Production of a Class II Two-Component Lantibiotic of
Streptococcus pneumoniae Using the Class I Nisin
Synthetic Machinery and Leader Sequence

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Recent studies showed that the nisin modification machinery can successfully dehydrate serines and threonines and introduce lantionine rings in small peptides that are fused to the nisin leader sequence. This opens up exciting possibilities to produce and engineer larger antimicrobial peptides in vivo. Here we demonstrate the exploitation of the class I nisin production machinery to generate, modify, and secrete biologically active, previously not-yet-isolated and -characterized class II two-component lantibiotics that have no sequence homology to nisin. The nisin synthesis machinery, composed of the modification enzymes NisB and NisC and the transporter NisT, was used to modify and secrete a putative two-component lantibiotic of Streptococcus pneumoniae. This was achieved by genetically fusing the propeptide-encoding sequences of the spr1765 (pneA1) and spr1766 (pneA2) genes to the nisin leader-encoding sequence. The chimeric prepeptides were secreted out of Lactococcus lactis, purified by cation exchange fast protein liquid chromatography, and further characterized. Mass spectrometry analyses demonstrated the presence and partial localization of multiple dehydrated serines and/or threonines and (methyl)lanthionines in both peptides. Moreover, after cleavage of the leader peptide from the prepeptides, both modified propeptides displayed antimicrobial activity against Micrococcus flavus. These results demonstrate that the nisin synthetase machinery can be successfully used to modify and produce otherwise difficult to obtain antimicrobially active lantibiotics.

Small antimicrobial peptides produced by Gram-positive bacteria are named bacteriocins. One group of bacteriocins, the nonlantibiotics, comprises peptides that do not require modification for their antimicrobial activity (46). Members of another group, the lantibiotics, require posttranslational modifications to acquire biological activity (11, 47). Lantibiotics are produced as inactive prepeptides, consisting of an N-terminal leader peptide and a C-terminal propeptide part. Most of the serine and threonine residues of the propeptide are dehydrated to dehydroalanine (Dha) and dehydrobutyrine (Dhb), respectively, by LanB- or LanM-type enzymes (“Lan” is a general abbreviation for proteins involved in lantibiotic biosynthesis). LanC or LanM enzymes can subsequently couple these dehydroresidues to cysteines, thus forming a (methyl)lanthionine ring. After the leader peptide is removed from the prepeptide by the extracellular LanP or transmembrane LanT proteins, the active lantibiotic is released. The immunity against the produced lantibiotics is provided by the LanI proteins, the active lantibiotic is released. The immunity against the produced lantibiotics is provided by the LanI proteins, the active lantibiotic is released. The immunity against the produced lantibiotics is provided by the LanI proteins, the active lantibiotic is released. The immunity against the produced lantibiotics is provided by the LanI proteins, the active lantibiotic is released. The immunity against the produced lantibiotics is provided by the LanI proteins, the active lantibiotic is released.

To understand the potential of lantibiotics as antimicrobial agents against Gram-positive bacteria, it is necessary to first understand the molecular mechanisms of their biosynthesis and secretion. This understanding will enable the development of new therapeutic strategies to combat drug-resistant bacterial infections. Understanding the biosynthesis and secretion of lantibiotics is essential for developing new therapeutic strategies to combat drug-resistant bacterial infections. Understanding the biosynthesis and secretion of lantibiotics is essential for developing new therapeutic strategies to combat drug-resistant bacterial infections. Understanding the biosynthesis and secretion of lantibiotics is essential for developing new therapeutic strategies to combat drug-resistant bacterial infections. Understanding the biosynthesis and secretion of lantibiotics is essential for developing new therapeutic strategies to combat drug-resistant bacterial infections. Understanding the biosynthesis and secretion of lantibiotics is essential for developing new therapeutic strategies to combat drug-resistant bacterial infections.
cently, it was shown that designed hexapeptides and nonlantibiotic peptides fused to the leader peptide of nisin could be successfully modified by NisB and NisC and exported out of *L. lactis* via NisT (16, 20, 38). The discovery that the lantibiotic modification enzymes LanB, LanC, and LanM possess rather low substrate specificities brings a new opportunity to use them as a tool to improve the stability and activity of peptides potentially valuable for medical applications (2, 16, 39).

