapies aiming to slow valve calcification.

Similar findings have been observed in patients with mitral annular calcification<sup>59</sup> and bioprosthetic valve degeneration. Marked Na[<sup>18</sup>F]F uptake is seen in explanted degenerated bioprosthetic valve tissue, and this correlated with valve degeneration on histology. In a prospective clinical cohort study of patients with surgically implanted bioprosthetic aortic valves, increased Na[<sup>18</sup>F]F PET uptake occurred in 34% of patients who did not have any clinical or echocardiographic suspicion of bioprosthetic valve degeneration. Baseline Na[<sup>18</sup>F]F PET went on to provide the most powerful prediction

of subsequent valve deterioration, outperforming all other known predictive markers.<sup>60</sup> These findings were subsequently confirmed in patients with transcatheter aortic valve implantation valves,<sup>61</sup> providing further support to Na[<sup>18</sup>F]F as an early marker of bioprosthetic valve degeneration. Further studies are required to evaluate how best to use this novel technique in routine clinical practice and how it might be used to complement the current strategy of echocardiographic surveillance. Other conditions where Na[<sup>18</sup>F]F PET has demonstrated exciting early results and where further research is ongoing include abdominal aortic aneurysmal disease, peripheral vascular disease, and carotid atherosclerosis.<sup>52</sup>

## Fibrosis

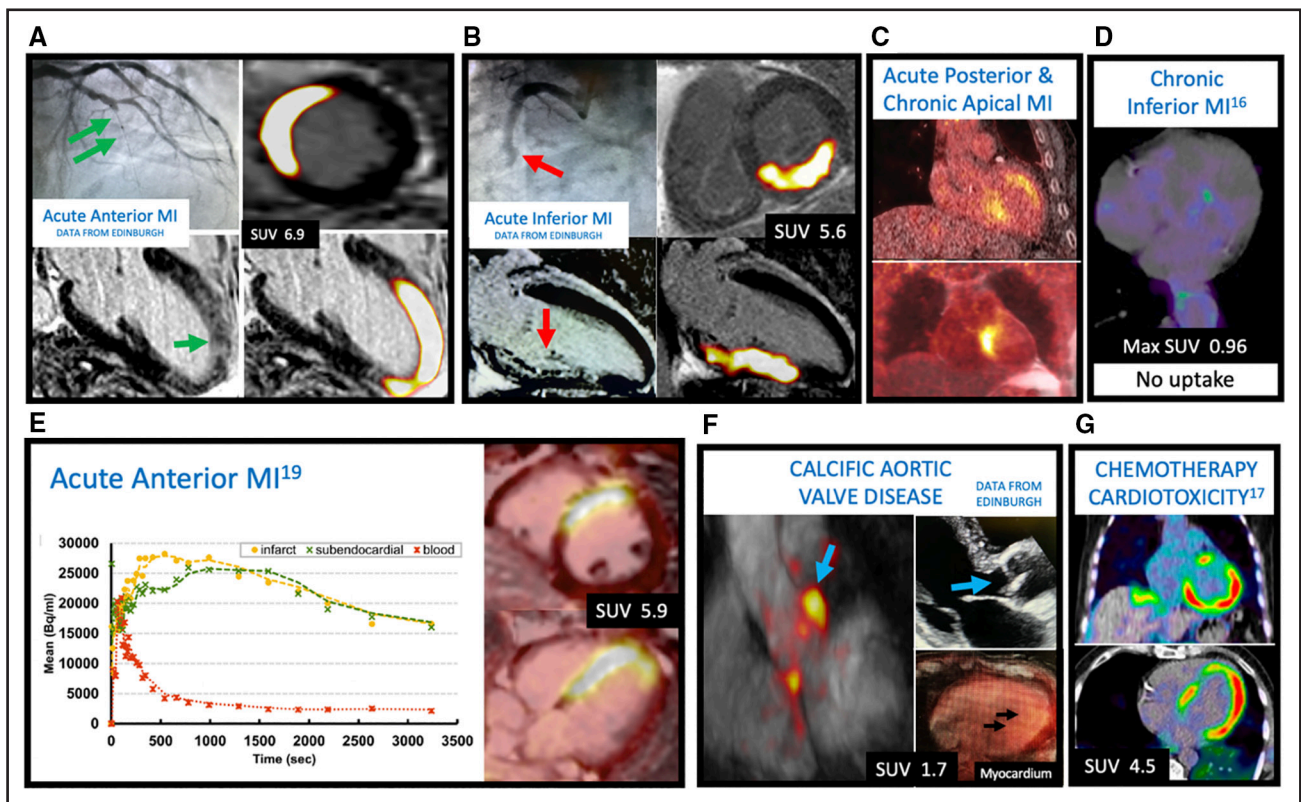
Fibrosis is a fundamental pathological process in cardiovascular disease and appears to be the common final pathway of injury in the myocardium, underlying almost every form of heart muscle disease. It plays a crucial role in cardiac remodeling that leads to heart failure, as well as ventricular dysrhythmia. Myocardial fibrosis takes 2 predominant forms: replacement and interstitial fibrosis. Replacement fibrosis is focal and nonreversible, often occurring as a result of myocyte necrosis, for example, following myocardial infarction. Interstitial fibrosis is diffuse and potentially reversible, characterizing conditions such as hypertension and valvular heart disease. Activated fibroblasts are the most prominent cells involved in myocardial fibrosis. Activation of fibroblasts occurs in response to myocardial injury under the influence of several pathways including the renin-angiotensin-aldosterone system, transforming growth factor-beta and interleukin-11.<sup>62</sup>

## RADIOTRACERS TARGETING FIBROBLAST ACTIVATION PROTEIN

FAP (fibroblast activation protein) is a cell surface proteinase that is almost exclusively expressed on activated fibroblasts, with no expression on myocytes, inflammatory cells, or quiescent fibroblasts.<sup>63</sup> FAP is an excellent imaging target to inform about fibroblast activation. Recently, FAP-specific inhibitors (FAPis) have been developed and radiolabeled with either gallium-68 or fluorine-18 aluminum fluoride. These PET tracers are highly specific for FAP, becoming internalized within the cell after receptor binding and demonstrating low levels of leakage thereafter. Combined with rapid renal clearance, [<sup>68</sup>Ga]-FAPi and Al[<sup>18</sup>F]F-FAPi have emerged as highly specific tracers for fibroblast activation across a range of different organ systems, demonstrating high-intensity signal in regions of activated fibroblasts and low background uptake, as well as low radiation doses.<sup>64</sup>

In the cardiovascular system, the first case report described increased myocardial  $[^{68}\text{Ga}]$ -FAPi uptake in a patient with treated pancreatic adenocarcinoma and chemotherapy-associated cardiotoxicity. Subsequent case reports have demonstrated increased myocardial  $[^{68}\text{Ga}]$ -FAPi uptake in patients with dilated cardiomyopathy, hypertensive heart disease, chemotherapy cardiotoxicity, and radiation heart disease.<sup>65</sup> Recently, a retrospective study described cardiac  $[^{68}\text{Ga}]$ -FAPi uptake in 229 oncology patients, undergoing PET-CT to evaluate metastatic cancer. Increased myocardial  $[^{68}\text{Ga}]$ -FAPi uptake was associated with cardiovascular risk factors including diabetes (odds ratio, 2.9;  $P=0.04$ ) and with previous platinum-based chemotherapy (odds ratio, 3.0;  $P=0.03$ ).  $[^{68}\text{Ga}]$ -FAPi uptake was also higher in patients with reduced versus preserved ejection fraction.<sup>66</sup> Several studies have now described intense  $[^{68}\text{Ga}]$ -FAPi uptake in patients following acute myocardial infarction with the PET signal extending beyond the infarct zone towards the infarct border zone and predictive of the evolution of ventricular dysfunction.<sup>67</sup> A recent study demonstrated

