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Genomic and expression analyses identify a disease modifying variant for fibrostenotic Crohn’s disease

Running title: Risk variant for fibrostenotic Crohn’s disease

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Abstract

Background and Aims

Crohn’s Disease (CD) is a chronic inflammatory disease with unpredictable behaviour. More than half of CD patients eventually develop complications such as stenosis, for which they then require endoscopic dilatation or surgery, as no anti-fibrotic drugs are currently available. We aim to identify disease-modifying genes associated with fibrostenotic CD.

Methods

We performed a within-case analysis comparing “extreme phenotypes” using the Immunochip and replication of the top SNPs with Agena Bioscience in two independent case-control cohorts totalling 322 cases with fibrostenotis (recurrent after surgery) and 619 cases with purely inflammatory CD.

Results

Combined meta-analysis resulted in a genome-wide significant signal for SNP rs11861007, \(P = 6.09 \times 10^{-11}\), located on chromosome 16, in IncRNA RP11-679B19.1, a IncRNA of unknown function, and close to exon 9 of the \(WWOX\) gene, which codes for WW domain-containing oxidoreductase. We analysed mRNA expression of \(TGF-\beta\) and downstream genes in ileocecal resection material from ten patients with and without the \(WWOX\) risk allele.

Patients carrying the risk allele (A) showed enhanced colonic expression of \(TGF-\beta\) compared to patients homozygous for the wild-type (G) allele \(P= 0.0079\).
Conclusion

We have identified a variant in WWOX and in lncRNA RP11-679B19.1, as a disease-modifying genetic variant associated with recurrent fibrostenotic CD and replicated this association in an independent cohort. WWOX can potentially play a crucial role in fibrostenosis in CD, being positioned at the crossroads of inflammation and fibrosis.

Keywords: Crohn’s Disease, Fibrosis, Genetics.
Introduction

Crohn’s Disease (CD) is a chronic Inflammatory Bowel Disease (IBD) with unpredictable disease behaviour; more than half of CD patients eventually develop complications such as fistulae and stenosis\(^1\). Patients with fibrosis-induced stenosis require endoscopic dilatation or surgery, as no anti-fibrotic drugs are currently available. The estimated lifetime risk for surgery in CD patients is 70\%, with fibrostenosis as the most common indication for surgery\(^2\). Within one and a half year after surgery, the endoscopic recurrence rate of fibrostenosis is 40-60\%\(^3\).

Fibrostenosis of the intestine is a result of chronic inflammation: the permanent scarring and the consequent luminal narrowing is induced by continuous events of injury and healing. Finally, the intestine is no longer able to restore the normal histological configuration and excessive deposition of extracellular matrix prevails. This leads to thickening of the intestinal wall, which in turn leads to luminal stricturing, finally causing stenosis and obstruction.

Genetic studies of IBD have identified over 240 risk loci associated to disease development\(^4\)\(^5\). A large genotype-phenotype association study was performed, including 34,819 IBD patients, which identified three loci (NOD2, MHC, and MST1 3p21) associated with disease location\(^6\). However, only a few risk variants are known that influence disease behaviour; such as SMAD3 with recurrent surgery in CD, MAGI1 with complicated structuring CD, and FOXO3, XACT and IGFBP1\(^-3\) with severe disease course in CD\(^7\)\(^-10\). Variants associated with disease onset seem not to be associated with disease behaviour, which suggests that the biological pathways that underlie disease behaviour are distinct from those that underlie disease onset\(^10\). In this study, we aim to identify disease-modifying genes associated with the fibrostenotic phenotype in CD. This will help us gain insight into the biological mechanisms underlying intestinal fibrosis development, which might open new avenues for the treatment of CD.
We performed a within-cases analysis comparing “extreme phenotypes”, an approach that provides power to detect associations within disease cohorts that has previously been successfully adopted for other traits, including obesity\textsuperscript{11,12}. We used an immune focused fine mapping chip (Immunochip) with 166,251 Single Nucleotide Polymorphisms (SNPs), in two independent case-control cohorts totalling 322 individuals with multiple (≥2) resections due to confirmed ileal stenosis, which we used as cases, and which we compared to patients with 619 purely inflammatory CD (non-penetrating/non-fibrostenotic with disease duration >5 years). Following this approach, we identified and replicated a variant within lncRNA RP11-679B19.1 and in WWOX, as disease-modifier, associated with recurrent fibrostenotic CD. We then studied the potential functional consequences of this variant by studying resection material of CD patients carrying the risk allele.

