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# Future of Team-based Basic and Translational Science in Radiation Oncology

R.P. Coppes, PhD,<sup>\*,†</sup> and L.V. van Dijk, PhD<sup>\*</sup>

To further optimize radiotherapy, a more personalized treatment towards individual patient's risk profiles, dissecting both patient-specific tumor and normal tissue response to multimodality treatments is needed. Novel developments in radiobiology, using *in vitro* patient-specific complex tissue resembling 3D models and multiomics approaches at a spatial single-cell level, may provide unprecedented insight into the radiation responses of tumors and normal tissue. Here, we describe the necessary team effort, including all disciplines in radiation oncology, to integrate such data into clinical prediction models and link the relatively "big data" from the clinical practice, allowing accurate patient stratification for personalized treatment approaches.

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## Introduction

The low-hanging fruits of radiation oncology have been discovered already, leaving more difficult challenges to discover novel paradigms. More accurate tumor irradiation with various irradiation qualities, dose tempo, improved target volume, and margin definition, (re)planning techniques, NTCP modeling, understanding of drug-radiation interaction, and precision medicine have been explored and implemented. These have improved tumor control and prolonged survival for many tumor sites<sup>1–3</sup> but have consequently resulted in an increased number of cancer survivors dealing with late normal tissue side effects. To further optimize cancer treatment, radiotherapy must be more personalized toward the individual patient's risk profile, dissecting both patient-specific tumor and normal tissue radiosensitivity, immune response, and drug/immune therapy radiation interaction. This to ensure improved tumor response with less normal tissue effects for cancer patients receiving radiation. Moreover, predicting treatment response and advancing knowledge on post-treatment tumor and normal tissue micro-environment reactions is essential to optimally

facilitate tumor control while stimulating normal tissue regeneration.<sup>4–6</sup> Unprecedented, combined expertise is required to achieve this in its full capacity. In other words, radiation biologists, molecular biologists, stem-cell biologists, geneticists, computational biologists, bioinformaticians, translational biologists/physicians, physicists, oncologists, radiologists, pathologists, surgeons, and radiation oncologists need to work together to delve into complex problems. This perspective paper focuses on novel developments in personalized radiotherapy from a radiobiologist's view matched with the clinical translational counterpart, where clinical information—available from routine clinical practice—can be extracted and used for toxicity or tumor outcome prediction.

Novel technological developments in radio-biology research have yielded unprecedented possibilities to study radiation effects on more complex animal and patient-derived models, ranging from *in vitro* cell lines and organoids to *ex vivo* tissue slices and *in vivo* humanized animal models. This combination of models allows for a better understanding of the damage mechanism and response and a more rapid translation to the clinic due to the closer resemblance of the patients. Next to that, current developments in multiomics approaches allow for an integrative analysis of radiation response parameters, providing unprecedented insight into the radiation responses of tumors and normal tissue. A multiomics approach may combine tumor and normal tissue data sets of genome, proteome, transcriptome, epigenome, metabolome, and/or microbiome to study responses of cells and tissues to, e.g., cancer treatments in a concerted way.

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The possibility of performing radiation research using multiomics at a single cell level in a spatial environment<sup>7,8</sup> will yield novel approaches to optimize radiation responses. The resulting complex biological big data set can be analyzed to find novel associations between biological processes, and indicate biomarkers, providing more in-depth knowledge of radiation response processes. A close interaction between bioinformaticians and computational biologists is needed to analyze these data. Regretfully, for analysis of patient-specific responses, these methodologies are presently too time-consuming and costly for individual prediction.

While generic response predictors/biomarker changes have already been tested in the clinic, this is where the gap with clinical translational research regarding prediction modeling needs to be bridged, as rapid advances in biotechnological and artificial intelligence (AI) methodologies allow the integration of multiomics data to predict treatment responses from diverse dimension levels. Particularly, since the bulk of translational research on the image "biomarkers"—otherwise referred to as radiomics features—that are extracted from routinely acquired imaging<sup>9-11</sup> often convey a similar message as that from the (radio)biological community, e.g., high tumor heterogeneity detected on CT is related to poorer outcomes<sup>12,13</sup>. Additionally, capturing the metabolic activity, FDG-PET imaging features have also shown that higher metabolic values in the tumor are linked to poorer outcome.<sup>14-18</sup> Higher PET intensity was linked to better normal gland tissue outcome, while for lung tissue, higher metabolic activity increases the risk of radiation-induced toxicity.<sup>18-23</sup> This shows how biological processes can be captured in imaging, which is also exemplified by the PET imaging studies aiming to capture hypoxia culture in tumor tissue<sup>24</sup> or Diffusion-Weighted Imaging, which estimates the diffusion of water molecules in tissue to detect tumor tissue or quantify damage.<sup>25-28</sup> Yet, other image biomarkers such as spatial tumor features—e.g., tumor volume, surface, sphericity, and compactness—often highly associated with the tumor response,<sup>11,29</sup> cannot be captured with *in vivo* biological biomarkers, which are typically obtained through biopsies or incisions. Hence, biological, omics, and image biomarkers can be highly complementary to each other in predicting tumor and normal tissue response. Moreover, relating multiomics biomarkers to predictive patient-specific imaging biomarkers extracted from PET-CT imaging data can strengthen individualized outcome prediction, as both suffer from institute and material susceptibility.<sup>30-33</sup> Not only could the current clinical prediction models be enriched with the right biological markers, but the link between the relatively "big data" from the clinical practice could also be delved to identify relevant biological markers that allow accurate patient stratification for personalized treatment approaches (see Fig. 1).