increased  $[^{68}\text{Ga}]$ -FAPi uptake in patients with hypertrophic cardiomyopathy, and this was association with cardiovascular events.<sup>65</sup> In patients with aortic stenosis planned for valve replacement,  $[^{68}\text{Ga}]$ -FAPi uptake was heterogeneous among individuals, but significantly higher than in matched controls, and correlated with left ventricular dysfunction.<sup>68</sup> While early in its development, there is considerable excitement about the investigation of activated fibroblasts with  $[^{68}\text{Ga}]$ -FAPi and  $[^{18}\text{F}]$ -FAPi across a range of cardiomyopathic conditions (Figure 6), which parallels the progression and understanding of cardiac fibrosis and allows for the development of novel targeted antifibrotic therapies. Indeed, not only molecular imaging with  $[^{68}\text{Ga}]$ -FAPi but also experimental approaches for the treatment of cardiac fibrosis by chimeric antigen receptor T cells targeted against FAP have seen rapid development,<sup>69</sup> with the recent introduction of a nanoparticle-based approach for transient in vivo FAP chimeric antigen receptor T-cell production to address the temporal dynamics and avoid long-term effects of fibrosis.<sup>70</sup>



**Figure 6. Data demonstrating increased  $[^{68}\text{gallium}]$  fibroblast activation protein-specific inhibitors ( $[^{68}\text{Ga}]$ FAPi) activity in different cardiovascular diseases.**

**A**, Patient with acute anterior myocardial infarction (MI) and increased myocardial  $[^{68}\text{Ga}]$ FAPi uptake in the anterior, anteroseptal, and apical regions. **B**, Patient with acute inferior MI demonstrating myocardial  $[^{68}\text{Ga}]$ FAPi uptake present in the inferior wall. **C**, Patient with acute posterior and chronic apical MI with increased myocardial  $[^{68}\text{Ga}]$ FAPi uptake present in the posterior wall but not in the apical region. **D**, Patient with chronic inferior MI with no evidence of myocardial  $[^{68}\text{Ga}]$ FAPi uptake. **E**, Patient with acute anterior MI demonstrating rapid myocardial  $[^{68}\text{Ga}]$ FAPi uptake in the infarct zone. **F**, Patient with aortic stenosis demonstrating increased  $[^{68}\text{Ga}]$ FAPi uptake in both the aortic valve leaflets and in the myocardium. **G**, Globally increased myocardial  $[^{68}\text{Ga}]$ FAPi uptake in patients with chemotherapy-induced cardiotoxicity (data from The University of Edinburgh). SUV indicates standardised uptake values.



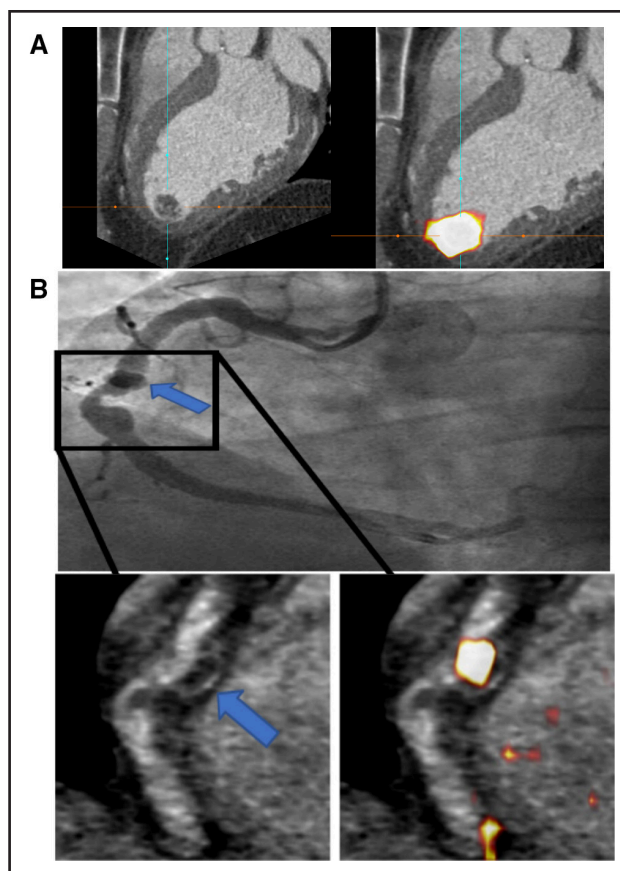
Although [ $^{68}\text{Ga}$ ]-FAPI PET is a promising tool for evaluating cardiovascular fibrosis, there are still gaps in knowledge and studies needing filling. Large-population and multicenter studies are warranted to evaluate the role of [ $^{68}\text{Ga}$ ]-FAPI PET imaging in the early detection and risk stratification of various cardiovascular diseases. Promising future research perspectives on [ $^{68}\text{Ga}$ ]-FAPI PET imaging could be the response to therapy and prediction of the evolution of left ventricle function, as there are merely a few related small-volume studies to date. The relationship between [ $^{68}\text{Ga}$ ]-FAPI PET imaging and other imaging modalities should be investigated to better understand the advantages and possible applications of [ $^{68}\text{Ga}$ ]-FAPI PET imaging. Moreover, given the current application of FAP inhibitors in cancer treatments,<sup>71</sup> FAPI PET imaging opens the door to new theranostic approaches to cardiovascular diseases.

A range of tracers targeting alternative pathways involved in tissue fibrosis have been investigated, but most work in this area is preclinical, and translation to humans is not as extensive compared with FAPI tracers.<sup>72</sup>

## Thrombosis

Arterial and venous thrombosis plays an important role in various cardiovascular conditions, such as myocardial infarction, stroke, transient ischemic attacks, deep vein thrombosis, and pulmonary embolism. Arterial thrombosis results from clot formation in the setting of atherosclerotic plaque rupture, leading to platelet aggregation, thrombus formation, vessel occlusion, and possible myocardial infarction or ischemic stroke. The glycoprotein IIb/IIIa receptor is integral to acute thrombus formation, with expression falling as the thrombus matures.<sup>73</sup> It is an attractive target for both therapeutic modulation and molecular imaging of acute thrombus formation.

Early *ex vivo* and *in vivo* data have demonstrated that [ $^{18}\text{F}$ ]GP1 is highly sensitive to the formation of acute arterial and venous thrombus in humans (Figure 7). Kim et al<sup>74</sup> performed [ $^{18}\text{F}$ ]GP1 PET-CT in patients ( $n=20$ ) with either suspected lower limb deep vein thrombosis or acute pulmonary embolism demonstrating excellent image quality, signal-to-noise ratio, and diagnostic accuracy. Similarly, Chae et al<sup>75</sup> demonstrated the ability of [ $^{18}\text{F}$ ]GP1 to diagnose acute arterial thrombosis in 10 patients across multiple sites in the body including the abdominal aorta ( $n=7$ ), internal carotid artery ( $n=1$ ), superficial femoral artery ( $n=1$ ), and popliteal artery ( $n=1$ ). Furthermore, [ $^{18}\text{F}$ ]GP1 uptake is higher in the presence of thrombus associated with bioprosthetic valves compared with normal native aortic valves, and its [ $^{18}\text{F}$ ]GP1 uptake regresses with anticoagulation.<sup>76</sup> Its role has also been reported in acute myocardial infarction.<sup>77</sup> Other clinical studies are underway exploring the efficacy