**Methods**

**Genetic analysis (cohort description, quality control and statistical analysis)**

*Subjects* The discovery cohort consisted of 521 Dutch CD patients, selected out of ~ 1,500 Dutch CD patients. Selection was based on reviewing the medical records of the total cohort of 1,500 patients. We selected patients on extreme ends of the fibrostenotic phenotype: a case group and a control group. The case group consisted of 242 patients with ‘recurrent fibrostenotic’ CD, which was defined as the need for multiple (≥2) resections due to confirmed ileal stenosis. The control group consisted of 279 patients with purely inflammatory disease behaviour CD (Montreal classification B1\textsuperscript{13}) defined as non-penetrating, non-fibrostenotic CD with a disease duration of >5 years, without the need for any kind of surgery. All patients of the discovery cohort were also included in the IIBDGC core-phenotype paper\textsuperscript{6}.

For the replication phase, patients were selected from the Parelsnoer Institute database (http://www.parelsnoer.org/). This database consists of 3,394 IBD patients (CD: n=2,118) collected...
from the IBD Centres in all eight University Medical Centres in the Netherlands. Patients already included in the discovery phase were excluded. Eighty CD patients (n=80) with recurrent fibrostenotic disease and 340 patients with purely inflammatory disease were selected with similar criteria. While the phenotypes of the discovery cohort could be checked to the patients’ medical charts, regulations prevented us from doing the same with the replication cohort. Hence, the replication cohort might have some non-cases and/or non-controls admixed. All patients were of Central European descent.

**Discovery phase**

*Immunochip Genotyping* In the discovery phase of the study we used genotype data of ~1,500 CD patients described by Jostins *et al.* In short, DNA was extracted from whole blood. Illumina Immunochip was used to genotype the DNA samples. The Immunochip is a custom Illumina Infinium immune focused fine-mapping chip comprising 196,524 Single Nucleotide Polymorphisms (SNPs) selected primarily based on Genome Wide Association Study (GWAS) analysis of 12 immune-mediated diseases.

*Quality Control* Quality control (QC) was performed using PLINKv1.07 software. QC removed SNPs with a minor allele frequency (MAF) less than 0.001, as well as SNPs with more than 10% missing genotypes. 547 patients were selected (248 fibrostenotic CD patients and 299 CD patients with purely inflammatory disease behaviour). After removing duplicate individuals (n=6) and individuals with more than 15% missing genotype data (n=20), 521 individuals remained (no differential missingness, \( \chi^2 P = 0.67 \)). After QC, the dataset consisted of 521 individuals and 166,251 SNPs with a genotyping call rate of 99%.

**Statistical analysis and prioritization of SNPs for replication.** After allele association analysis (\( \chi^2 \) test) and cluster plot inspection, 34 SNPs with a p-value <1.0×10\(^{-3}\) remained. In order to adequately test
each locus for association in the replication phase we selected 2 SNPs per locus where possible. In addition, we decided to test the SNP that previously showed the strongest association with severe CD disease course (rs12212067, FOXO3), although this SNP was not associated with fibrostenotic disease in our discovery cohort\(^{10,16}\). 43 SNPs passed the design of 2 plexes with the Assay Design suite of Agena Bioscience (https://seqpws1.agenacx.com/AssayDesignerSuite.html).

**Replication phase**

*Genotyping* 43 SNPs were genotyped using the Agena Bioscience Massarray in the replication phase (http://agenabio.com). During QC we excluded SNPs with a Hardy–Weinberg equilibrium p-value <0.0001 in controls and an overall call rate <90% and individuals with <85% of SNPs confidently genotyped. After QC, the replication cohort consisted of 80 CD cases with recurrent fibrostenotic disease behaviour, 340 CD patients with purely inflammatory disease and 39 SNPs with a genotype call rate of 99%. Since rs11861007 failed the Agena Bioscience design and it was the only Immunochip SNP in the WWOX region, genotyping of SNP rs11861007 was performed using Taqman technology (Applied Biosystems).