Here, we present the successful application of the nisin expression/modification system to produce, modify, and secrete silent lantibiotics, i.e., those for which production conditions are not known, that otherwise would be difficult to obtain from their natural sources.

### MATERIALS AND METHODS

**Bacterial strains and growth conditions.** Strains and plasmids used in this study are listed in Table 1. Strains were stored in 10% (vol/vol) glycerol at 30°C in GM17 or minimal medium (39) supplemented with 5 μg/ml chloramphenicol. *S. aureus* RN6390B, *S. mitis* NYCC10712, *E. faecalis* V583, and *M. flavus* NIZO B423 were purchased from Pepscan Lelystad NL. Peptides were purified to homogeneity with matrix-assisted laser desorption ionization–time of flight (MALDI-TOF) mass spectrometry. Importantly, the modified peptides were dehydrated multiple times, as shown by their N-terminal sequences. The modified peptides were analyzed with MALDI-TOF mass spectrometry and used for screening of antimicrobial activity.

**Expression and purification of microbially produced peptides** were performed as follows. Overnight cultures of *L. lactis* NZ9000 containing pIL3BTC and a chimeric peptide expression plasmid, namely, pNZnisA-E3, pNZnisE3-1765, or pNZnisE3-1766, in GM17 were diluted 1:50 in minimal medium containing appropriate antibiotics and 0.5 ng/ml nisin (Sigma) for induction. Cultures were grown for 24 h at 30°C. Subsequently, supernatants were separated from cells by centrifugation. Next, supernatants were filtered through a 0.2-μm filter (Millipore). Prior to purification on a 5-ml Hitrap SP cation exchange column (GE Healthcare) using fast protein liquid chromatography (FPLC), an Akta purifier (Amersham Biosciences), supernatants were diluted 1:1 with a 100 mM lactic acid solution and filtered through 0.2-μm filters. After passage of supernatant through a column, unbound compounds were washed away with 100 mM lactic acid. Elution was performed with 1 M NaCl in 50 mM lactic acid solution. The fractions containing prepeptides were concentrated and desalted with 50 mM Tris-HCl of pH 5.5 on Microcon columns (Millipore). Intact prepeptides and peptides without a leader sequence were analyzed with MALDI-TOF mass spectrometry and used for screening of antimicrobial activity.

**Terminal sequence removal.** The N-terminal sequence from the FPLC-purified prepeptides prePneA1 and prePneA2 was removed by trypsin. Prepeptides were incubated for 2 h at 37°C with 20 μg/ml trypsin in 100 mM Tris-HCl buffer (pH 8) containing 10 mM CaCl₂. Alternatively, to remove the leader from prePneA2, 180 μl of the prepeptide was incubated for 30 min at 37°C with 20 μl

### TABLE 1. Strains and plasmids used in this study

<table>
<thead>
<tr>
<th>Strain/plasmid</th>
<th>Description</th>
<th>Reference or source</th>
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<tr>
<td><em>S. pneumoniae</em> R6</td>
<td>D39 (Δ<em>pepS2</em> 2538–9862) with increased transformation efficiency</td>
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<tr>
<td><em>L. lactis</em> NZ9000</td>
<td>MG1363 Δ<em>pepN</em>:nisRK</td>
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<td>42</td>
</tr>
<tr>
<td><em>S. mitis</em> NYCC10712</td>
<td>Lab collection</td>
<td>42</td>
</tr>
<tr>
<td><em>E. faecalis</em> V583</td>
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<td></td>
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<tr>
<td><em>M. flavus</em> NIZO B423</td>
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Plasmids