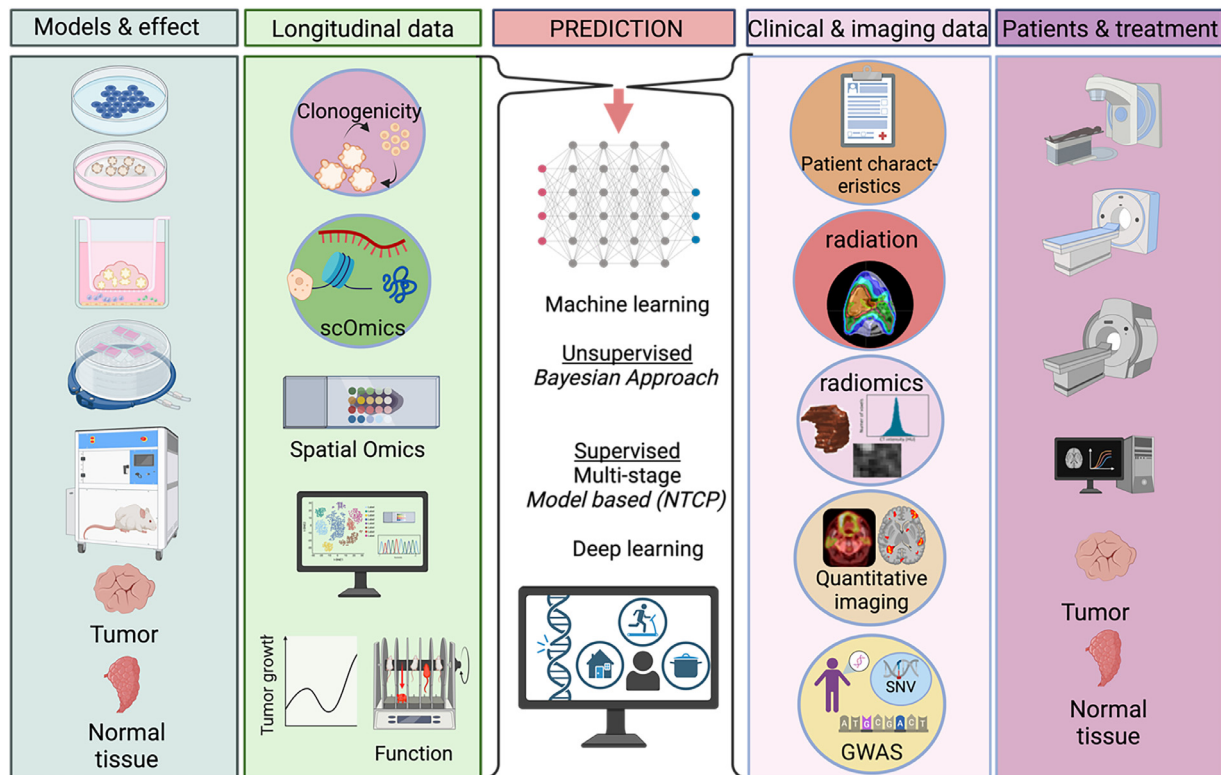
This perspective will address the possibilities available and how (patient-specific) radiobiology and clinical parameters could be combined to yield a rapid and highly individualized prediction of future patient responses.

## Biological Approaches

Cell lines have provided invaluable information on cellular radiation response characteristics, DNA damage response, cell cycle, radiosensitivity, etc., and led to one of the most used prediction models, the LQ model.<sup>2</sup> Cell lines will also, in the future, provide us with more mechanistic insight into the molecular response to radiation and combined therapies and can be used to validate big data-derived correlative mechanisms. However, although it is valuable to test molecular mechanisms, they will not provide individual patient-specific response parameters. Moreover, cell lines have performed poorly in establishing normal tissue responses, mainly because no good cell lines represent normal tissue complexity. Fibroblasts have been used to predict normal tissue responses, but besides showing some relation with fibrosis, they have largely failed to describe and predict the complex response of normal tissues. Recently, the landscape has changed with the development of 3-dimensional (3D) culture systems, allowing cell lines to grow as spheroids, showing more realistic cell-cell interactions and the radiation dose response.<sup>34,35</sup> However, even these still consist of only a single cell type, which does not represent the diversity of cell types in tumors or normal tissues. Other models are now available for these.

Organoids are 3D models derived from stem cells capable of self-renewal and contain multiple cell types of a complex structure that resembles the normal or tumor tissue of origin. They recapitulate the characteristics and functionality of the tissue, including genetic mutations.<sup>36</sup> Therefore, organoids represent an excellent model for studying patient-specific tumor and normal tissue radiation responses.<sup>35,37</sup> Organoids can be cultured from embryonic stem cells, induced pluripotent stem cells, animal tissues, and patient biopsies, allowing diagnostics, disease prognosis, treatment outcome prediction, and organoid cell-based therapies.<sup>38</sup> Indeed, organoids can provide treatment prediction for a variety of cancers such as colorectal,<sup>39</sup> head and neck cancer,<sup>40</sup> and pancreatic cancer,<sup>41</sup> while trials on lung cancer, liver cancer, kidney cancer, brain cancer, esophageal cancer, and breast cancer are ongoing.<sup>38</sup>

Normal tissue organoids have also revealed exciting characteristics, as shown with salivary gland and thyroid gland-derived organoids showing low-dose hypersensitivity.<sup>42</sup> Moreover, the feasibility of a patient-tailored evaluation of gastrointestinal response to radiation using human intestinal organoids using comparative transcriptome profiling to photon, proton, and carbon irradiation was shown.<sup>43</sup> Another interesting approach was recently published, where bulk and single-cell RNAseq reveal different transcriptomic changes of organoid-derived cells after photon and proton irradiation.<sup>44</sup> Interestingly, upon proton irradiation, a higher expression *INF-β* for many tumor sites related genes, cytoplasmic dsRNA, and expression of latent endogenous retroviruses was observed, leading to a robust immune activation and inflammatory response, like what is seen after a viral infection.<sup>45,46</sup> A loss of histone H3K9 methylation and consequential derepression of transposable elements



**Figure 1** Schematic representation of the potential methods and "bio" markers to predict the patient's treatment response. From the left to the middle, models for radiobiology (blue) allow the collection of data on radiation responses, (cancer) stem cell function, specific cellular and spatial responses, and tumor plus normal tissue responses (green). From the right to the middle in purple, patient treatment planning and dose distribution can be correlated to the patient's characteristic genetic information (pink), imaging data, normal tissue function, and tumor response measurements. The white column in the middle shows how the biology data can be merged with clinical data to establish response prediction models.

accompanied this. This process could be directly related to the stem cell response within the organoids, inducing an enhanced regenerative potential after proton compared to photon irradiation.<sup>44</sup> Interestingly, it was shown that a similar resurrection of endogenous retroviruses was observed during aging, which reinforced cellular senescence.<sup>47</sup> These studies indicate the potential of using organoids to study biological processes involved in the radiation response, allow the detection of biomarkers, and predict patient responses. However, organoids do not represent the complete tissue, as they lack blood vessels, immune cells, and (tumor-associated) fibroblasts. For this, co-culture "assembloids" have been developed to aggregate cells of different lineages able to self-organize, giving rise to more complex multilineage interactive organoids.<sup>48,49</sup> Next to this, a co-culture system where semi-permeable membranes separate organoids from cell types of other lineages may provide additional information.<sup>50</sup> Subsequent postirradiation multiomics will provide novel information on the radiation response of the different cell types and how they interact. Although promising still, these models do not entirely reflect the tissue in its normal state.

Human precision-cut tissue slices represent an organ mini-model that closely resembles the organ from which it is prepared, with all cell types in an original tissue-matrix configuration. Tissue slices have been used for drug discovery

and development.<sup>51</sup> They may yield information on metabolism and radiation-drug interactions in normal and tumor tissue. The multicellularity, intact cell-matrix interaction, and metabolism can be used to study the viability and functionality of radiation-affected (e.g., fibrotic) tissue. In capable hands, slices can remain viable for many days.<sup>51</sup> As such, an ex vivo tissue slice culture model of clear cell renal cell carcinoma was developed to elucidate the impact of nivolumab on tumor-infiltrating immune cells,<sup>52</sup> indicating the robustness of the model in identifying patient response to specific therapeutics.