**Statistical analysis** Allelic association analysis ($\chi^2$ test) and meta-analysis of the complete cohort was performed with PLINKv1.07 software\(^{15}\). To test if disease localization could be a cofounding factor, we performed, additionally, an allelic association analysis for rs11861007 ($\chi^2$ test) in ileal disease patients (n=86) versus colonic disease patients (n=36). Similarly, allelic association analysis for rs11861007 ($\chi^2$ test) in Montreal Class B2 phenotype patients (n=165) versus Montreal Class B1 and B3 phenotype patients (776) was performed to distinguish if rs11861007 was associated with the Montreal B2 phenotype (and not re-current fibrostenotic disease.) Logistic regression analyses with disease as a covariate was performed with R statistics.
Given that the prevalence of recurrent stenotic disease in CD was 0.04 in our replication cohort, that the allele frequency of our rs11861007 risk allele A is 0.06 in CEU, and that we expected the risk variant to have an OR of 4.3, we expected to have >80% power to discover this association in our replication cohort. Ultimately, we only had 60% power to detect the association that we did detect for rs11861007 in our replication cohort at OR 2.1 (http://zzz.bwh.harvard.edu/gpc/cc2.html).

**SNP annotation**

LocusZoom was used to construct regional association plots\(^\text{17}\) (Supplementary Figure 1). Exploration of the linkage disequilibrium (\(r^2 > 0.8\)) was performed with Haploview\(^\text{18}\). We assessed whether associated SNPs had known functional consequences or regulatory features. The Encyclopedia of DNA Elements (ENCODE)\(^\text{19}\) was searched using the UCSC Genome Browser\(^\text{20}\). Specifically, SNPs located in the following regulatory features were searched: DNaseI-hypersensitivity sites, transcription factor binding sites, histone modification and DNA-polymerase sites. We tested whether associated variants (or SNPs in high LD (\(r^2 > 0.8\))) showed an effect on gene expression levels of genes. The expression Quantitative Trait Loci (eQTL)-analysis was performed with the eQTL browser (http://genenetwork.nl/bloodeqtlbrowser/), based on non-transformed peripheral blood in 5,311 individuals\(^\text{21}\). Additional enhancer analyses were performed with the Fantom5 enhancer atlas\(^\text{22}\).

**In Silico analysis (rs11861007)**

The strongest association for fibrostenotic disease was found in the RP11-679B19.1 lncRNA, and in the WWOX gene. The function of the RP11-679B19.1 lncRNA is unknown. To predict in which biological or cellular process and molecular function the WWOX gene is involved, we used an in house developed RNA network tool (http://www.genenetwork.nl)\(^\text{23}\). The RNA network uses a method, based on principal component analysis, to build transcriptional profiles for biological pathways, which can
be used to predict gene functions. A more detailed description of the method of the RNA network can be found in the paper of Fehrmann et al.\textsuperscript{23}.

**Expression analysis**

**Resection material of Crohn’s Disease patients**

*Subjects* The resection material was collected during surgical resection procedures (samples of the small bowel and colon) from patients for whom we had extensive phenotype data (University Medical Center Groningen). Twenty-nine patients with fibrostenotic CD behaviour were included. The resection material of CD patients was preserved (storage at -80°C) in three parts: the ileum (proximal of the stenosis), the stenosis itself (medial part) and the colon (distal of the stenosis). As stenotic tissue is a final stage in the process of fibrosis formation, we considered the stenotic tissue, medial part of resection, as not representative. Therefore, we included only samples of the ileum and colon, respectively proximal and distal from the stenosis. Pathology records of the resection material were checked and report no severe inflammation or fibrosis at the end of the resection sides, proximal and distal, of the stenosis.

*Genotyping and patient selection* After DNA isolation, genotyping of the WWOX SNP (rs11861007) in 71 patients was performed using Taqman technology with a call rate of 96\% (Life Technologies). We selected the five CD patients that carried the WWOX-genotype (risk allele, A), considering these cases (we found no patients homozygous for the risk allele in our cohort). We selected five matched CD patients with the GG genotype (wild-type) as controls (matched based on age, disease duration, inflammation and stenosis).

Quantitative Polymerase Chain Reaction (qPCR) was performed of WWOX, TGF-\(\beta\), iNOS, IL1-\(B\), TNF-\(\alpha\), FOXP3, \(\alpha\)-SMA, Collagen Type 1 and downstream genes PAI-1 (SERPINE) and CTGF of the
ileocolonic tissue of five CD patients carrying the WWOX risk allele and five CD patients homozygous for the WWOX wild-type allele (Online Methods).