<table>
<thead>
<tr>
<th>Plasmid</th>
<th>Description</th>
<th>Reference or source</th>
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<tbody>
<tr>
<td>pIL3BTC</td>
<td><em>nisBTC</em>, encoding nisin modification machinery; Ery&lt;sup&gt;a&lt;/sup&gt;</td>
<td>39</td>
</tr>
<tr>
<td>pNZ8048</td>
<td>Nisin-inducible <em>PnisA</em>: Cm&lt;sup&gt;b&lt;/sup&gt;</td>
<td>7</td>
</tr>
<tr>
<td>pNG8048E</td>
<td>Nisin-inducible <em>PnisA</em>, pNZ8048 derivative containing Ery&lt;sup&gt;a&lt;/sup&gt; gene to facilitate cloning; Cm&lt;sup&gt;b&lt;/sup&gt;</td>
<td>Lab collection</td>
</tr>
<tr>
<td>pNZnisA-E3</td>
<td><em>nisA</em>, encoding nisin</td>
<td>19</td>
</tr>
<tr>
<td>pNZNisE3-1765</td>
<td>Contains <em>nisA</em> gene and SPR1765 gene</td>
<td>This work</td>
</tr>
<tr>
<td>pNZNisE3-1766</td>
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<td>This work</td>
</tr>
<tr>
<td>pNZNisE3-1765</td>
<td>Contains a part of <em>nisA</em> gene which encodes leader peptide of nisin and leaderless part of SPR1765 gene fused in frame</td>
<td>This work</td>
</tr>
<tr>
<td>pNZNisE3-1766</td>
<td>Contains a part of <em>nisA</em> gene which encodes leader peptide of nisin and leaderless part of SPR1766 gene fused in frame</td>
<td>This work</td>
</tr>
<tr>
<td>pNGSspr1765-1766</td>
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<sup>a</sup> Dutch Institute of Dairy Research.

<sup>b</sup> Ery<sup>a</sup>, erythromycin resistance; Cm<sup>b</sup>, chloramphenicol resistance.
of 0.5 M phosphate buffer (pH 7.4) and with 10 μl of leucine aminopeptidase (Sigma; suspension in 3.5 M ammonium sulfate).

Mass spectrometry analysis. To investigate whether chimeric peptides possess free cysteine residues, reactions with 1-cyano-4-dimethylaminopyridinium tetrafluoroborate (CDAP) were performed. To obtain higher mass spectrum resolutions with MALDI-TOF, both prepeptides and propeptides, prior to CDAP treatment, were purified on a Hewlett Packard 1050 HPLC apparatus using a Jupiter Proteo C18 (4-μm, 90-Å, 250- by 4.6-mm) column. Reverse-phase purification was used with a gradient of 10% to 40% acetonitrile in purified water. All buffers contained 0.1% trifluoroacetic acid (TFA). The reactions with CDAP were performed as described previously (16, 35). Briefly, the pH of vacuum-dried trypsinated and nontrypsinated peptides and a control peptide, termed NisB2 (H-CRYTDKPHHRLIK-OH), resuspended in 16 μl MilliQ was adjusted to 2 or 3 with 0.1% TFA. Prior to treatment with CDAP, 1 μl of a 100 mg/ml concentration of the reducing agent triscarboxyethyl phosphine (TCEP) was added to each mixture, and a reaction was carried out for 5 min at room temperature. Subsequently, 2 μl of 100 mg/ml CDAP was added to the mixtures, and incubation followed for 15 min at room temperature.

Analysis was performed essentially as described before (19). Briefly, ZipTips (C18, ZipTip; Millipore) were wetted with 100% acetonitrile and washed with 0.1% TFA. Subsequently, the supernatant containing the peptides was mixed with 0.1% TFA and applied to a ZipTip. Peptides that were bound to the column were washed with 0.2% TFA and eluted with 50% acetonitrile and 0.1% TFA. The eluent was mixed in a ratio of 1:1 with 10 mg/ml a-cyano-4-hydroxycinnamic acid (matrix). A total of 1.5 μl of this prepared mixture was spotted on the target and allowed to dry. Mass spectra were recorded with a Voyager-DE Pro (Applied Biosystems) MALDI-TOF. In order to increase the sensitivity, external calibration was applied with six different peptides (protein MALDI-MS calibration kit; Sigma).

Amino acid sequence alignment. Amino acid sequence alignment of nisin with pneumococcins A1 and A2 (SPR1765 and SPR1766, respectively) was performed with Clustal W (25), a program for multiple sequence alignments. The peptide sequences were derived from the NCBI database.

Electrophoresis. Prepeptides and mature peptides were analyzed on Tris-Tricine gels (44) and stained with Coomassie (Fermentas).

Peptide concentration determination. Peptide concentrations were determined using the DC protein assay of Bio-Rad. HPLC-purified nisin was used as a standard.