That acts as a decoy hijacking the DNA damage response. It was observed that the drug protected the normal tissue by triggering a G1/S arrest.<sup>53</sup> Regretfully, precision cut slides have a limited viability of about 2 weeks maximum under optimal conditions and may also be overgrown by a specific rapidly proliferating cell type. However, this model may represent a midway between organoids and humanized animal models.

Humanized mouse models are immunodeficient mice co-grafted with human tumors and innate immune components. They can be used to study patient-specific interactions between engrafted matched tumors and immune cells. They are used in immuno-oncology research with potential for clinical translation.<sup>54</sup> Regretfully, there is a lack of preclinical studies using this model for radiobiology-based research.

However, it could be used to study radiation responses involving the innate immune system, such as the "abscopal effect".<sup>55</sup> Interestingly, using a lung squamous cell carcinoma humanized mouse model, it was shown that targeting NK cells may be an essential strategy for combining radiotherapy with immune checkpoint inhibitors.<sup>56</sup>

However, humanized mouse models show a restricted development of mature innate immune cells and lack HLAs and antigen-specific antibody responses.<sup>54</sup> Next-generation novel humanized mouse platforms may accelerate the research on the immune response to radiation in combination with (new) immunotherapies.

## Clinical Approaches

Clinical prediction models are based on clinical tabular data and can predict various treatment outcomes. Examples are clinical models that predict overall survival for head and neck cancer,<sup>57-60</sup> Non-Small Cell Lung Cancer,<sup>61</sup> and oligometastatic disease metastatic disease<sup>62</sup> or biochemical outcome in prostate cancer patients.<sup>63</sup> These models typically incorporate a range of predictors, including patient characteristics (e.g., age, sex, performance score, pack years), tumor characteristics (e.g., AJCC stage, tumor location, Gleason score), and occasionally treatment information (e.g., chemotherapy, radiation dose) to predict a patient-specific risk score. Some also consider semantic imaging features (e.g., extranodal extension) or biomarkers (e.g., HPV status or PSA levels). In contrast to the biological approaches, larger quantities of data are relatively simple to collect, particularly with the institution of efficient standardized follow-up programs and with the (near) future potential of mining all clinical data from electronic health records with natural language processing (NLP) techniques into model-ready tabular data.<sup>64</sup> Big clinical datasets have allowed for robustly trained and validated models supported by large, representative, multi-institutional datasets.<sup>58,60</sup>

Clinical models are often presented in a nomogram format,<sup>57,59,62,65</sup> while graphical user interface applications<sup>66</sup> or web-based applications<sup>58,60</sup> are more sophisticated methods to present the treatment outcome probability.<sup>58,60,66</sup> Regardless, prediction models are rarely used in the clinic but rather indirectly incorporated in our oncologic decision protocols, typically using binary cutoffs of risk factors, e.g., adding concomitant chemotherapy based on the AJCC stage and an age limit only. Yet, mortality and tumor outcome remain highly variable within these defined groups, highlighting the need for improved personalization of treatment with model-based risk estimations.<sup>58</sup>

Normal Tissue Complication Probability (NTCP) models predict radiation-induced side effects that are generally based on pretreatment dose parameters of organs-at-risk, combined with clinical variables, such as age or baseline complaints.<sup>67</sup> Examples are NTCP models that predict xerostomia with mean salivary gland doses<sup>68</sup> with mean lung dose<sup>69</sup> and rectal toxicity with doses related to the

substructures within and around the anorectum.<sup>70</sup> Several countries have implemented NTCP models in clinical decision-making to select patients for advanced normal tissue sparing radiotherapy techniques, such as proton therapy.<sup>71</sup> Moreover, the paradigm shift of radiation dose optimization based on NTCP models (model-based optimization) instead of radiation dose objectives may become a commercially available application in the future.<sup>72-74</sup>

Radiomics models are based on features (i.e., image "biomarkers") that quantify patient-specific tissue characteristics from clinical images, translating intensity, texture, and geometric properties region-of-interest into single variables that can be fed into a machine learning modeling pipeline.<sup>9</sup> Radiomics hypothesizes that these high-dimensional features are minable and can quantitatively reveal more information, as "medical images are more than pictures, they are data."<sup>9,75-77</sup> The first radiomics study showed that pretreatment CT-based radiomics features that predict overall survival for nonsmall cell lung cancer patients performed well in an independent lung and a head and neck cancer patient cohort.<sup>10</sup> Several subsequent studies have shown that tumor radiomics features can contribute to the prediction of overall, disease-free, progression-free survival in multiple disease sites: e.g., head and neck,<sup>10,78-81</sup> lung<sup>10,11</sup> cervical,<sup>82</sup> prostate and rectal<sup>83</sup> cancer patients. Gross modo, these studies suggest that high texture values in the tumor, capturing higher tumor heterogeneity, are typically associated with a higher risk of recurrences and poorer survival.<sup>75,81</sup> Geometric features typically indicate that larger and more irregularly shaped (e.g., volume surface, bounding box) tumors are related to recurrences and worse survival rates.<sup>81</sup> Moreover, radiomic features have also been associated with pathology or molecular markers (e.g., HPV)<sup>84-87</sup> and when combined with genomic markers, they can improve prediction.<sup>88</sup> This demonstrates the potential of associating radiomics with biology and highlights the complementary effects it can have in improving the prediction of treatment outcomes.

Radiomics has also been applied for normal tissue response or toxicity, addressing the challenge that patients who receive similar radiation doses to organs can have highly variable toxicity presentations. Radiomics features of the parotid glands obtained from pretreatment CT and <sup>18</sup>F-FDG-PET images were added to the conventional dose-based xerostomia prediction model,<sup>20,89</sup> something that was independently validated.<sup>19,21</sup> This work demonstrated that high heterogeneous CT intensities and low metabolic activity in parotid gland tissue were associated with a higher probability of developing permanent late xerostomia, suggesting that larger ratios of fat-to-functional parotid gland tissue were related to a higher risk of xerostomia. This was later confirmed by the strong relation between fat-related MR-features and late xerostomia.<sup>90,91</sup> Another example, radiomics features that indicated that a large volume of high PET intensity was associated with the development of radiation pneumonitis, suggested that pretreatment subclinical inflammation may already be a risk factor for pneumonitis.<sup>22</sup> In addition, images during treatment have been shown to give useful delta radiomics features to predict toxicities.<sup>92-95</sup>

Clinical implementation of radiomics models has yet to be realized in radiotherapy. One of the major challenges that these models face is imaging and image processing variation.<sup>30,96-98</sup> While the image biomarker standardization initiative (IBSI) has produced guidelines to harmonize image processing, feature definition, and extraction,<sup>33</sup> the variability in imaging acquisition still challenges the robustness of radiomics features<sup>98,99</sup> and likely attributes to why radiomics features fail to predict treatment outcome or validate in some centers.<sup>100-104</sup> Advancements in MRI, such as quantitative modalities like Diffusion Weighted Imaging (DWI) and Dynamic Contrast Enhanced (DCE) MR imaging, can provide meaningful intensity values related to diffusion and perfusion of tissue, respectively.<sup>105-107</sup> These methods are, however, more susceptible to motion and geometric distortion and are limited by noise and resolution.<sup>108</sup> This field needs to mature to propel the field of personalized radiotherapy forward.