**Figure 7. Case examples of [ $^{18}\text{F}$ ]glycoprotein 1 imaging.**

Data demonstrating increased [ $^{18}\text{F}$ ]GP1 ([ $^{18}\text{F}$ ]glycoprotein 1) uptake in (A) patient with left ventricular thrombus following heart infarct with tracer uptake overlying the thrombus on computed tomography (CT) and (B) in a patient with acute inferior myocardial infarction with high thrombus burden in the right coronary artery detected on angiography and clear [ $^{18}\text{F}$ ]GP1 uptake overlying the area of thrombus on CT. Note that there is almost no background myocardial uptake resulting in a high signal-to-noise ratio (data from The University of Edinburgh).

of [ $^{18}\text{F}$ ]GP1 PET-CT in detecting a range of pathological conditions, including left ventricular thrombus. Tantalizingly, a recent case report demonstrated that [ $^{18}\text{F}$ ]GP1 was able to identify acute thrombus in both the circumflex coronary artery and the left atrial appendage, leading to a change in clinical diagnosis from type 1 to type 2 myocardial infarction.<sup>78</sup> As demonstrated by the above, [ $^{18}\text{F}$ ]GP1 PET-CT imaging can be a promising tool in different cardiovascular conditions; however, a limited number and volumes of studies are available to date, and its potential role in clinical practice needs to be identified in future research.

Alternatively, Izquierdo-Garcia et al<sup>79</sup> demonstrated the feasibility of detecting (sub)acute left atrial appendage thrombus using combined PET/CMR of the fibrin-binding radiotracer [ $^{64}\text{Cu}$ ]FBP8 with good accuracy. This first-in-human study is promising, but further validation in larger prospective clinical trials is warranted.



## Cardiac Innervation Imaging

Increased myocardial sympathetic activity is a prominent feature of heart failure and is associated with progressive cardiac remodeling, decline in left ventricular function, and worsening symptoms. Alterations in myocardial sympathetic activity also play an important role in the generation of ventricular arrhythmias and sudden cardiac death.

### [<sup>123</sup>I]-META-IODOBENZYLGUANIDE

Cardiac SPECT with [<sup>123</sup>I]-meta-iodobenzylguanidine ([<sup>123</sup>I]-MIBG) is the most commonly used method to assess cardiac sympathetic innervation.<sup>80</sup> [<sup>123</sup>I]-MIBG is a guanethidine analog whose uptake at the postganglionic presynaptic nerve ending is mediated by noradrenaline uptake and storage mechanisms. Chronic high sympathetic cardiac activity results in reduced presynaptic noradrenaline uptake as an adaptive mechanism. The intensity of [<sup>123</sup>I]-MIBG uptake in the heart (quantified as a heart-to-mediastinum ratio) is, thus, inversely correlated with the degree of sympathetic activation. In patients with ischemic heart disease, measuring the heart-to-mediastinum ratio on [<sup>123</sup>I]-MIBG scintigraphy helps identify patients at high risk for developing heart failure, while in patients with established heart failure, irrespective of the cause, reduced cardiac uptake is associated with poor survival and a higher incidence of cardiac arrhythmias.<sup>81</sup> Indeed, in a study of 90 patients with chronic heart failure (New York Heart Association classes II–IV; ejection fraction <45%), the [<sup>123</sup>I]-MIBG heart-to-mediastinal ratio emerged as the best predictor of survival.<sup>82</sup> This powerful prognostic information has been confirmed in several other reports, including a study of 414 patients with known or suspected cardiac disease, where [<sup>123</sup>I]-MIBG scintigraphy again emerged as the most powerful predictor of cardiac death.<sup>83</sup>

### CARBON-11 META-HYDROXYEPHEDRINE

Carbon-11 meta-hydroxyephedrine ([<sup>11</sup>C]mHED) is a norepinephrine analog and the most commonly used PET radiotracer in cardiac sympathetic imaging. [<sup>11</sup>C]mHED was developed based on metamadol, a synthetic false transmitter analog of norepinephrine, which accumulates in nerve terminals such as norepinephrine. It is released by nerve stimulation but has only weak postsynaptic effects. Like MIBG, [<sup>11</sup>C]mHED shares the same neuronal uptake mechanism as norepinephrine and is resistant to enzymatic degradation, making it a good radioligand to assess the integrity of the presynaptic sympathetic nerve terminal.<sup>84</sup> High myocardium [<sup>11</sup>C]mHED uptake (79±31%) denotes an intact cardiac sympathetic nervous system, whereas reduced uptake is observed in diseased states such as heart failure.<sup>85</sup> Vesalainen

et al<sup>86</sup> demonstrated that global myocardial [<sup>11</sup>C]mHED uptake was 30% lower in patients with chronic heart failure compared with healthy controls. [<sup>11</sup>C]mHED also provides prognostic information. In the PAREPET trial (Prediction of Arrhythmic Events With Positron Emission Tomography), 204 patients with ischemic cardiomyopathy (left ventricular ejection fraction ≤35%) were imaged with [<sup>11</sup>C]mHED PET. The risk of sudden cardiac death was higher in patients with lower [<sup>11</sup>C]mHED uptake.<sup>87</sup> Further studies are needed, in particular, studies investigating whether its prognostic power might be used to guide the implantation of implantable cardioverter defibrillators for primary prevention.

## CHALLENGES AND OPPORTUNITIES

Advances in hybrid imaging technologies such as PET-CT and PET-MRI alongside improved motion correction and image analysis techniques make the noninvasive assessment of cardiovascular disease activity a clinical reality. Hybrid imaging combines the strengths of 2 powerful imaging modalities, allowing for the assessment of disease activity together with detailed anatomic information and tissue characterization. Nuclear cardiology has, nowadays, a large armamentarium of tools to prevent, detect, and correct motion, including techniques addressing heart contractions, tidal breathing, and patient repositioning; new algorithms to further improve the overall quality of the imaging study are under development. Recent technical advances have led to new possibilities for accurate image quantification and quantification of radiotracer kinetics, allowing for better evaluation of disease activity, and treatment response monitoring.<sup>88</sup> While the lack of specific radiotracers was previously an important barrier, we now have an array of tracers allowing us to measure inflammation, infection, activated fibroblasts, calcification activity, myocardial sympathetic activity, and thrombus formation as they are occurring in the body, potentially heralding a new era of cardiovascular imaging. However, it is important to acknowledge that not all molecular imaging targets hold the same clinical value. For example, the proximity of the imaging target to the therapeutic target and the differences between hot spot and cold spot imaging in therapy guidance are critical factors that influence the clinical impact of specific tracers. Some targets may have more immediate clinical relevance, particularly in guiding therapeutic decisions, while others might offer broader insights into disease processes. Although a detailed exploration of these variations is beyond the scope of this review, it is crucial to consider these factors in future research and clinical applications. A range of other tracers targeting yet more pathological processes are currently under development or are being tested in preclinical studies (Table S1) but are beyond the scope of this review article.

Despite its numerous benefits, molecular imaging remains expensive and not readily available at all centers. The increasing use of PET to assess patients with cancer is gradually addressing both of these issues, but further collaborative work is required to make molecular imaging more accessible and equitable for patients. Future clinical studies will need to demonstrate the clinical utility and cost-effectiveness of molecular imaging to justify its increased clinical use. These will need to be matched by improved education of cardiologists and imaging specialists, as well as patients and allied health professionals.

## FUTURE DIRECTIONS

Precision-based medicine, defined as tailored therapy based on an individual's underlying disease biology, now underpins research and clinical aspirations within many medical specialties. With further evolution of the field, molecular imaging has the potential to deliver precision-based cardiovascular medicine by identifying the active processes driving disease in a particular patient, identifying the presence or absence of a therapeutic target before initiation of therapy, and measuring the efficacy of treatment by repeated imaging. Theranostic application, or the coupling of imaging agent to the therapy, is already entering clinical practice best exemplified by cardiac sarcoidosis and amyloidosis but with major potential to revolutionize the application of molecular imaging in a wider range of cardiovascular conditions. In addition, molecular assessments of disease activity are increasingly used as an end point in clinical trials of novel therapies, providing rapid readouts that might accelerate the development of new treatments.