MIC determination. MIC assays for M. flarus, S. pneumoniae, E. faecalis, S. aureus, and S. mitis were performed in 96-well microtiter plates in GM17. The assay was performed as follows. Overnight cultures of the above-mentioned strains were diluted 1:50, and growth was continued to an optical density at 600 nm (OD600) of 0.2. Subsequently, 150-μl cultures were mixed with 50 μl of appropriate medium and various concentrations of peptides. The microtiter plates were incubated in a GENios (TECAN Benelux) at a suitable temperature for overnight growth of the strain, and the OD600 was measured every 30 min. The MIC values were determined at the time when the cells without antimicrobial substance reached half of the maximal optical density. MICs were calculated from the lowest concentration of the antimicrobial substance that was able to inhibit the growth of the tested strain. All the susceptibility assays were performed in triplicate at least.

RESULTS

In silico analysis of the putative two-peptide lantibiotic-like cluster from S. pneumoniae. After in silico analysis of 11 putative bacteriocin genes, identified by BAGEIL (5), of S. pneumoniae R6 and their adjacent open reading frames (ORFs), we selected two of them, i.e., spr1765 and spr1766. In silico analysis of these two genes, as well as their nine adjacent ORFs, which most likely constitute a single cluster (Fig. 1), indicated that SPR1765 and SPR1766 belong to the class II two-component lantibiotics. We here propose to name SPR1765 and SPR1766 pneumococcins A1 and A2 (PneA1 and PneA2), respectively. So far there are no experimental data indicating that these bacteriocin-like peptides are expressed under laboratory growth conditions or in or under any other growth media or conditions we have tried (data not shown). In silico analysis showed that the nine adjacent genes (Fig. 1) are likely involved in pneumococcin A1 and A2 modification, transport, processing, and immunity. Gene spr1767 encodes a protein with amino acid sequence similarity to a classic bifunctional LanM-like modification enzyme. The spr1768 gene encodes a putative flavin adenine dinucleotide (FAD)-dependent flavoprotein that could catalyze the oxidative decarboxylation of C-terminal residues. The functions of SPR1764, SPR1769, and SPR1774 are unknown. The SPR1770 protein is a predicted ABC transporter containing putative N-terminal double-glycine peptidase activity (peptidase of C39 family) and is most likely responsible for transport of modified bacteriocins and for prepeptide processing. The SPR1771 protein shares 48% identity with NisP, the nisin leader peptidase. The last two genes of the regulon, spr1772 and spr1773, encode a putative immunity protein and a putative ABC transporter, respectively. This analysis clearly shows that PneA1 and PneA2 very likely belong to the class II two-component lantibiotics.

Additionally, the gene cluster encoding the pneumococci contains the gene encoding a putative flavoprotein (the spr1768 gene product). The gene encoding this type of enzyme is also found in the gene clusters of epidermin and mersacidin. The modifications made by the flavoproteins are required for full activity of these peptides (24, 29). Whereas NisT displays rather broad transport specificity of peptides fused to the leader peptide of nisin, NisP has been shown to process only fully modified prenisin (19). Figure 2 shows the amino acid sequence alignment of pneumococci A1 and A2 with the nisin leader peptide. There is a low similarity between the peptides both in the part of the Pne leader peptides end with ProArg. Strikingly, prenisin, prePneA1, and prePneA2 cleave), one behind a shared GlyAla sequence, and one behind one of the candidate cleavage sites or in or under any other growth media or conditions we have tried (data not shown). In silico analysis analysis showed that the nine adjacent genes (Fig. 1) are likely involved in pneumococcin A1 and A2 modification, transport, processing, and immunity. Gene spr1767 encodes a protein with amino acid sequence similarity to a classic bifunctional LanM-like modification enzyme. The spr1768 gene encodes a putative flavin adenine dinucleotide (FAD)-dependent flavoprotein that could catalyze the oxidative decarboxylation of C-terminal residues. The functions of SPR1764, SPR1769, and SPR1774 are unknown. The SPR1770 protein is a predicted ABC transporter containing putative N-terminal double-glycine peptidase activity (peptidase of C39 family) and is most likely responsible for transport of modified bacteriocins and for prepeptide processing. The SPR1771 protein shares 48% identity with NisP, the nisin leader peptidase. The last two genes of the regulon, spr1772 and spr1773, encode a putative immunity protein and a putative ABC transporter, respectively. This analysis clearly shows that PneA1 and PneA2 very likely belong to the class II two-component lantibiotics.