Radiomics combined with clinical variable models have improved prediction performance. The radiomics feature can add information to clinical variables such as age, TNM stage, and performance score but can also replace them. The latter was shown by Zhai et al., where the feature major axis length of all lymph nodes (i.e., the maximum length between pathological lymph nodes) better predicted survival than the N-stage.<sup>81</sup> This indicates the potential for quantification of clinical variables and the importance of being wary of multicollinearity (i.e., high inter-associations between the independent variables).<sup>109</sup>

Deep learning models or convolutional neural networks differentiate themselves from conventional machine learning approaches by extracting weights and features directly from the input data, such as 3D imaging and radiotherapy dose data, without predefining variables. Typically, the initial layers of these networks are convolutional layers, applying various imaging filters in sequence. The processed data is then flattened and passed through fully connected layers that perform a function analogous to variable selection.<sup>110,111</sup> Deep learning applications can be found throughout the radiotherapy workflow, from image acquisition segmentation to treatment planning and delivery.<sup>112</sup> Deep learning in radiotherapy outcome prediction is also growing rapidly, with studies focusing on using imaging data to predict tumor responses and overall survival.<sup>110,111,113-116</sup> Moreover, combining imaging and 3D dose distributions as deep learning inputs has been explored for tumor outcome and toxicity prediction.<sup>117-120</sup> Additionally, adding genomics data as e.g., obtained from genome-wide association study (GWAS) can identify genetic differences between patients related to certain toxicities, as has been shown for radiation-induced xerostomia.<sup>121</sup> Adding these may improve prediction. However, deep learning is intrinsically data-hungry, and the required enormous datasets are not readily available in radiotherapy. Even though algorithmic measures (e.g., dropout, permutations) have been implemented to enhance generalizability, this still illustrates the need to train models on collective multi-institutional datasets.

By directly learning from images, deep learning comes with the trade-off of being a "black box" compared to conventional machine learning approaches, as defined features can be more easily related to semantic concepts, such as tumor size and heterogeneity. Deep learning attention maps can provide insights into the focal areas of the deep learning network models, visually highlighting key regions on the input image.<sup>122,123</sup> This can aid the model's reliability by identifying potential biases in predictions and generating hypotheses by showing new regions associated with tumor outcome or toxicity development. The potential to link imaging and outcome variation to genome data or organoid processes is even more exciting. However, explainable AI is still in an early stage, and interpretation can be ambiguous.

## Merged Approaches

If we want to move towards a more personalized treatment strategy, we will need to adapt to model-based probability as a marker for treatment selection. It is essential to take personalized radiotherapy to the next level and to alter the radiation dose to the patient's risk, i.e., dose escalation for patients at high risk of treatment failure versus dose de-escalation for low-risk patients. For this, combining clinically derived data and knowledge of biological processes will increase the predictive potential of such models.

Although biology models can be used to describe mechanisms, show potential biomarkers, and predict the response of patients to cancer therapies, regrettably, for patient-specific response prediction most of the biology models may take weeks to provide a prediction, due to slow growth/response of the biological entity studies. Waiting for treatment until this prediction is finished is not feasible. While clinical machine learning models—once trained—take only seconds to minutes to run, they lack the capacity to prove causality in radiation effect. With explainable AI, this is attempted, but it will not surpass being hypothesis generating. However, when AI points to a potential biological process/mechanism, models of biology can show the causality of such processes through gene expression modulation. Retrospectively, biological assays can validate the predictive potential of specific biomarkers, as has been performed for organoid response testing.<sup>38-41,124</sup> If a model can predict patients' responses, single-cell multiomics can be used to show the biological processes involved. When such a process is observed and biologically validated by, for instance, knocking down the gene involved (Cinat et al.<sup>44</sup>), this process could also be assessed in patients. The stem cell genetic or biological process markers can be correlated with the response of general patients and their gene profile. An example could be the differential expression of cytokines involved in viral responses after irradiation, such as interferons (INFs)<sup>44</sup> between patients for tumor and normal tissue response. High levels of, e.g., INF- $\beta$  may improve regeneration of normal tissue.<sup>44</sup> Say that some patients have a low, and others have a high INF response after irradiation; this could be linked to image "biomarker," clinical tumor characteristics, or GWAS data. Particularly, new image biomarkers from quantitative

imaging (e.g., DWI MRI or PET imaging with novel tracers) may be identified that need compelling rationale from biology to be introduced into the clinical workflow. Via this avenue, the biology causality can be subsequently related to clinical response data in larger datasets to develop effective bioinformatics and AI/machine learning models. An approach involving high-level data analysis, bioinformatics, and artificial intelligence (AI)/machine learning is required to combine the complex and large amount of data on omics and patients-related parameters. The big data collected for a comprehensive multiomics project requires combining different data processing and integration approaches to explore the quantitative aspects (correlation analysis, multivariate factorization, and molecular network learning). Dedicated machine learning approaches (e.g., multivariable statistics, random forest, and possibly novel methodologies to be developed) need to be adequately tuned to analyze the data. For example, by applying unsupervised clustering methods, multifactorial clinical (image) biomarkers can be linked to relevant biological markers. Next, a machine learning Bayesian approach may be promising, assuming that higher irradiation doses will produce a higher response and that different radiation qualities produce a different response. It is also essential to link the biological approaches with the clinical prediction model, as clinical prediction models are trained on patients receiving the current status-quo radiotherapy fractionation schedule and prescribed dose. This means that the risk classification of patients may be altered if a different radiotherapy regimen would have been given, while this cannot be tested *in vivo*, as we can only give the primary radiation once. This could be done with biological methods, such as organoids. To effectively personalize the radiation dose, a symbiotic collaboration between the disciplines is essential.

The potential of multiomics data analysis is yet to be unraveled; in consulting with the team mentioned above and based on the diversity and quality of the data, potentially different methodologies may be followed.<sup>125</sup> This should lead to models per tissue/tumor type that, e.g., predict patients' specific responses. Multiomics data from validation experiments must be included to test further and verify the models. In the near future, this may lead to improved patient-specific prediction of tumor and normal tissue response, allowing optimization of radiotherapy and combined treatments, resulting in optimal tumor control with the best possible quality of life.