The future of molecular cardiovascular imaging will likely see a significant contribution from artificial intelligence and machine learning platforms. Artificial intelligence may help improve molecular cardiovascular imaging in various methods and domains, for example, reconstruction, denoising, partial volume correction, motion compensation, image registration, reconstruction and segmentation, and automated quantification.<sup>89</sup> Machine learning algorithms can efficiently connect information from multiple complex data sets and are well suited to hybrid imaging where information from PET can be combined with the rich information provided by CT and MRI.<sup>89</sup> Moreover, newer total body PET camera systems and newer technologies such as CardioFreeze will be able to offer higher sensitivity and better signal-to-noise ratio.

## CONCLUSIONS

Advances in molecular imaging and radiotracer discovery now allow the assessment of disease activity across a range of cardiovascular conditions, complementing the structural and functional information provided by

traditional imaging techniques. We currently have an array of radiotracers that inform about each of the major processes driving cardiovascular diseases, including inflammation, calcification, fibrosis, thrombosis, myocardial metabolism, and sympathetic innervation, and molecular imaging is, nowadays, an integral part of the clinical workup of patients with acute myocardial infarction, cardiac amyloidosis, endocarditis, and sarcoidosis. Cardiovascular molecular imaging is likely to play a pivotal role in improving our understanding of disease mechanisms, developing and screening novel therapeutics, as well as playing an increasing role in patient management in a wider range of cardiovascular conditions.

## ARTICLE INFORMATION

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### Disclosures

None.

### Supplemental Material

Table S1  
References 90–100

## REFERENCES

- Habib G, Lancellotti P, Antunes MJ, Bongiorno MG, Casalta J, Del Zotti F, Dulgheru R, El Khoury G, Erba PA, lung B, et al. 2015 ESC guidelines for the management of infective endocarditis. *Eur Heart J*. 2015;36:3075–3128. doi: 10.5603/KP.2015.0227
- Slart RHJA, Glaudemans AWJM, Lancellotti P, Hyafil F, Blankstein R, Schwartz RG, Jaber WA, Russell R, Gimelli A, Rouzet F, et al; Document Reading Group. A joint procedural position statement on imaging in cardiac