Results

In this study we included two independent case-control cohorts totalling 322 recurrent fibrostenotic and 619 purely inflammatory CD cases, 60% of the cohort is female with a mean age at diagnosis of 25 years. The cohorts were selected based on “extreme phenotypes”, resulting in a statistically significant difference in disease location and behaviour between patients (Table 1). After analysis of 166,251 SNPs in 242 fibrostenotic and 279 purely inflammatory CD cases and replication of 34 selected SNPs in an independent cohort of 80 fibrostenotic and 340 purely inflammatory CD cases, the combined meta-analysis resulted in a genome-wide significant signal for SNP rs11861007 ($P = 6.09 \times 10^{11}$, odds ratio $= 3.2$, heterogeneity ($I^2 < 75\%$) (Table 2, Supplementary Table 1). The minor heterogeneity for SNP rs11861007 between the discovery and the replication cohort, might be caused by a slight admixture of non-cases and/or non-controls in the replication cohort, as described in the Methods section. To assess whether the association between rs11861007 and fibrostenotic disease was not mainly determined by ileal disease localization we tested this marker for association to ileal disease localization. We found no association between rs11861007 and ileal disease localization ($P = 0.27$). Additional logistic regression analyses for the association between SNP rs11861007 and re-fibrosis, with disease location as a covariate, showed a still remaining significant association ($p=0.004$), although it reduces the significance level. We also tested for association between rs11861007 and the Montreal Class B2 phenotype (any stricuring disease) in our cohort, but did not find an association ($P= 0.25$).
**SNP Annotation and in silico analyses**

rs11861007 is located on chromosome 16, in an intron close to exon 9 of the *WWOX* gene, which codes for WW domain-containing oxidoreductase. Neither rs11861007, nor SNPs in high linkage disequilibrium (r^2 > 0.8), have known functional or regulatory features (eQTL or regulatory elements assessed in the ENCODE Encyclopedia of DNA Elements or FANTOM5 enhancers). *WWOX* is the only coding gene in the locus (defined as 250-kb on either side of our top hit) making it the most likely positional candidate gene. The SNP is also located in the lncRNA RP11-679B19.1, in which it might affect folding, but for this lncRNA there is also no eQTL effect, and no function is known for RP11-679B19.1.

Co-transcriptional pathway analysis using an in-house-developed RNA network tool predicts *WWOX* involvement with cellular components in the *extracellular matrix compartment* (P = 1.41 × 10^-3) and an association with *collagen binding* (P = 9.78 × 10^-5). See Supplementary Table 2 for involvement in other pathways.

**Expression analyses**

Normal ileal and colonic tissue residing proximal and distal from the stenotic part was sampled from 5 CD patients heterozygous for the *WWOX* risk allele (A) and 5 CD patients homozygous for the *WWOX* wild-type allele (G). *WWOX* mRNA levels were similar both in the ileal and colonic tissue of patients with or without the risk allele (Supplementary Figure 2). In contrast, TGF-β expression was significantly higher in the colonic tissue of risk-allele-carrying patients compared to patients homozygous for the wild-type allele (Mann-Whitney U-test, P = 0.0079) (Figure 1A). Similarly, downstream targets involved in fibrosis, like Connective Tissue Growth Factor (*CTGF*) and matricellular *PAI-1 (SERPINE1)*, showed a trend towards increased expression in the risk-allele-carrying CD patients compared to the patients homozygous for the wild-type allele (Figure 1B, Supplementary Figure 2 for all qPCR results).
Discussion

In this study, we have identified a variant in the WWOX gene as a disease-modifier associated with a recurrent fibrostenotic phenotype in CD, and we have replicated this association in an independent cohort. We found that patients carrying the risk allele show enhanced profibrotic signalling, through higher expression of TGF-β, in the colon. This would suggest that the genetic variant in WWOX is associated with a decrease of WWOX function. Alternatively, the effect could be caused by a configuration change of the lncRNA RP11-679B19.1, which has recently been recorded in the same locus and is, yet, of unknown function.