## References

- Montay-Gruel P, Meziani L, Yakkala C, et al: Expanding the therapeutic index of radiation therapy by normal tissue protection. *Br J Radiol* 92, 2019(1093):20180008
- Joiner M, Kogel Avd: *Basic clinical radiobiology*. 4th ed London: Hodder Arnold, 2009.
- Reda M, Bagley AF, Zaidan HY, et al: Augmenting the therapeutic window of radiotherapy: A perspective on molecularly targeted therapies and nanomaterials. *Radiother Oncol* 150:225-235, 2020
- Cinat D, Coppes RP, Barazzuol L: DNA damage-induced inflammatory microenvironment and adult stem cell response. *Front Cell Dev Biol* 9, 2021:729136
- Peng X, Wu Y, Brouwer U, et al: Cellular senescence contributes to radiation-induced hyposalivation by affecting the stem/progenitor cell niche. *Cell Death Dis* 11(10):854-859, 2020
- Barazzuol L, Coppes RP, van Luijk P: Prevention and treatment of radiotherapy-induced side effects. *Mol Oncol* 14(7):1538-1554, 2020
- Li J, Li L, You P, et al: Towards artificial intelligence to multi-omics characterization of tumor heterogeneity in esophageal cancer. *Semin Cancer Biol* 91:35-49, 2023
- Heller G, Fuereder T, Grandits AM, et al: New perspectives on biology, disease progression, and therapy response of head and neck cancer gained from single cell RNA sequencing and spatial transcriptomics. *Oncol Res* 32(1):1-17, 2023
- Kumar V, Gu Y, Basu S, et al: Radiomics: the process and the challenges. *Magn Reson Imaging* 30(9):1234-1248, 2012
- Aerts HJWL, Velazquez ER, Leijenaar RTH, et al: Decoding tumour phenotype by noninvasive imaging using a quantitative radiomics approach. *Nat Commun* 5:4006, 2014
- Thawani R, McLane M, Beig N, et al: Radiomics and radiogenomics in lung cancer: A review for the clinician. *Lung Cancer* 115:34-41, 2018
- Zhai T, Langendijk JA, van Dijk LV, et al: The prognostic value of CT-based image-biomarkers for head and neck cancer patients treated with definitive (chemo-)radiation. *Oral Oncol* 95:178-186, 2019
- Vallières M, Freeman CR, Skamene SR, et al: A radiomics model from joint FDG-PET and MRI texture features for the prediction of lung metastases in soft-tissue sarcomas of the extremities. *Phys Med Biol* 60 (14):5471-5496, 2015
- Xiong J, Yu W, Ma J, et al: The role of PET-based radiomic features in predicting local control of esophageal cancer treated with concurrent chemoradiotherapy. *Sci Rep* 8(1):9902, 2018
- Cook GJR, Siddique M, Taylor BP, et al: Radiomics in PET: Principles and applications. *Clin Translat Imag* 2(3):269-276, 2014
- Carvalho S, Leijenaar RTH, Troost EGC, et al: 18F-fluorodeoxyglucose positron-emission tomography (FDG-PET)-Radiomics of metastatic lymph nodes and primary tumor in non-small cell lung cancer (NSCLC) - a prospective externally validated study. *PLoS One* 13, 2018(3):e0192859
- Dissaux G, Visvikis D, Da-Ano R, et al: Pretreatment (18)F-FDG PET/CT radiomics predict local recurrence in patients treated with stereotactic body radiotherapy for early-stage non-small cell lung cancer: A multicentric study. *J Nucl Med* 61(6):814-820, 2020
- Xie P, Yue J, Fu Z, et al: Prognostic value of 18F-FDG PET/CT before and after radiotherapy for locally advanced nasopharyngeal carcinoma. *Ann Oncol* 21(5):1078-1082, 2010
- Wilkie JR, Mierzwa ML, Casper KA, et al: Predicting late radiation-induced xerostomia with parotid gland PET biomarkers and dose metrics. *Radiother Oncol* 148:30-37, 2020
- van Dijk LV, Noordzij W, Brouwer CL, et al: (18)F-FDG PET image biomarkers improve prediction of late radiation-induced xerostomia. *Radiother Oncol* 126(1):89-95, 2018
- Li Y, Sijtsma NM, de Vette SPM, et al: Validation of the (18)F-FDG PET image biomarker model predicting late xerostomia after head and neck cancer radiotherapy. *Radiother Oncol* 180, 2023:109458
- Castillo R, Pham N, Ansari S, et al: Pre-radiotherapy FDG PET predicts radiation pneumonitis in lung cancer. *Radiat Oncol* 9:74, 2014
- Castillo R, Pham N, Castillo E, et al: Pre-radiation therapy fluorine 18 fluorodeoxyglucose PET helps identify patients with esophageal cancer at high risk for radiation pneumonitis. *Radiology* 275(3):822-831, 2015
- Srinivasan A, Mohan S, Mukherji SK: Biologic imaging of head and neck cancer: the present and the future. *AJNR Am J Neuroradiol* 33 (4):586-594, 2012
- Zhou N, Guo T, Zheng H, et al: Apparent diffusion coefficient histogram analysis can evaluate radiation-induced parotid damage and predict late xerostomia degree in nasopharyngeal carcinoma. *Oncotarget* 8(41):70226-70238, 2017
- Dirix P, De Keyzer F, Vandecaveye V, et al: Diffusion-weighted magnetic resonance imaging to evaluate major salivary gland function before and after radiotherapy. *Int J Radiat Oncol Biol Phys* 71 (5):1365-1371, 2008