- sarcoidosis: from the cardiovascular and inflammation & infection committees of the European Association of Nuclear Medicine, the European Association of Cardiovascular Imaging, and the American Society of Nuclear Cardiology. *J Nucl Cardiol*. 2018;25:298–319. doi: 10.1007/s12350-017-1043-4
3. Start RHJA. FDG-PET/CT(A) imaging in large vessel vasculitis and polymyalgia rheumatica: joint procedural recommendation of the EANM, SNMMI, and the PET Interest Group (PIG), and endorsed by the ASNC. *Eur J Nucl Med Mol Imaging*. 2018;45:1250–1269. doi: 10.1007/s00259-018-3973-8
  4. Davidson CQ, Phenix CP, Tai TC, Khaper N, Lees SJ. Searching for novel PET radiotracers: imaging cardiac perfusion, metabolism and inflammation. *Am J Nucl Med Mol Imaging*. 2018;8:200–227. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6056242/>
  5. Almeida AG, Carpenter J, Cameli M, Donal E, Dweck MR, Flachskampf FA, Maceira AM, Muraru D, Neglia D, Pasquet A, et al. Multimodality imaging of myocardial viability: an expert consensus document from the European Association of Cardiovascular Imaging (EACVI). *Eur Hear J Cardiovasc Imaging*. 2021;22:e97–e125. doi: 10.1093/ehjci/jeab053
  6. Di Carli MF, Asgarzadeh F, Schelbert HR, Brunken RC, Laks H, Phelps ME, Maddahi J. Quantitative relation between myocardial viability and improvement in heart failure symptoms after revascularization in patients with ischemic cardiomyopathy. *Circulation*. 1995;92:3436–3444. doi: 10.1161/01.cir.92.12.3436
  7. Beanlands RSB, Nichol G, Huszti E, Humen D, Racine N, Freeman M, Gulenchyn KY, Garrard L, DeKemp R, Guo A, et al; PARR-2 Investigators. F-18-fluorodeoxyglucose positron emission tomography imaging-assisted management of patients with severe left ventricular dysfunction and suspected coronary disease. *J Am Coll Cardiol*. 2007;50:2002–2012. doi: 10.1016/j.jacc.2007.09.006
  8. Wang A, Athan E, Pappas PA, Fowler VG, Olaison L, Paré C, Almirante B, Muñoz P, Rizzi M, Naber C, et al; International Collaboration on Endocarditis-Prospective Cohort Study Investigators. Contemporary clinical profile and outcome of prosthetic valve endocarditis. *JAMA*. 2007;297:1354–1361. doi: 10.1001/jama.297.12.1354
  9. Sarrazin JF, Philippon F, Tessier M, Guimond J, Molin F, Champagne J, Nault I, Blier L, Nadeau M, Charbonneau L, et al. Usefulness of fluorine-18 positron emission tomography/computed tomography for identification of cardiovascular implantable electronic device infections. *J Am Coll Cardiol*. 2012;59:1616–1625. doi: 10.1016/j.jacc.2011.11.059
  10. de Camargo RA, Sommer Bitencourt M, Meneghetti JC, Soares J, Gonçalves LFT, Buchpiguel CA, Paixão MR, Felício MF, de Matos Soeiro A, Varejão Strabelli TM, et al. The role of 18F-fluorodeoxyglucose positron emission tomography/computed tomography in the diagnosis of left-sided endocarditis: native vs prosthetic valves endocarditis. *Clin Infect Dis*. 2020;70:583–594. doi: 10.1093/cid/ciz267
  11. Libby P, Bhatt DL, Di Carli M. Fluorodeoxyglucose uptake in atheroma. *J Am Coll Cardiol*. 2019;74:1233–1236. doi: 10.1016/j.jacc.2019.07.009
  12. Rudd JHF, Warburton EA, Fryer TD, Jones HA, Clark JC, Antoun N, Johnström P, Davenport AP, Kirkpatrick PJ, Arch BN, et al. Imaging atherosclerotic plaque inflammation with [18F]-fluorodeoxyglucose positron emission tomography. *Circulation*. 2002;105:2708–2711. doi: 10.1161/01.cir.0000020548.60110.76
  13. Hetterich H, Rominger A, Walter L, Habs M, Volpers S, Hacker M, Reiser MF, Bartenstein P, Saam T. Natural history of atherosclerotic disease progression as assessed by 18F-FDG PET/CT. *Int J Cardiovasc Imaging*. 2016;32:49–59. doi: 10.1007/s10554-015-0660-8
  14. Fayad ZA, Mani V, Woodward M, Kallend D, Abt M, Burgess T, Fuster V, Ballentyne CM, Stein EA, Tarif JC, et al. Safety and efficacy of dalcetrapib on atherosclerotic disease using novel non-invasive multimodality imaging (dal-PLAQUE): a randomised clinical trial. *Lancet*. 2011;378:1547–1559. doi: 10.1016/S0140-6736(11)61383-4
  15. Treglia G, Albano D, Dondi F, Bertagna F, Gheysens O. A role of FDG PET/CT for response assessment in large vessel disease? *Semin Nucl Med*. 2023;53:78–85. doi: 10.1053/j.semnuclmed.2022.08.002
  16. Ekström K, Lehtonen J, Nordenswan HK, Mäyränpää MI, Räisänen-Sokolowski A, Kandolin R, Simonen T, Rietilä-Effafi P, Alatalo A, Utriainen S, et al. Sudden death in cardiac sarcoidosis: an analysis of nationwide clinical and cause-of-death registries. *Eur Heart J*. 2019;40:3121–3128. doi: 10.1093/eurheartj/ehz428
  17. Birnie DH, Kandolin R, Nery PB, Kupari M. Cardiac manifestations of sarcoidosis: diagnosis and management. *Eur Heart J*. 2017;38:2663–2670. doi: 10.1093/eurheartj/ehw328
  18. Kusano KF, Satomi K. Diagnosis and treatment of cardiac sarcoidosis. *Heart*. 2016;102:184–190. doi: 10.1136/heartjnl-2015-307877
  19. Dweck MR, Abgral R, Trivieri MG, Robson PM, Karakatsanis N, Mani V, Palmisano A, Miller MA, Lala A, Chang HL, et al. Hybrid magnetic resonance imaging and positron emission tomography with fluorodeoxyglucose to diagnose active cardiac sarcoidosis. *JACC Cardiovasc Imaging*. 2018;11:94–107. doi: 10.1016/j.jcmg.2017.02.021
  20. Youssef G, Leung E, Mylonas I, Nery P, Williams K, Wisenberg G, Gulenchyn KY, deKemp RA, DaSilva J, Birnie D, et al. The use of 18 F-FDG PET in the diagnosis of cardiac sarcoidosis: a systematic review and meta-analysis including the Ontario experience. *J Nucl Med*. 2012;53:241–248. doi: 10.2967/jnumed.111.090662
  21. Ahmadian A, Pawar S, Govender P, Berman J, Ruberg FL, Miller EJ. The response of FDG uptake to immunosuppressive treatment on FDG PET/CT imaging for cardiac sarcoidosis. *J Nucl Cardiol*. 2017;24:413–424. doi: 10.1007/s12350-016-0490-7
  22. James OG, Christensen JD, Wong TZ, Borges-Neto S, Koweek LM. Utility of FDG PET/CT in inflammatory cardiovascular disease. *Radiographics*. 2011;31:1271–1286. doi: 10.1148/rg.315105222
  23. Pauwels E, Cleeren F, Bormans G, Deroose CM. Somatostatin receptor PET ligands - the next generation for clinical practice. *Am J Nucl Med Mol Imaging*. 2018;8:311–331.
  24. Lapa C, Reiter T, Siang L, Werner RA, Samnick S, Jahns R, Buck AK, Ertl G, Bauer WR. Imaging of myocardial inflammation with somatostatin receptor based PET/CT – a comparison to cardiac MRI. *Int J Cardiol*. 2015;194:44–49. doi: 10.1016/j.ijcard.2015.05.073
  25. Bartlett B, Ludewick HP, Lee S, Verma S, Francis RJ, Dwivedi G. Imaging inflammation in patients and animals: focus on pet imaging the vulnerable plaque. *Cells*. 2021;10:2573. doi: 10.3390/cells10102573
  26. Li X, Samnick S, Lapa C, Israel I, Buck AK, Kreissl MC, Bauer W. 68Ga-DOTATATE PET/CT for the detection of inflammation of large arteries: correlation with 18F-FDG, calcium burden and risk factors. *EJNMMI Res*. 2012;2:52. doi: 10.1186/2191-219X-2-52
  27. Kaushik P, Patel C, Gulati GS, Seth S, Parakh N, Guleria R, Kumar R, Gupta P, Bal C. Comparison of 68Ga-DOTANOC PET/CT with cardiac MRI in patients with clinical suspicion of cardiac sarcoidosis. *Ann Nucl Med*. 2021;35:1058–1065. doi: 10.1007/s12149-021-01641-4
  28. Dusi V, Ghidoni A, Ravera A, De Ferrari GM, Calvillo L. Chemokines and heart disease: a network connecting cardiovascular biology to immune and autonomic nervous systems. *Mediators Inflamm*. 2016;2016:1–16. doi: 10.1155/2016/5902947
  29. Reiter T, Kircher M, Schirbel A, Werner RA, Kropf S, Ertl G, Buck AK, Wester HJ, Bauer WR, Lapa C. Imaging of C-X-C motif chemokine receptor CXCR4 expression after myocardial infarction with [68Ga]pentixafor-PET/CT in correlation with cardiac MRI. *JACC Cardiovasc Imaging*. 2018;11:1541–1543. doi: 10.1016/j.jcmg.2018.01.001
  30. Derlin T, Sedding DG, Dutzmann J, Haghikia A, König T, Napp LC, Schütze C, Owsianski-Hille N, Wester HJ, Kropf S, et al. Imaging of chemokine receptor CXCR4 expression in culprit and nonculprit coronary atherosclerotic plaque using motion-corrected [68Ga]pentixafor PET/CT. *Eur J Nucl Med Mol Imaging*. 2018;45:1934–1944. doi: 10.1007/s00259-018-4076-2
  31. Hess A, Derlin T, Koenig T, Diekmann J, Witteben A, Wang Y, Wester HJ, Ross TL, Wollert KC, Bauersachs J, et al. Molecular imaging-guided repair after acute myocardial infarction by targeting the chemokine receptor CXCR4. *Eur Heart J*. 2020;41:3564–3575. doi: 10.1093/eurheartj/ehaa598
  32. Heo GS, Bajpai G, Li W, Luehmann HP, Sultan DH, Dun H, Leuschner F, Brody SL, Gropler RJ, Kreisel D, et al. Targeted PET imaging of chemokine receptor 2-positive monocytes and macrophages in the injured heart. *J Nucl Med*. 2021;62:111–114. doi: 10.2967/jnumed.120.244673
  33. English SJ, Sastriques SE, Detering L, Sultan D, Luehmann H, Arif B, Heo FS, Zhang S, Laforest R, Zheng J, et al. CCR2 positron emission tomography for the assessment of abdominal aortic aneurysm inflammation and rupture prediction. *Circ Cardiovasc Imaging*. 2020;13:E009889. doi: 10.1161/CIRCIMAGING.119.009889
  34. Gaemperli O, Shalhoub J, Owen DRJ, Lamare F, Johansson S, Fouladi N, Davies AH, Rimoldi OE, Camici PG. Imaging intraplaque inflammation in carotid atherosclerosis with 11C-PK11195 positron emission tomography/computed tomography. *Eur Heart J*. 2012;33:1902–1910. doi: 10.1093/eurheartj/ehr367
  35. Pugliese F, Gaemperli O, Kinderlerer AR, Lamare F, Shalhoub J, Davies AH, Rimoldi OE, Mason JC, Camici PG. Imaging of vascular inflammation with [11C]-PK11195 and positron emission tomography/computed tomography angiography. *J Am Coll Cardiol*. 2010;56:653–661. doi: 10.1016/j.jacc.2010.02.063
  36. Kreisl WC, Jendo KJ, Hines CS, Lyoo CH, Corona W, Morse CL, Zoghbi SS, Hyde T, Kleinman JE, Pike VW, et al. A genetic polymorphism for translocator protein 18 Kda affects both in vitro and in vivo radioligand binding in human brain to this putative biomarker of neuroinflammation. *J Cereb Blood Flow Metab*. 2013;33:53–58. doi: 10.1038/jcbfm.2012.131