The genetic variants that contribute to disease behaviour can be different from the variants that contribute to disease susceptibility. This has been shown for a variant affecting FOXO3A expression, which was found to be significantly associated with disease prognosis but which was not associated with CD susceptibility\(^ {16}\). In this study, we confirm the concept that disease-modifying genes can differ from variants contributing to disease development. Our most associated SNP (rs11861007) has not been associated with disease development in previous studies\(^ {4}\). Previous genotype-phenotype studies in IBD patients have not studied the specific phenotype in the present study: recurring ileal fibrostenosis in CD\(^ {6}\). Previous studies focusing on genetic variants associated with a single fibrotic event in CD found associations with variants in SMAD3 and MAGI, however due to the diverse coverage of the genotyping platforms we could not replicate these findings\(^ {7,8}\). We could also not confirm a previously described association between NOD2 variants and ileal fibrostenotic disease, which might be due to the fact that we study recurrent fibrostenosis, not necessarily ileal, or because the NOD2 variants are relatively rare in our cohort.

The SNP (rs11861007) is located in an intron close to exon 9 of the WWOX gene. Although we could not find an eQTL effect, regulatory features or enhancer activity, the WWOX gene is the only coding gene in the locus (defined as 250-kb on either side of our top hit) making it the most likely positional candidate gene. The SNP is also located in the lncRNA RP11-679B19.1, in which it might...
affect folding, but for this lncRNA there is also no eQTL effect, and no function is known for RP11-679B19.1. WWOX is a known tumor suppressor gene and encodes a protein that contains 2 WW domains and a short-chain dehydrogenase/reductase domain (SRD). The mechanism of tumor suppression of WWOX involves apoptosis, modulation of the extracellular matrix, and modulation of cell bioenergetics. Genome-wide association studies have shown that WWOX also plays a role in the pathogenesis of pulmonary fibrosis. Targeted deletion of Wwox in epithelium in the mammary gland increased fibronectin levels and conditional deletion of Wwox in the mammary gland significantly upregulated multiple collagen genes. Moreover, molecular functions predicted by our RNA network tool show that WWOX plays a role in fibrosis formation, in agreement with the previously described reports from literature.

The main mediator between intestinal inflammation and fibrosis in IBD is TGF-β, which is overexpressed in intestinal tissue in CD patients. Upon TGF-β stimulation, WWOX acts as an inhibitor of SMAD3 transcriptional activity by sequestering it in the cytoplasm. We show an enhanced TGF-β expression in CD patients carrying the WWOX risk allele. TGF-β stimulates downstream signalling pathways resulting in expression of several profibrotic genes, including CTGF.

qPCR analysis of CTGF in this study showed a trend towards increased expression in the WWOX risk-allele-carrying CD patients compared to the patients homozygous for the WWOX wild-type allele. We conclude that WWOX risk-allele-carrying individuals have enhanced TGF-β expression with a trend towards elevated expression of profibrotic genes as a downstream effect.

There are some limitations to this study. First, the cohort size is relatively small. However, by using a within-case analysis comparing “extreme phenotypes” we increased the power of our analysis. This approach provides power to detect associations within disease cohorts and the approach has been successfully adopted for other traits. Moreover, we replicated our findings in an independent cohort. Second, the upregulation of TGF-β in the ileal part of the resection material (upstream of the stenosis) in patients carrying the WWOX risk allele was a trend and not statistically significant. However, in the colonic part of the resection material (downstream of the stenosis) in patients carrying the WWOX risk allele we do show statistically significant enhanced TGF-β expression. All available
WWOX risk-allele-carrying patients in our centre were included, since the risk allele has a low frequency this number is quite small, which means our study is relatively underpowered. Increasing the sample size may turn the trend we observe into a significant association. Finally, the WWOX-SMAD3-TGF-β pathway has been described previously\textsuperscript{31} and it has been proven that the proteins encoded by the genes interact, but the exact pathways through which they interact have not yet been elucidated, making it difficult to interpret the results from our study.

In conclusion, we have identified and replicated WWOX as a disease-modifying gene associated with the recurrent fibrostenotic phenotype in CD. Our expression analyses suggest a functional effect of the risk allele through enhanced expression of TGF-β in risk-allele carriers, which propose profibrotic expression. CD patients carrying the WWOX risk allele appear to have a profibrotic profile. To avoid fibrotic complications, it might be advisable to refrain from prescribing anti-inflammatory medication that enhances TGF-β signalling in intestinal fibroblasts in these patients.