27. Vandecaveye V, Dirix P, De Keyzer F, et al: Predictive value of diffusion-weighted magnetic resonance imaging during chemoradiotherapy for head and neck squamous cell carcinoma. *Eur Radiol* 20(7):1703-1714, 2010
28. Vandecaveye V, Dirix P, De Keyzer F, et al: Diffusion-weighted magnetic resonance imaging early after chemoradiotherapy to monitor treatment response in head-and-neck squamous cell carcinoma. *Int J Radiat Oncol Biol Phys* 82(3):1098-1107, 2012
29. Wong AJ, Kanwar A, Mohamed AS, et al: Radiomics in head and neck cancer: from exploration to application. *Transl Cancer Res* 5(4):371-382, 2016
30. Shafiq-UL-Hassan M, Zhang GG, Latifi K, et al: Intrinsic dependencies of CT radiomics features on voxel size and number of gray levels. *Med Phys* 44(3):1050-1062, 2017
31. Park JE, Park SY, Kim HJ, et al: Reproducibility and generalizability in radiomics modeling: Possible strategies in radiologic and statistical perspectives. *Korean J Radiol* 20(7):1124-1137, 2019
32. Shiri I, Abdollahi H, Shaysteh S, et al: Test-retest reproducibility and robustness analysis of recurrent glioblastoma MRI radiomics texture features. *Iranian Journal of Radiology* 14, 2017:e48035
33. Zwanenburg A, Vallières M, Abdalah MA, et al: The image biomarker standardization initiative: Standardized quantitative radiomics for high-throughput image-based phenotyping. *Radiology* 295(2):328-338, 2020
34. Eke I, Cordes N: Radiobiology goes 3D: How ECM and cell morphology impact on cell survival after irradiation. *Radiotherapy and Oncology* 99(3):271-278, 2011
35. Nagle PW, Hosper NA, Ploeg EM, et al: The in vitro response of tissue stem cells to irradiation with different linear energy transfers. *International Journal of Radiation Oncology Biology Physics* 95(1):103-111, 2016
36. Huch M, Gehart H, Van Boxtel R, et al: Long-term culture of genome-stable bipotent stem cells from adult human liver. *Cell* 160(1-2):299-312, 2015
37. Nagle PW, Plukker JTM, Muijs CT, et al: Patient-derived tumor organoids for prediction of cancer treatment response. *Semin Cancer Biol* 53:258-264, 2018
38. Soto-Gamez A, Gunawan JP, Barazzuol L, et al: Organoid-based personalized medicine: from tumor outcome prediction to autologous transplantation. *Stem Cells* 42(6):499-508, 2024
39. Sakshaug BC, Folkesson E, Haukaas TH, et al: Systematic review: predictive value of organoids in colorectal cancer. *Sci Rep* 13(1):18124-18128, 2023
40. Millen R, De Kort WWB, Koomen M, et al: Patient-derived head and neck cancer organoids allow treatment stratification and serve as a tool for biomarker validation and identification. *Med* 4(5):290-310.e12, 2023
41. Shukla HD, Dukic T, Roy S, et al: Pancreatic cancer derived 3D organoids as a clinical tool to evaluate the treatment response. *Front Oncol* 12, 2023:1072774
42. Nagle PW, Hosper NA, Barazzuol L, et al: Lack of DNA damage response at low radiation doses in adult stem cells contributes to organ dysfunction. *Clin Cancer Res* 24(24):6583-6593, 2018
43. Nowrouzi A, Sertorio MG, Akbarpour M, et al: Personalized assessment of normal tissue radiosensitivity via transcriptome response to photon, proton and carbon irradiation in patient-derived human intestinal organoids. *Cancers (Basel)* 12(2):469, 2020. <https://doi.org/10.3390/cancers12020469>
44. Cinat Davide, van der Wal Ryan, Baanstra Mirjam, et al: Derepression of transposable elements enhances interferon beta signaling and stem/progenitor cell activity after proton irradiation. *bioRxiv* 2024. <https://doi.org/10.1101/2024.02.14.580306>
45. Rehwinkel J, Gack MU: RIG-I-like receptors: their regulation and roles in RNA sensing. *Nat Rev Immunol* 20(9):537-551, 2020
46. Onomoto K, Onoguchi K, Yoneyama M: Regulation of RIG-I-like receptor-mediated signaling: interaction between host and viral factors. *Cell Mol Immunol* 18(3):539-555, 2021
47. Liu X, Liu Z, Wu Z, et al: Resurrection of endogenous retroviruses during aging reinforces senescence. *Cell* 186(2):287-304.e26, 2023
48. Workman MJ, Mahe MM, Trisno S, et al: Engineered human pluripotent-stem-cell-derived intestinal tissues with a functional enteric nervous system. *Nat Med* 23(1):49-59, 2017
49. Birey F, Andersen J, Makinson CD, et al: Assembly of functionally integrated human forebrain spheroids. *Nature* 545(7652):54-59, 2017
50. Soto-Gamez AA, van Es M, Hageman E, et al: Mesenchymal stem cell-derived HGF attenuates radiation-induced senescence in salivary glands via compensatory proliferation. *Radiother Oncol* 190, 2024:109984
51. IAD Graaf, Groothuis GM, Olinga P: Precision-cut tissue slices as a tool to predict metabolism of novel drugs. *Expert Opin Drug Metab Toxicol* 3(6):879-898, 2007
52. Stenzel PJ, Hörner N, Foersch S, et al: Nivolumab reduces PD1 expression and alters density and proliferation of tumor infiltrating immune cells in a tissue slice culture model of renal cell carcinoma. *Cancers (Basel)* 13(18):4511, 2021. <https://doi.org/10.3390/cancers13184511>
53. Sesink A, Becerra M, Ruan J, et al: The AsiDNA™ decoy mimicking DSBs protects the normal tissue from radiation toxicity through a DNA-PK/p53/p21-dependent G1/S arrest. *NAR Cancer* 6(1):zcae011, 2024
54. Chuprin J, Buettner H, Seedhom MO, et al: Humanized mouse models for immuno-oncology research. *Nat Rev Clin Oncol* 20(3):192-206, 2023
55. Cogels MM, Rouas R, Ghanem GE, et al: Humanized mice as a valuable pre-clinical model for cancer immunotherapy research. *Front Oncol* 11, 2021:784947
56. Yu D, Yang P, Lu X, et al: Single-cell RNA sequencing reveals enhanced antitumor immunity after combined application of PD-1 inhibitor and Shenmai injection in non-small cell lung cancer. *Cell Commun Signal* 21(1):169, 2023
57. Larsen CG, Jensen DH, Carlander AF, et al: Novel nomograms for survival and progression in HPV+ and HPV- oropharyngeal cancer: a population-based study of 1,542 consecutive patients. *Oncotarget* 7(44):71761-71772, 2016
58. van Dijk LV, Mohamed AS, Ahmed S, et al: Head and neck cancer predictive risk estimator to determine control and therapeutic outcomes of radiotherapy (HNC-PREDICTOR): development, international multi-institutional validation, and web implementation of clinic-ready model-based risk stratification for head and neck cancer. *Eur J Cancer* 178:150-161, 2023
59. Fakhry C, Zhang Q, Nguyen-Tân PF, et al: Development and validation of nomograms predictive of overall and progression-free survival in patients with oropharyngeal cancer. *J Clin Oncol* 35(36):4057-4065, 2017
60. Hoesseini A, van Leeuwen N, Offerman MPJ, et al: Predicting survival in head and neck cancer: External validation and update of the prognostic model OncologIQ in 2189 patients. *Head Neck* 43(8):2445-2456, 2021
61. Oberije C, De Ruyscher D, Houben R, et al: A validated prediction model for overall survival from stage iii non-small cell lung cancer: toward survival prediction for individual patients. *Int J Radiat Oncol Biol Phys* 92(4):935-944, 2015
62. Tanadini-Lang S, Rieber J, Filippi AR, et al: Nomogram based overall survival prediction in stereotactic body radiotherapy for oligo-metastatic lung disease. *Radiother Oncol* 123(2):182-188, 2017
63. Zelefsky MJ, Kattan MW, Fearn P, et al: Pretreatment nomogram predicting ten-year biochemical outcome of three-dimensional conformal radiotherapy and intensity-modulated radiotherapy for prostate cancer. *Urology* 70(2):283-287, 2007
64. Yang X, Chen A, PourNejatian N, et al: A large language model for electronic health records. *NPJ Digit Med* 5(1):194, 2022
65. Balachandran VP, Gonen M, Smith JJ, et al: Nomograms in oncology: more than meets the eye. *Lancet Oncol* 16(4):173, 2015
66. Baatenburg de Jong RJ, Hermans J, Molenaar J, et al: Prediction of survival in patients with head and neck cancer. *Head Neck* 23(9):718-724, 2001
67. Brodin NP, Kabarriti R, Garg MK, et al: Systematic review of normal tissue complication models relevant to standard fractionation radiation