37. MacAskill MG, Stadulyte A, Williams L, Morgan TEF, Sloan NL, Alcaide-Corral CJ, Walton T, Wimberley C, McKenzie CA, Spath N, et al. Quantification of macrophage-driven inflammation during myocardial infarction with 18 F-LW223, a novel TSPO radiotracer with binding independent of the rs6971 human polymorphism. *J Nucl Med*. 2021;62:536–544. doi: 10.2967/jnumed.120.243600
38. Pijl JP, Kwee TC, Slart RHJA, Glaudemans AWJM. PET/CT imaging for personalized management of infectious diseases. *J. Pers Med*. 2021;11:133. doi: 10.3390/jpm11020133
39. Creager MD, Hohl T, Hutcheson JD, Moss AJ, Schlotter F, Blaser MC, Park MA, Lee LH, Sing SA, Alcaide-Corral CJ, et al. 18F-fluoride signal amplification identifies microcalcifications associated with atherosclerotic plaque instability in positron emission tomography/computed tomography images. *Circ Cardiovasc Imaging*. 2019;12:e007835. doi: 10.1161/CIRCIMAGING.118.007835
40. Fontana M, orovi A, Scully P, Moon JC. Myocardial amyloidosis. *JACC Cardiovasc Imaging*. 2019;12:2345–2356. doi: 10.1016/j.jcmg.2019.06.023
41. Maurer MS, Bokhari S, Damy T, Dorbala S, Drachman BM, Fontana M, Grogan M, Kristen AV, Lousada I, Nativi-Nicolau J et al. Expert consensus recommendations for the suspicion and diagnosis of transthyretin cardiac amyloidosis. *Circ Heart Fail*. 2019;12:e006075. doi: 10.1161/CIRCHEARTFAILURE.119.006075
42. Perugini E, Guidalotti PL, Salvi F, Cooke RMT, Pettinato C, Riva L, Leone O, Farsad M, Ciliberti P, Bacchi-Reggiani L, et al. Noninvasive etiologic diagnosis of cardiac amyloidosis using 99m Tc-3,3-Diphosphono-1,2-propanodimethylcarboxylic acid scintigraphy. *J Am Coll Cardiol*. 2005;46:1076–1084. doi: 10.1016/j.jacc.2005.05.073
43. Martinez-Naharro A, Hawkins PN, Fontana M. Cardiac amyloidosis. *Clin Med (Lond)*. 2018;18:s30–s35. doi: 10.7861/clinmedicine.18-2-s30
44. Gillmore JD, Maurer MS, Falk RH, Merlini G, Damy T, Dispenzieri A, Weckalekar AD, Berk JL, Quarta CC, Grogan M, et al. Nonbiopsy diagnosis of cardiac transthyretin amyloidosis. *Circulation*. 2016;133:2404–2412. doi: 10.1161/CIRCULATIONAHA.116.021612
45. Sperry BW, Bock A, DiFilippo FP, Donnelly JP, Hanna M, Jaber WA. Pilot study of F18-florbetapir in the early evaluation of cardiac amyloidosis. *Front Cardiovasc Med*. 2021;8:693194. doi: 10.3389/fcvm.2021.693194
46. Dorbala S, Ando Y, Bokhari S, Dispenzieri A, Falk RH, Ferrari VA, Fontana M, Gheysens O, Gillmore JD, Glaudemans AWJM et al. ASNC/AHA/ASE/EANM/HFSA/ISA/SCMR/SNMMI expert consensus recommendations for multimodality imaging in cardiac amyloidosis: part 1 of 2 - evidence base and standardized methods of imaging. *Circ Cardiovasc Imaging*. 2021;14:e000029. doi: 10.1161/HCI.0000000000000029
47. Genovesi D, Vergaro F, Giorgetti A, Marzullo P, Scipioni M, Santarelli MF, Pucci A, Buda G, Vopi E, Emdin M. [18F]-Florbetaben PET/CT for differential diagnosis among cardiac immunoglobulin light chain, transthyretin amyloidosis, and mimicking conditions. *JACC Cardiovasc Imaging*. 2021;14:246–255. doi: 10.1016/j.jcmg.2020.05.031
48. Lee SP, Suh HY, Park S, Oh S, Kwak S, Kim H, Koh Y, Park J, Kim HK, Cho HJ, et al. Pittsburgh B compound positron emission tomography in patients with AL cardiac amyloidosis. *J Am Coll Cardiol*. 2020;75:380–390. doi: 10.1016/j.jacc.2019.11.037
49. Rosengren S, Skibsted CT, Tolbod L, Granstam S, Eiskjaer H, Wikström G, Vedin O, Kero T, Lubberink M, Harms H, et al. Diagnostic accuracy of [11C]PIB positron emission tomography for detection of cardiac amyloidosis. *JACC Cardiovasc Imaging*. 2020;13:1337–1347. doi: 10.1016/j.jcmg.2020.02.023
50. Garcia-Pavia P, aus dem Siepen F, Donal E, Lairez O, van der Meer P, Kristen AV, Mercuri MF, Michalon A, Frost RJA, Grimm J, et al. Phase 1 trial of antibody NIO06 for depletion of cardiac transthyretin amyloid. *N Engl J Med*. 2023;389:239–250. doi: 10.1056/NEJMoa2303765
51. Santarelli MF, Genovesi D, Scipioni M, Positano V, Favilli B, Giorgetti A, Vergaro G, Landini L, Emdin M, Marzullo P. Cardiac amyloidosis characterization by kinetic model fitting on [18F]florbetaben PET images. *J Nucl Cardiol*. 2022;29:1919–1932. doi: 10.1007/s12350-021-02608-8
52. Tzolos E, Dweck MR. 18F-Sodium Fluoride (18F-NaF) for imaging microcalcification activity in the cardiovascular system. *Arterioscler Thromb Vasc Biol*. 2020;40:1620–1626. doi: 10.1161/ATVBAHA.120.313785
53. Dweck MR, Chow MWL, Joshi NV, Williams MC, Jones C, Fletcher AM, Richardson H, White A, McKillop G, van Beek EJR, et al. Coronary arterial 18F-sodium fluoride uptake: a novel marker of plaque biology. *J Am Coll Cardiol*. 2012;59:1539–1548. doi: 10.1016/j.jacc.2011.12.037
54. Joshi NV, Vesey AT, Williams MC, Shah ASV, Calvert PA, Craighead FHM, Yeoh SE, Wallace W, Salter D, Fletcher AM, et al. 18F-fluoride positron emission tomography for identification of ruptured and high-risk coronary atherosclerotic plaques: a prospective clinical trial. *Lancet*. 2014;383:705–713. doi: 10.1016/S0140-6736(13)61754-7