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Production, secretion, and purification of the chimeric peptides. To investigate the production and secretion of chimeric peptides, pNZE3-spr1765 and pNZE3-spr1766 were introduced into L. lactis NZ9000 containing pIL3BTC. Cultures of L. lactis NZ9000 plasmid-containing derivatives induced with nisin were grown for 24 h in minimal medium. Subsequently, the supernatants were collected, and the prepeptides were purified on a Hitrap SP cation exchange column. The same procedure was applied for prenisin, a positive control. All prepeptides were produced and secreted, as visualized by Tris-Tricine electrophoresis (Fig. 3). Prepeptide concentrations were determined with HPLC-nisin induction, no secreted peptides were observed (data not shown). 

Streptococcal chimeric peptides are modified by nisin synthetase enzymes. To test whether the purified chimeras prePneA1 and prePneA2 were modified, they were analyzed by MALDI-TOF spectrometry. Table 2 presents a summary of the obtained masses. Interestingly, analysis of prePneA1 and prePneA2 showed that both prepeptides were modified multifold. PrePneA1 showed four- and threefold dehydrations and prePneA2 four-, three-, and twofold. Chimeric prepeptides were processed by trypsin or leucine aminopeptidase and further characterized. Since the leader peptide keeps the prepeptide inactive, its removal allows the assessment of the antimicrobial activity of the mature peptides. Trypsin cleaves a peptide bond behind lysine or arginine, with arginine being preferred over lysine, allowing arginine-specific cleavage under controlled conditions (Table 2).

The prePneA1 was processed by trypsin, and only the nisin leader sequence was clipped off, leaving the mature peptide with multiple dehydrations (Table 2). However, prolonged digestion at a higher concentration of trypsin resulted in two additional mass peaks corresponding to two fragments (Table 2). The N-terminal part of PneA1 (Table 2) was cleaved off and showed no dehydrations, likely due to protection against dehydration of Ser/Thr by their directly flanking residues (39). The identity of this N-terminal fragment was confirmed by sequence data obtained by postsource decay (data not shown). The other lysine residue in the mature peptide (SSK) appears protected against proteolysis, likely due to posttranslational modifications in its vicinity (26). The five dehydrations that were observed with the mature PneA1 are therefore located in the C-terminal part of the peptide, in which six serines are present. Since one of those serines is located next to the lysine that is cleaved by trypsin, it is most likely that the observed dehydrations are all within the last 25 amino acids of PneA1, except for the first serine at SSK.

PneA2 contains a number of residues that are substrates for trypsin, and some of them are not protected by modified residues (Fig. 2). Therefore, we initially obtained a smaller fragment (Table 2, PneA2 fragment 2), which lacked the C-terminal extension but contained multiple dehydrated residues. Additionally, in HPLC-purified chimeric prePneA2, there was also a clear fraction containing a peptide fragment consisting of part of the nisin leader sequence and the N-terminal part of the PneA2 peptide (Table 2, prePneA2 fragment 1). Interestingly, this peptide fragment contained up to four dehydrations, which clearly shows that four out of five dehydrated residues are located in the first 11 amino acids of the mature peptide.

Subsequently, removal of the N-terminal leader peptide from chimeric peptides by NisP overexpressed in L. lactis was also investigated. However, neither the release of the leader peptide nor the antimicrobial activity of PneA1 or PneA2 was detected (data not shown). Intact PneA2 was obtained using leucine aminopeptidase and appeared to be three- to sixfold dehydrated (Table 2). Importantly, the activity of the leucine aminopeptidase apparently had stopped after the last Arg. This indicates the presence of a thioether bridge starting at either Ser1 or Thr2. Studies by Rink et al. indicated that flanking hydrophilic residues on both sides of a Ser or Thr would not favor dehydration. Furthermore, serines are less readily dehydrated than threonines (39). Therefore, a large extent of the dehydration of Ser1 flanked by Arg and Thr is not likely. Taken together, these findings indicate that Thr2 is fully dehydrated and is thioether cross-linked to Cys5.