- therapy of the head and neck region published after the QUANTEC reports. *Int J Radiat Oncol Biol Phys* 100(2):391-407, 2018
68. Beetz I, Schilstra C, van der Schaaf A, et al: NTCP models for patient-rated xerostomia and sticky saliva after treatment with intensity modulated radiotherapy for head and neck cancer: the role of dosimetric and clinical factors. *Radiother Oncol* 105(1):101-106, 2012
  69. Appelt AL, Vogelius IR, Farr KP, et al: Towards individualized dose constraints: Adjusting the QUANTEC radiation pneumonitis model for clinical risk factors. *Acta Oncol* 53(5):605-612, 2014
  70. Schaake W, van der Schaaf A, van Dijk LV, et al: Normal tissue complication probability (NTCP) models for late rectal bleeding, stool frequency and fecal incontinence after radiotherapy in prostate cancer patients. *Radiother Oncol* 119(3):381-387, 2016
  71. Langendijk JA, Lambin P, De Ruyscher D, et al: Selection of patients for radiotherapy with protons aiming at reduction of side effects: the model-based approach. *Radiother Oncol* 107(3):267-273, 2013
  72. Witte MG, van der Geer J, Schneider C, et al: IMRT optimization including random and systematic geometric errors based on the expectation of TCP and NTCP. *Med Phys* 34(9):3544-3555, 2007
  73. Thomas E, Chapet O, Kessler ML, et al: Ten Haken RK. Benefit of using biologic parameters (EUD and NTCP) in IMRT optimization for treatment of intrahepatic tumors. *Int J Radiat Oncol Biol Phys* 62(2):571-578, 2005
  74. Deasy JO, Mayo CS, Orton CG: Treatment planning evaluation and optimization should be biologically and not dose/volume based. *Med Phys* 42(6):2753-2756, 2015
  75. Yip SSF, Liu Y, Parmar C, et al: Associations between radiologist-defined semantic and automatically computed radiomic features in non-small cell lung cancer. *Sci Rep* 7(1):3519, 2017
  76. Gillies RJ, Kinahan PE, Hricak H: Radiomics: Images are more than pictures, they are data. *Radiology* 278(2):563-577, 2016
  77. Lambin P, Rios-Velazquez E, Leijenaar R, et al: Radiomics: extracting more information from medical images using advanced feature analysis. *Eur J Cancer* 48(4):441-446, 2012
  78. Abgral R, Keromnes N, Robin P, et al: Prognostic value of volumetric parameters measured by 18F-FDG PET/CT in patients with head and neck squamous cell carcinoma. *Eur J Nucl Med Mol Imaging* 41(4):659-667, 2014
  79. Koyasu S, Nakamoto Y, Kikuchi M, et al: Prognostic value of pretreatment 18F-FDG PET/CT parameters including visual evaluation in patients with head and neck squamous cell carcinoma. *AJR Am J Roentgenol* 202(4):851-858, 2014
  80. Alluri KC, Tahari AK, Wahl RL, et al: Prognostic value of FDG PET metabolic tumor volume in human papillomavirus-positive stage III and IV oropharyngeal squamous cell carcinoma. *AJR Am J Roentgenol* 203(4):897-903, 2014
  81. Zhai T, van Dijk LV, Huang B, et al: Improving the prediction of overall survival for head and neck cancer patients using image biomarkers in combination with clinical parameters. *Radiother Oncol* 124(2):256-262, 2017
  82. Lucia F, Visvikis D, Desseroit M, et al: Prediction of outcome using pretreatment (18)F-FDG PET/CT and MRI radiomics in locally advanced cervical cancer treated with chemoradiotherapy. *Eur J Nucl Med Mol Imaging* 45(5):768-786, 2018
  83. Yi X, Pei Q, Zhang Y, et al: MRI-based radiomics predicts tumor response to neoadjuvant chemoradiotherapy in locally advanced rectal cancer. *Front Oncol* 9:552, 2019
  84. Bogowicz M, Riesterer O, Ikenberg K, et al: Computed tomography radiomics predicts hpv status and local tumor control after definitive radiochemotherapy in head and neck squamous cell carcinoma. *Int J Radiat Oncol Biol Phys* 99(4):921-928, 2017
  85. Corredor G, Bharadwaj S, Pathak T, et al: A review of AI-based radiomics and computational pathology approaches in triple-negative breast cancer: Current applications and perspectives. *Clin Breast Cancer* 23(8):800-812, 2023
  86. Bagher-Ebadian H, Lu M, Siddiqui F, et al: Application of radiomics for the prediction of HPV status for patients with head and neck cancers. *Med Phys* 47(2):563-575, 2020
  87. Tang C, Hobbs B, Amer A, et al: Development of an Immune-Pathology Informed Radiomics Model for Non-Small Cell Lung Cancer. *Scient Rep* 8(1):1922, 2018
  88. Li S, Zhou B: A review of radiomics and genomics applications in cancers: the way towards precision medicine. *Radiat Oncol* 17(1):217, 2022
  89. van Dijk LV, Brouwer CL, van der Schaaf A, et al: CT image biomarkers to improve patient-specific prediction of radiation-induced xerostomia and sticky saliva. *Radiother Oncol* 122(2):185-191, 2017
  90. van Dijk LV, Thor M, Steenbakkers RJHM, et al: Parotid gland fat related magnetic resonance image biomarkers improve prediction of late radiation-induced xerostomia. *Radiother Oncol* 128(3):459-466, 2018
  91. Sheikh K, Lee SH, Cheng Z, et al: Predicting acute radiation induced xerostomia in head and neck cancer using MR and CT Radiomics of parotid and submandibular glands. *Radiat Oncol* 14(1):131-134, 2019
  92. Rosen BS, Hawkins PG, Polan DF, et al: Early changes in serial CBCT-measured parotid gland biomarkers predict chronic xerostomia after head and neck radiation therapy. *Int J Radiat Oncol Biol Phys* 102(4):1319-1329, 2018
  93. Wu H, Chen X, Yang X, et al: Early prediction of acute xerostomia during radiation therapy for head and neck cancer based on texture analysis of daily CT. *Int J Radiat Oncol Biol Phys* 102(4):1308-1318, 2018
  94. van Dijk LV, Brouwer CL, van der Laan HP, et al: Geometric image biomarker changes of the parotid gland are associated with late xerostomia. *Int J Radiat Oncol Biol Phys* 99(5):1101-1110, 2017
  95. Cunliffe A, Armato SG3, Castillo R, et al: Lung texture in serial thoracic computed tomography scans: correlation of radiomics-based features with radiation therapy dose and radiation pneumonitis development. *Int J Radiat Oncol Biol Phys* 91(5):1048-1056, 2015
  96. He L, Huang Y, Ma Z, et al: Effects of contrast-enhancement, reconstruction slice thickness and convolution kernel on the diagnostic performance of radiomics signature in solitary pulmonary nodule. *Sci Rep* 6:34921, 2016
  97. Larue RTHM, van Timmeren JE, de Jong EEC, et al: Influence of gray level discretization on radiomic feature stability for different CT scanners, tube currents and slice thicknesses: a comprehensive phantom study. *Acta Oncol* 56(11):1544-1553, 2017
  98. Mackin D, Fave X, Zhang L, et al: Measuring computed tomography scanner variability of radiomics features. *Invest Radiol* 50(11):757-765, 2015
  99. Ger RB, Zhou S, Chi PM, et al: Comprehensive Investigation on Controlling for CT Imaging Variabilities in Radiomics Studies. *Sci Rep* 8(1):13047, 2018
  100. Ger RB, Zhou S, Elgohari B, et al: Radiomics features of the primary tumor fail to improve prediction of overall survival in large cohorts of CT- and PET-imaged head and neck cancer patients. *PLoS One* 14, 2019(9):e0222509
  101. Wang Y, Zhao H, Zhang Z, et al: MR imaging prediction of local control of nasopharyngeal carcinoma treated with radiation therapy and chemotherapy. *Br J Radiol* 87, 2014(1039):20130657
  102. Loo CE, Straver ME, Rodenhuis S, et al: Magnetic resonance imaging response monitoring of breast cancer during neoadjuvant chemotherapy: relevance of breast cancer subtype. *J Clin Oncol* 29(6):660-666, 2011
  103. Wahid KA, He R, McDonald BA, et al: Intensity standardization methods in magnetic resonance imaging of head and neck cancer. *Phys Imaging Radiat Oncol* 20:88-93, 2021
  104. Nyúl LG, Udupa JK, Zhang X: New variants of a method of MRI scale standardization. *IEEE Trans Med Imaging* 19(2):143-150, 2000
  105. Rosenkrantz AB, Mendiratta-Lala M, Bartholmai BJ, et al: Clinical utility of quantitative imaging. *Acad Radiol* 22(1):33-49, 2015. Jan
  106. Abramson RG, Burton KR, Yu JJ, et al: Methods and challenges in quantitative imaging biomarker development. *Acad Radiol* 22(1):25-32, 2015
  107. Koh D, Collins DJ: Diffusion-weighted MRI in the body: applications and challenges in oncology. *AJR Am J Roentgenol* 188(6):1622-1635, 2007
  108. Schmidt MA, Payne GS: Radiotherapy planning using MRI. *Phys Med Biol* 60(22):323, 2015
  109. Welch ML, McIntosh C, Haibe-Kains B, et al: Vulnerabilities of radiomic signature development: The need for safeguards. *Radiother Oncol* 130:2-9, 2019 Jan