55. Doris MK, Meah MN, Moss AJ, Andrews JPM, Bing R, Gillen R, Weir N, Syed M, Daghmagh M, Shah A, et al. Coronary 18 F-fluoride uptake and progression of coronary artery calcification. *Circ Cardiovasc Imaging*. 2020;13:e011438. doi: 10.1161/CIRCIMAGING.120.011438
56. Kwiecinski J, Tzolos E, Adamson PD, Cadet S, Moss AJ, Joshi N, Williams MC, van Beek EJR, Dey D, Berman DS, et al. Coronary 18F-sodium fluoride uptake predicts outcomes in patients with coronary artery disease. *J Am Coll Cardiol*. 2020;75:3061–3074. doi: 10.1093/ehjci/jez152
57. Moss AJ, Daghmagh M, Tzolos E, Meah MN, Wang KL, Bularga A, Adamson PD, Kwiecinski J, Fletcher A, Dawson D, et al; PREFFIR Investigators. Coronary atherosclerotic plaque activity and future coronary events. *JAMA Cardiol*. 2023;8:755–764. doi: 10.1001/jamacardio.2023.1729
58. Dweck MR, Jones C, Joshi NV, Fletcher AM, Richardson H, White A, Marsden M, Pessotto R, Clark JC, Wallace WA, et al. Assessment of valvular calcification and inflammation by positron emission tomography in patients with aortic stenosis. *Circulation*. 2012;125:76–86. doi: 10.1161/CIRCULATIONAHA.111.051052
59. Massera D, Trivieri MG, Andrews JPM, Sartori S, Abgall R, Chapman AR, Jenkins WSA, Vesey AT, Doris MK, Pawade TA, et al. Disease activity in mitral annular calcification. *Circ Cardiovasc Imaging*. 2019;12:e008513. doi: 10.1161/CIRCIMAGING.118.008513
60. Cartledge TRG, Doris MK, Sellers SL, Pawade TA, White AC, Pessotto R, Kwiecinski J, Fletcher A, Alcaide C, Lucatelli C, et al. Detection and prediction of bioprosthetic aortic valve degeneration. *J Am Coll Cardiol*. 2019;73:1107–1119. doi: 10.1016/j.jacc.2018.12.056
61. Kwiecinski J, Tzolos E, Cartledge TRG, Fletcher A, Doris MK, Bing R, Tarkin JM, Seidman MA, Gulsin GS, Cruden NL, et al. Native aortic valve disease progression and bioprosthetic valve degeneration in patients with transcatheter aortic valve implantation. *Circulation*. 2021;144:1396–1408. doi: 10.1161/CIRCULATIONAHA.121.056891
62. Bing R, Dweck MR. Myocardial fibrosis: why image, how to image and clinical implications. *Heart*. 2019;105:1832–1840. doi: 10.1136/heartjnl-2019-315560
63. Kelly T, Huang Y, Simms AE, Mazur A. Fibroblast activation protein-1: a key modulator of the microenvironment in multiple pathologies. *Int Rev Cell Mol Biol*. 2012;297:83–116. doi: 10.1016/B978-0-12-394308-8.00003-0
64. Loktev A, Lindner T, Mier W, Debus J, Altmann A, Jäger D, Giesel F, Kratochwil C, Barthe P, Roumestand C, et al. A Tumor-imaging method targeting cancer-associated fibroblasts. *J Nucl Med*. 2018;59:1423–1429. doi: 10.2967/jnumed.118.210435
65. Bengel FM, Diekmann J, Hess A, Jerosch-Herold M. Myocardial fibrosis: emerging target for cardiac molecular imaging and opportunity for image-guided therapy. *J Nucl Med*. 2023;64:49S–58S. doi: 10.2967/jnumed.122.264867
66. Heckmann MB, Reinhardt F, Finke D, Katus HA, Haberkorn U, Leuschner F, Lehmann LH. Relationship between cardiac fibroblast activation protein activity by positron emission tomography and cardiovascular disease. *Circ Cardiovasc Imaging*. 2020;13:e010628. doi: 10.1161/CIRCIMAGING.120.010628
67. Diekmann J, Koenig T, Thackeray JT, Derlin T, Czerner C, Neuser J, Toss TL, Schäfer A, Tillmanns J, Bauersachs J, et al. Cardiac fibroblast activation in patients early after acute myocardial infarction: integration with MR tissue characterization and subsequent functional outcome. *J Nucl Med*. 2022;63:1415–1423. doi: 10.2967/jnumed.121.263555
68. Diekmann J, Neuser J, Röhrich M, Derlin T, Zwadlo C, Koenig T, Weiberg D, Jäckle F, Kempf T, Ross TL, et al. Molecular imaging of myocardial fibroblast activation in patients with advanced aortic stenosis before transcatheter aortic valve replacement: a pilot study. *J Nucl Med*. 2023;64:1279–1286. doi: 10.2967/jnumed.122.265147
69. Aghajanian H, Kimura T, Rurik JG, Hancock AS, Leibowitz MS, Li L, Scholler J, Wonslow J, Lo A, Han W, et al. Targeting cardiac fibrosis with engineered T cells. *Nature*. 2019;573:430–433. doi: 10.1038/s41586-019-1546-z
70. Rurik JG, Tombacz I, Yadegari A, Fernandez POM, Shewale SV, Li L, Kimura T, Soliman OY, Papp TE, Tam YK, et al. CAR T cells produced in vivo to treat cardiac injury. *Science*. 2022;375:91–96. doi: 10.1126/science.abm0594
71. Xin L, Gao J, Zheng Z, Chen Y, Lv S, Zhao Z, Chunhai Y, Yang X, Zhang R. Fibroblast activation protein-1 as a target in the bench-to-bedside diagnosis and treatment of tumors: a narrative review. *Front Oncol*. 2021;11:648187. doi: 10.3389/fonc.2021.648187
72. De Haas HJ, Arbustini E, Fuster V, Kramer CM, Narula J. Molecular imaging of the cardiac extracellular matrix. *Circ Res*. 2014;114:903–915. doi: 10.1161/CIRCRESAHA.113.302680
73. Fullard J. The role of the platelet glycoprotein IIb/IIIa in thrombosis and haemostasis. *Curr Pharm Des*. 2004;10:1567–1576. doi: 10.2174/1381612043384682