To investigate whether thioether rings were formed, CDAP, which reacts only with free cysteines, was used. The control NisB2 peptide that contains one free cysteine was used as a positive control (not shown) for CDAP modification. CDAP modification converts a free thiol group of cysteine into an isothiocyanate, yielding a mass increase of 25 Da. The fivefold dehydrated PneA1 showed hardly any CDAP modification.
indicating that there were no free cysteine residues and thus the formation of two thioether rings. The threefold dehydrated PneA1 peptide showed single and double CDAP modifications, indicating the formation of either one or no thioether rings (Fig. 4). Similar studies of CDAP modification of PrePneA2 indicated the presence of two thioether rings in the extensively dehydrated peptides (not shown). These data demonstrate that the putative lantibiotics, which are entirely unrelated to nisin, can be successfully produced, modified, and secreted by the nisin synthetase machinery.

The produced and modified peptides have significant antimicrobial activity. To investigate the antimicrobial activity of the modified peptides, the chimeric prepeptides were incubated with trypsin or leucine aminopeptidase to remove the N-terminal leader sequence. Various dilutions of trypsin-treated peptides, namely, prenisin (positive control) and pneumococcins A1 and A2, were tested for antimicrobial activity in the MIC assay (Table 3). Of all microorganisms tested, i.e., M. flavus, S. pneumoniae, E. faecalis, S. aureus, and S. mitis, only M. flavus was susceptible to the tested peptides, i.e., PneA1 and PneA2 (Table 3). In control experiments, no significant inhibition was found with either buffer or bovine serum albumin (BSA) treated with trypsin or with empty samples, i.e., fractions from prepeptide purifications that did not contain peptides (Table 3). Additionally, undigested chimeric prepeptides did not show significant antimicrobial activity against the indicator strain (Table 3). Unmodified PneA1 (MIC of >50 μM) and PneA2 (MIC of >1.5 mM) propeptides obtained by chemical synthesis were at least 30-fold and 170-fold less active,
TABLE 3. Susceptibility of *M. flavus* to NisBC-modified PneA1 and PneA2 peptides

<table>
<thead>
<tr>
<th>Sample</th>
<th>MIC (μM) for indicated digestion type</th>
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<tr>
<td></td>
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<tr>
<td>Prenisin</td>
<td>20</td>
</tr>
<tr>
<td>Nisin leader peptide PneA1</td>
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</tr>
<tr>
<td>Nisin leader peptide PneA2</td>
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<tr>
<td>BSA</td>
<td>NI</td>
</tr>
<tr>
<td>Buffer</td>
<td>NI</td>
</tr>
<tr>
<td>Empty sample</td>
<td>NI</td>
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*ni*, no inhibition. MICs are calculated using the molecular weights of the mature peptides.

respectively, than the HPLC-purified active fraction of the corresponding NisBC-modified peptides, without leader peptide. This proves that NisBC-induced modifications are required for lantibiotic activity. PneA1 inhibited the growth of *M. flavus* at a peptide concentration of 0.6 μM. PneA2, from which the leader sequence was removed by either trypsin or leucine aminopeptidase, inhibited growth at approximately 10 or 8.5 μM, respectively (Table 3). The combination of both modified chimeric peptides, PneA1 and PneA2, did not act synergistically (Table 3). Thus, the data demonstrate that it is possible to utilize the nisin synthetase machinery for the production of antimicrobially active peptides unrelated to nisin.

**DISCUSSION**

To the best of our knowledge, we present here for the first time the successful expression, modification, secretion, and biological activity of novel class II lantibiotics by the nisin synthetases, which normally produce nisin, a class I lantibiotic. To present a significant challenge as substrate peptides, pneumococci A1 and A2 from *S. pneumoniae* R6, which presumably belong to the class II two-component lantibiotics, were chosen as substrates for the nisin enzymes. The class II two-component lantibiotics require a LanM-type enzyme that performs both dehydration and cyclization, whereas class I lantibiotics require LanB dehydratases and LanC cyclases. It has already been shown that LanBC-type enzymes can modify peptides other than nisin which are fused to the nisin leader sequence. Kluskens et al. and Kuipers et al. demonstrated that both medically relevant nonlantibiotic peptides and a truncated lantibiotic, lactin 3147, that were fused with the nisin leader sequence, modified by NisB and NisC, were exported via NisT and contained dehydrated amino acids and lanthionine rings (16, 20). The same was proven by Rink et al. for various hexapeptides (38, 39). Thus, based on the discovery that the nisin synthetase machinery can accept various peptides as templates for modification, the propeptide part, which is the predicted maturing part of either the PneA1 or the PneA2 peptide, was fused to the nisin leader sequence and introduced into *L. lactis* that overexpresses NisBTC. The produced peptides were dehydrated multifold and contained thioether rings. Some dehydrated residues and one thioether ring could be localized by studying peptide fragments and by applying leucine aminopeptidase.

