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The major amino acid transporter superfamily has a similar core structure as Na⁺-galactose and Na⁺-leucine transporters

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Abstract

The sodium solute symporters (SSS) and neurotransmitter sodium symporters (NSS) are two families of secondary transporters that are not related in amino acid sequence. Nonetheless, recent crystal structures showed that the Na⁺/galactose (SSS) and Na⁺/leucine (NSS) transporters have similar core structures. The structural relatedness highlights the need for classification methods for membrane protein structures based on other criteria than amino acid similarity. Here, we demonstrate that a method based on hydropathy profile alignments convincingly identifies structural similarity between the NSS and SSS families. Most importantly, the method shows that one of the largest transporter families for which a crystal structure is elusive (the amino acid/polyamine/organocation or APC superfamily), also shares the similar core structure observed for the Na⁺/galactose and Na⁺/leucine transporters. The APC superfamily contains the major amino acid transporter families that are found throughout life. Insight into their structure will significantly facilitate the studies of this important group of transporters.

Keywords: Transporter classification, transport protein structure, MemGen, hydropathy profile alignment, structural class

Introduction

Very recently, the crystal structure of a Na⁺/galactose symporter vSGLT was reported at a resolution of ~ 3 Å [1]. The transporter found in Vibrio parahaemolyticus is the first member of the sodium solute symporter (SSS) family for which a crystal structure has been solved. The SSS family is a ubiquitous family of Na⁺-coupled transporters including the well studied mammalian transporters for glucose (SGLT) and iodide (NIS) and the bacterial proline transporters (PutP) [2]. The Na⁺/galactose symporter has 14 transmembrane segments (TMSs) with a core of two structurally similar domains of five TMSs each. The two domains have opposite orientations in the membrane and bind the galactose molecule at their interface. The domains are further characterized by a break in the first transmembrane α-helix (Figure 1). Surprisingly, the structural arrangement of the core of the galactose transporter had been seen before in LeuT [3], a leucine transporter of the neurotransmitter sodium symporter (NSS) family, a family unrelated to the SSS family in amino acid sequence. The observation stresses the importance of methods other than amino acid sequence analysis to structurally classify membrane proteins.

The MemGen method uses hydropathy profile alignments to predict structural similarity of membrane proteins not related in sequence [4]. Although the method predicted the structural similarity between the transporters of the SSS and NSS families [5], the limited data set available at the time (1998) made the method prone to the identification of false positives. Here, we show that using the vast number of sequences in current databases, the MemGen method convincingly demonstrates structural similarity of the SSS and NSS families. In fact, TMSs II-XI of the SSS transporters corresponding to TMSs I-X of the NSS transporters are identified correctly as the core of the proteins. More importantly, based on the larger dataset, the method also decidedly predicts the same structural core in the transporters of the amino acid-polyamine-organocation (APC) superfamily [6], one of the largest remaining families of secondary transporters for which no crystal structure is available. The APC superfamily is the main family of amino acid transporters found in all domains of life.
Molecular Membrane Biology online.

Methods

Computational methods

Members of the SSS, NSS and AAT families were collected by BLAST searches [7] of a database of proteins coded on the genomes of 649 available microbes. vSGLT of *Vibrio heamolyticus* and the Na⁺-leucine transporter LeuT of *Aquifex aeolicus* (NSS) and the aromatic amino acid transporter AroP of *Escherichia coli* were used as queries. AroP is a member of the AAT family, one of the families in the APC superfamily. Sequences were selected from each set that shared between ~20 and 60% sequence identity by pairwise alignment. Family hydropathy profiles were calculated following multiple sequence alignment by CLUSTALW2.0 [8] as described [4]. The structure divergence scores of the family profiles (SDS, see [4]) were 0.123, 0.125, and 0.125 for the SSS, NSS and AAT families, respectively. The algorithm to find the optimal alignment of the family profiles and calculation of the S-factor to discriminate between similar and dissimilar structures have been described as well [4].