110. Litjens G, Kooi T, Bejnordi BE, et al: A survey on deep learning in medical image analysis. *Med Image Anal* 42:60-88, 2017
111. LeCun Y, Bengio Y, Hinton G: Deep learning. *Nature* 521(7553):436-444, 2015
112. Meyer P, Noblet V, Mazzara C, et al: Survey on deep learning for radiotherapy. *Comput Biol Med* 98:126-146, 2018
113. Diamant A, Chatterjee A, Vallières M, et al: Deep learning in head & neck cancer outcome prediction. *Sci Rep* 9(1):2764, 2019
114. Lou B, Doken S, Zhuang T, et al: An image-based deep learning framework for individualizing radiotherapy dose. *Lancet Digit Health* 1(3): e136-e147, 2019
115. Naser MA, Wahid KA, Mohamed ASR, et al: Progression free survival prediction for head and neck cancer using deep learning based on clinical and PET/CT imaging data. *Head Neck Tumor Segm Chall* 13209:287-299, 2021
116. Nie D, Lu J, Zhang H, et al: Multi-channel 3D deep feature learning for survival time prediction of brain tumor patients using multi-modal neuroimages. *Sci Rep* 9(1):1103-1109, 2019
117. Appelt AL, Elhaminia B, Gooya A, et al: Deep learning for radiotherapy outcome prediction using dose data - a review. *Clin Oncol (R Coll Radiol)* 34(2):e87-e96, 2022
118. Men K, Geng H, Zhong H, et al: A deep learning model for predicting xerostomia due to radiation therapy for head and neck squamous cell carcinoma in the RTOG 0522 clinical trial. *Int J Radiat Oncol Biol Phys* 105(2):440-447, 2019
119. de Vette SP, Chu H, Neh H, et al: OC-0088 Deep learning NTCP model for late dysphagia based on 3D dose, CT and segmentations. *Radiother Oncol* 182:S51-S52, 2023
120. Chu H, de Vette SP, Neh H, et al: OC-0449 Deep learning based NTCP-modelling using 3D-information for predicting late xerostomia. *Radiother Oncol* 182:S352-S354, 2023
121. Naderi E, Crijns APG, Steenbakkens RJHM, et al: A two-stage genome-wide association study of radiation-induced acute toxicity in head and neck cancer. *J Transl Med* 19(1):481, 2021
122. Grad-cam: Visual explanations from deep networks via gradient-based localization. In: *Proceedings of the IEEE international conference on computer vision*; 2017
123. Grad-cam: Generalized gradient-based visual explanations for deep convolutional networks. In: *2018 IEEE winter conference on applications of computer vision (WACV)*, IEEE; 2018.
124. Nagle PW, Coppes RP: Current and Future Perspectives of the Use of Organoids in Radiobiology. *Cells* 9(12):2649, 2020. <https://doi.org/10.3390/cells9122649>
125. Babu M, Snyder M: Multi-omics profiling for health. *Mol Cell Proteomics* 22, 2023(6):100561