To be biologically active, lantibiotics require prepeptide processing, i.e., removal of the leader sequence. The prepeptide sequences and the homology of the peptidase with NisP indicate that the site behind PR might be the processing site from which the propeptide starts. To liberate the mature peptides, we used trypsin or leucine aminopeptidase. Exported, purified, and processed peptides were tested for antimicrobial activity, and *M. flavus* was found to be highly susceptible to both the PneA1 and PneA2 peptides. Despite the fact that PneA1 and PneA2 are predicted two-component lantibiotics, we did not observe any significant synergistic effect when these peptides were combined (data not shown). The PneA1 and PneA2 peptides are a mixture of extensively modified and hardly modified peptides. These mixtures probably contain active, less active, and inactive peptides. Thus, the determined antimicrobial activity is the mean of the activities of all these peptides, indicating that the specific activity for a single active peptide might be higher. With respect to this, a preliminary experiment was performed with processed and NisBC-modified PneA1 and PneA2 peptides which were dehydrated four- and fivefold, separated, and purified from the total mixture. However, the MICs of both peptides were not significantly different from the MICs of the unpurified peptides (data not shown). A challenge for future work might be to sort out the active peptide fraction from the inactive fraction in order to get a better picture of which modifications yield antimicrobially active peptides.

The putative cluster of pneumococci consists of 11 ORFs. In the cluster, two genes might be required for proper modification of PneA1 and PneA2. These genes encode a single putative LanM-type modification enzyme and a putative LanD-type flavoprotein. Flavoproteins catalyze the oxidative decarboxylation of a C-terminal cysteine residue involved in ring formation. A FAD-dependent flavoprotein catalyzes this reaction for mersacidin, a lantibiotic produced by *Bacillus* sp. (29). Another flavoprotein, which is flavin mononucleotide (FMN) dependent, catalyzes the same reaction for epidermin, a lantibiotic of *S. epidermidis*, and this enzyme is essential for formation of a biologically active peptide (24). It is not known whether the putative LanD-type flavoprotein of PneA1 and PneA2 performs a similar function in this cluster. Because the original cluster of pneumococci contains LanM- and LanD-type modification enzymes, peptides modified by NisB and NisC might not be fully active by the lack of the oxidative decarboxylation. Furthermore, we do not know whether the native dehydration and ring pattern is exactly similar to the one installed heterologously by NisB and NisC. These factors might explain the presumably suboptimal antimicrobial activities and lack of synergism within this putative two-component lantibiotic system. However, both peptides, PneA1 and PneA2, still showed significant antimicrobial activity.

The production of nonlantibiotic or lantibiotic chimeras with a heterologous system has been reported using either closely related or nonlantibiotic peptides. For example, production of chimeric nonlantibiotic bacteriocin pediocin PA-1, which is fused to the leader of lactococcin A and/or to enterocin P, or enterocin A, which is fused to the leader of enterocin P, resulted in the secretion of active peptides (13, 30, 31). These
cases of successful production of biologically active bacteriocins concerns nonlantibiotic bacteriocins, which in contrast to lantibiotics do not require posttranslational modifications for antimicrobial activity. Production of class I lantibiotic chimeras, such as nisin/subtilin or subtilin/nisin, with either subtilin or nisin expression machineries was performed successfully (1, 23). Of the amino acid residues of the leaders and mature peptides of subtilin and nisin, 57% were identical (23). Studies using lactacin 481 synthetase demonstrated its ability to prepare other lantibiotics in the class II lactacin 481 family, including nukacin ISK-1, mutacin II, and ruminococcin A (37).

In contrast, we show here for the first time that it is possible to use the nisin synthetase system to produce, modify, and secrete lantibiotics, from a very different source and class, that exhibit considerable antimicrobial activity.

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