74. Kim C, Lee JS, Han Y, Chae SY, Jin S, Sung C, Son HJ, Oh SJ, Lee SJ, Oh JS, et al. Glycoprotein IIb/IIIa receptor imaging with 18 F-GP1 PET for acute venous thromboembolism: an open-label, nonrandomized, phase 1 study. *J Nucl Med*. 2019;60:244–249. doi: 10.2967/jnumed.118.212084
75. Chae SY, Kwon TW, Jin S, Kwon SU, Sung C, Oh SJ, Lee SJ, Oh JS, Han Y, Cho YP, et al. A phase 1, first-in-human study of 18F-GP1 positron emission tomography for imaging acute arterial thrombosis. *EJNMMI Res*. 2019;9:3. doi: 10.1186/s13550-018-0471-8
76. Bing R, Deutsch MA, Sellers SL, Corral CA, Andrews JPM, van Beek EJR, Bleiziffer S, Burchert W, Clark T, Dey D, et al. 18F-GP1 positron emission tomography and bioprosthetic aortic valve thrombus. *JACC Cardiovasc Imaging*. 2022;15:1107–1120. doi: 10.1016/j.jcmg.2021.11.015
77. Tzolos E, Bing R, Jack AJ, MacAskill MG, Tavares AAS, Macnaught G, Clark T, Mills ML, Fujisawa T, Nash J, et al. Non-invasive in vivo coronary artery thrombus imaging. *JACC Cardiovasc Imaging*. 2022;16:820–832. doi: 10.1016/j.jcmg.2022.10.002
78. Tzolos E, Bing R, Newby DE, Dweck MR. Categorising myocardial infarction with advanced cardiovascular imaging. *Lancet*. 2021;398:e9. doi: 10.1016/S0140-6736(21)01329-5
79. Izquierdo-Garcia D, Désogère P, Philip AL, Mekkaoui C, Weiner RB, Catalano OA, Chen YCI, Yeh DD, Mansour M, Catana C, et al. Detection and characterization of thrombosis in humans using fibrin-targeted positron emission tomography and magnetic resonance. *JACC Cardiovasc Imaging*. 2022;15:504–515. doi: 10.1016/j.jcmg.2021.08.009
80. Gimelli A, Liga R, Agostini D, Bengel FM, Ernst S, Hyafil F, Saraste A, Scholte AJHA, Verberne HJ, Verschure DO et al. The role of myocardial innervation imaging in different clinical scenarios: an expert document of the European Association of Cardiovascular Imaging and Cardiovascular Committee of the European Association of Nuclear Medicine. *Eur Heart J Cardiovasc Imaging*. 2021;22:480–490. doi: 10.1093/ehjci/jeab007
81. Kioka H, Yamada T, Mine T, Morita T, Tsukamoto Y, Tamaki S, Masuda M, Okuda K, Hori M, Fukunami M. Prediction of sudden death in patients with mild-to-moderate chronic heart failure by using cardiac iodine-123 metaiodobenzylguanidine imaging. *Heart*. 2007;93:1213–1218. doi: 10.1136/hrt.2006.094524
82. Merlet P, Valette H, Dubois-Randé JL, Moysé D, Duboc D, Dove P, Bourguignon MH, Benvenuti C, Duval AM, Agotini D, et al. Prognostic value of cardiac metaiodobenzylguanidine imaging in patients with heart failure. *J Nucl Med*. 1992;33:471–477.
83. Nakata T. Cardiac death prediction and impaired cardiac sympathetic innervation assessed by MIBG in patients with failing and nonfailing hearts. *J Nucl Cardiol*. 1998;5:579–590. doi: 10.1016/s1071-3581(98)90112-x
84. Rosenspire KC, Haka MS, Van Dort ME, Jewett DM, Gildersleeve DL, Schwaiger M, Wieland DM. Synthesis and preliminary evaluation of carbon-11-meta-hydroxyephedrine: a false transmitter agent for heart neuronal imaging. *J Nucl Med*. 1990;31:1328–1334.
85. Schwaiger M, Kalff V, Rosenspire K, Haka MS, Molina E, Hutchins GD, Deeb M, Wolfe E Jr, Wieland DM. Noninvasive evaluation of sympathetic nervous system in human heart by positron emission tomography. *Circulation*. 1990;82:457–464. doi: 10.1161/01.cir.82.2.457
86. Vesalainen RK, Pietilä M, Tahvanainen KU, Jartti T, Teräs M, Nagren K, Lehtikoinen P, Huupponen R, Ukkonen H, Saraste M, et al. Cardiac positron emission tomography imaging with [11C]hydroxyephedrine, a specific tracer for sympathetic nerve endings, and its functional correlates in congestive heart failure. *Am J Cardiol*. 1999;84:568–574. doi: 10.1016/s0002-9149(99)00379-3
87. Fallavollita JA, Heavey BM, Luisi AJ Jr, Michalek SM, Baldwa S, Mashtare TL Jr, Hutson AD, Dekemp RA, Haka MS, Sajjad M, et al. Regional myocardial sympathetic denervation predicts the risk of sudden cardiac arrest in ischemic cardiomyopathy. *J Am Coll Cardiol*. 2014;63:141–149. doi: 10.1016/j.jacc.2013.07.096
88. Tingen HAS, van Praagh GD, Nienhuis PH, Tubben A, van Rijsewijk ND, ten Hove D, Mushari NA, Martinez-Lucio TS, Mendoza-Ibanez OI, van Sluis J, et al. The clinical value of quantitative cardiovascular molecular imaging: a step towards precision medicine. *Br J Radiol*. 2023;96:20230704. doi: 10.1259/bjr.20230704
89. Slart RHJA, Williams MC, Juarez-Orozco LE, Rischpler C, Dweck MR, Glaudemans AWJM, Fimelli A, Georgoulas P, Gheysens O, Gaemperli O, et al. Position paper of the EACVI and EANM on artificial intelligence applications in multimodality cardiovascular imaging using SPECT/CT, PET/CT, and cardiac CT. *Eur J Nucl Med Mol Imaging*. 2021;48:1399–1413. doi: 10.1007/s00259-021-05341-z
90. Moisis O, Palani S, Virta J, Elo P, Liljenbäck H, Tolvanen T, Käkälä M, Miner MG, Herre EA, Marjamäki P, et al. Radiosynthesis and preclinical evaluation of [68Ga]Ga-NOTA-folate for PET imaging of folate receptor  $\beta$ -positive macrophages. *Sci Rep*. 2020;10:13593. doi: 10.1038/s41598-020-70394-3
91. Gona K, Toczek J, Ye Y, Sanzida N, Golbazi A, Boodagh P, Salarian M, Jung JJ, Rajendran S, Kukreja G, et al. Hydroxamate-based selective macrophage elastase (MMP-12) inhibitors and radiotracers for molecular imaging. *J Med Chem*. 2020;63:15037–15049. doi: 10.1021/acs.jmedchem.0c01514
92. Huisman LA, Steinkamp PJ, Hillebrands JL, Zeebregts CJ, Linsen MD, Jorritsma-Smit A, Slart RHJA, van Dam GM, Boersma HH. Feasibility of ex vivo fluorescence imaging of angiogenesis in (non-) culprit human carotid atherosclerotic plaques using bevacizumab-800CW. *Sci Rep*. 2021;11:2899. doi: 10.1038/s41598-021-82568-8
93. Woodard P, Zayed M, Laforest R, Li R, Zheng J, Lin CY, Detering L, Sultan D, Luehmann H, Heo G, et al. Targeted NPR-C PET/MR imaging of carotid atherosclerosis in humans: correlation with ex vivo plaque immunohistochemistry (IHC) and patient outcomes. *J Nucl Med*. 2021;62:131. [https://jnm.snmjournals.org/content/62/supplement\\_1/131](https://jnm.snmjournals.org/content/62/supplement_1/131)
94. Makowski MR, Rischpler C, Ebersberger U, Keithahn A, Kasel M, Hoffmann E, Rassaf T, Kessler H, Wester HJ, Nekolla SG, et al. Multiparametric PET and MRI of myocardial damage after myocardial infarction: correlation of integrin  $\alpha\beta3$  expression and myocardial blood flow. *Eur J Nucl Med Mol Imaging*. 2021;48:1070–1080. doi: 10.1007/s00259-020-05034-z
95. Andrews JPM, Portal C, Walton T, Macaskill MG, Hadoke PWF, Corral CA, Lucatelli C, Wilson S, Wilson I, MacNaught G, et al. Non-invasive in vivo imaging of acute thrombosis: development of a novel factor XIIIa radiotracer. *Eur Heart J Cardiovasc Imaging*. 2020;21:673–682. doi: 10.1093/ehjci/jez207
96. Balogh V, Spath N, Alcaide-Corral C, Walton T, Lennen R, Jansen M, Lucatelli C, Newby DE, MacAskill M, Hadoke P, et al. Assessment of myocardial fibrosis activity using 18 f-fluoropropyl positron emission tomography (pet) in rat models of cardiovascular disease. *Heart*. 2020;106:A11.2–A11.1.
97. Burban A, Slupic D, Reda A, Szczerba E, Grabowski M, Kolodzinska A. Novel diagnostic methods for infective endocarditis. *Int J Mol Sci*. 2024;25:1245. doi: 10.3390/ijms25021245
98. Jenkins WSA, Vesey AT, Stirrat C, Connell M, Lucatelli C, Neale A, Moles C, Vickers A, Fletcher A, Pawade T, et al. Cardiac  $\alpha\beta3$  integrin expression following acute myocardial infarction in humans. *Heart*. 2017;103:607–615. doi: 10.1136/heartjnl-2016-310115
99. Sun Y, Zng Y, Shu Y, Feng F, Su W, Wu C, Xing B, Zhang W, Wu P, Cui L, et al. Application of 68Ga-PRGD2 PET/CT for  $\alpha\beta3$ -integrin imaging of myocardial infarction and stroke. *Theranostics*. 2014;4:778–786. doi: 10.7150/thno.8809
100. Sivapackiam J, Kabra S, Speidel S, Sharma M, Laforest R, Salter A, Rettig MP, Sharma V. 68Ga-galmydar: a PET imaging tracer for noninvasive detection of Doxorubicin-induced cardiotoxicity. *PLoS One*. 2019;14:e0215579. doi: 10.1371/journal.pone.0215579