Results and discussion

The hydropathy profile of the amino acid sequence of a membrane protein shows a characteristic peak pattern due to the clustering of hydrophobic residues in the parts of the polypeptide chain that are embedded in the membrane. MemGen uses the hydropathy profile as a ‘reporter’ or ‘fingerprint’ of the global folding of the protein to compare the structures of different proteins, i.e., for structural classification. First, an average ‘family hydropathy profile’ is calculated based on the multiple sequence alignment of a set of homologous proteins (e.g. from the SSS family), which show unambiguous similarity at the amino acid level. Then, in a way similar to comparing amino acid sequences, MemGen compares averaged hydropathy profiles of different families of membrane proteins (e.g. SSS and NSS) by finding the optimal alignment of the profiles, without using the amino acid sequences, and allowing for insertions and deletions. The divergence of the hydropathy profiles within a family (e.g. the variation in the single protein profiles in the SSS family) is then compared to the divergence between the averaged hydropathy profiles of different families (e.g. between NSS and SSS) to provide a numerical criterion for structural similarity. Similarity is expressed in the similarity score S that takes values of 1 or below for similar structures. When the method was introduced, and only a very limited set of sequences was available, 13 families of secondary transporters were classified into 4 structural classes, termed ST[1], ST[2], ST[3], and ST[4] [4]. Later, by screening the available sequence databases, many more families were assigned to these structural classes [5,9]. All the families in one particular class are predicted to have the same global fold. The SSS, NSS and APC families are all found in structural class ST[2].

The limited data set used before made the method prone to errors because of poorly defined average family hydropathy profiles. We now reevaluate the predicted structural similarity of the SSS and NSS families, for which crystal structures of representatives are now available, and APC families, for which no crystal structure has been determined. The sequences of members of the SSS, NSS, and APC families were extracted from a database of proteins coded on 649 bacterial and archaeal genomes. The families contained 36, 113 and 133 sequences, respectively, with pair wise sequence identities in each family ranging between 20 and 60%. Optimal alignment of the SSS and NSS profiles resulted in an S (similarity) score of 0.879, which is strongly indicative of similar structures (Figure 2, top). Similarly, optimal alignments of the SSS and APC profiles (Figure 2, bottom) and of the NSS and APC profiles (not shown) result in similarity scores of 0.900 and 0.944, respectively, showing that the APC superfamily belongs to the same structural class. The section marked as ‘core’ is present in all three profiles and can be found in all other families in structural class ST[2], while regions outside the core may be present or not (not shown). Clearly, the SSS family contains an extra TMS at the N-terminus that is not present in the NSS and APC families and the
number TMSs at the C-terminal side of the core is variable even within the families.

Structural class ST[1] in the MemGen classification system largely correspond to the Major Facilitator Superfamily (MFS) from sequence homology [4,5]. ST[2] contains the APC superfamily and the SSS and NSS families discussed here plus a number of smaller families of transporters, mainly for amino acids. ST[3] has been analyzed most extensively and contains a total of 36 families among which the families of the Ion Transporter superfamily (IT) and 2-hydroxycarboxylate transporter family (2HCT) and Na⁺-coupled glutamate transporters (ESS) [9,10]. Finally, ST[4] is unique in that it contains a single family of glutamate and neutral amino acid transporters (DAACS) [10]. Until now, available crystal structures of secondary transporter proteins supported the MemGen classification in the sense that proteins from different classes (LacY and GlpT in ST[1] [11,12], LeuT in ST[2] [3] and GlpPh in ST[4] [13] clearly represented different structures (see also reference [14]). The high resolution structures of vSGLT and LeuT now also provide support for the MemGen classification of families within a structural class: two proteins in the same MemGen class, but unrelated in sequence, share the same global fold. Before, this had been demonstrated only at the low resolution level of the membrane topology for families in class ST[3] [15].

In the MemGen classification, the APC superfamily is in the same structural class (ST[2]) as the SSS and NSS families, meaning that they share the core structure observed in the vSGLT and LeuT transporters. This should significantly facilitate the characterization of this large and important group of transporters.

Figure 2. Hydropathy profile alignment of the SSS and NSS (top) and the SSS and AAT (bottom) families. The family hydropathy profile of the SSS family is indicated in red, those of the NSS (top) and AAT (bottom) families in blue. The AAT family is one of the families in the APC superfamily. Red and blue bars at the top indicate positions where gaps were introduced in the profiles by the alignment algorithm. Dashed boxes show the two domains in the core structure. The position of the 2 times five TMS is indicated. This Figure is reproduced in colour in Molecular Membrane Biology online.
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