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This manuscript, including related data, figures and tables has not been previously published and is not under consideration elsewhere.

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We thank Dianne Jansen and Tjasso Blokzijl for their expert technical help and Kate McIntyre for editorial assistance.

Author's contributions

Genetic study designs: MCV, EAMF, RKW. Expression analysis design: MCV, EAMF, GD, KNF, MA, RKW. Clinical information collection: MCV, LMS, EVL. Sample collection: MCV, LMS, FI, DJD, MP, LMP, FI, PWJM, CJW, AAB, BO, ML, GD, CW, SS, RKW. Genotyping: MCV, SS, RA. Expression, wet lab: BHJ, TB, HK. Statistical analyses, figures: MCV, BHJ, SCC, FI. Writing: MCV, EAMF, RKW. All authors read, critically revised, and approved the final manuscript.

Competing interests

The authors declare that they have no competing interests.

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**Legends**

**Figure 1 TGF-β + CTGF expression in Human and macrophage polarization of WWOX**

TGF-β is a crucial factor in the inflammation and fibrosis equilibrium.

A, B) TGF-β and CTGF (profibrotic downstream gene of TGF-β) expression in the non-stenotic ileoceanal resection tissue from 5 CD patients carrying the risk allele (AG) and 5 CD patients homozygous for the wild-type allele (GG).

*CD; Crohn’s disease, qPCR; Quantification Polymerase Chain Reaction, Tgf-β; Transforming Growth Factor, CTGF; Connective Tissue Growth Factor; homozyg; homozygous, heterozyg; heterozygous.*
Table 1 Clinical characteristics of patients in the combined cohort.

This table provides the clinical characteristics of patients with recurrent fibrostenotic and purely inflammatory CD in the combined cohort. The disease location and behaviour is based on the Montreal Classification for CD. Chi-squared and Mann-Whitney U-test (only in Age of onset) are used to calculate the p-value. CD; Crohn’s disease

Table 2 Allelic association analysis

The five most significant allelic association analysis results (χ² test) for the 166,251 SNPs in the discovery cohort (242 patients with recurrent fibrostenotic CD vs 279 patients with purely inflammatory CD). Replication was performed for 39 SNPs in a replication cohort (80 patients with recurrent fibrostenotic CD vs 340 patients with purely inflammatory CD). Meta-analysis was performed in the combined cohort.

CD; Crohn’s disease, SNP; Single Nucleotide Polymorphism


Table 1 Clinical characteristics of patients in the combined cohort

<table>
<thead>
<tr>
<th></th>
<th>Fibrostenotic CD</th>
<th>Purely inflammatory CD</th>
<th>P-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>n (%)</td>
<td>322 (100%)</td>
<td>619 (100%)</td>
<td></td>
</tr>
<tr>
<td><strong>Disease Characteristics</strong></td>
<td></td>
<td></td>
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<tr>
<td>Female, n (%)</td>
<td></td>
<td></td>
<td></td>
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<tr>
<td></td>
<td>180 (56%)</td>
<td>371 (60%)</td>
<td>0.23</td>
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<tr>
<td>Age of onset median yrs. (IQR 25-75)</td>
<td></td>
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<tr>
<td></td>
<td>24 (18-30)</td>
<td>26 (19-36)</td>
<td>0.38</td>
</tr>
<tr>
<td><strong>Disease Location, n (%)</strong></td>
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<tr>
<td>Montreal-L1, ileal</td>
<td>86 (27%)</td>
<td>93 (15%)</td>
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<td>36 (11%)</td>
<td>291 (47%)</td>
<td>&lt;0.0001</td>
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<td>Montreal-L3 ileocolonic</td>
<td>200 (62%)</td>
<td>235 (38%)</td>
<td>0.0011</td>
</tr>
<tr>
<td>Montreal-L4, additional upper disease localisation</td>
<td>32 (10%)</td>
<td>55 (9%)</td>
<td>1</td>
</tr>
<tr>
<td><strong>Disease Behaviour, n (%)</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Montreal-B1, inflammatory</td>
<td></td>
<td></td>
<td>0</td>
</tr>
<tr>
<td>Montreal-B2, stricturing</td>
<td>165 (51%)</td>
<td>0</td>
<td></td>
</tr>
<tr>
<td>Montreal-B3, penetrating</td>
<td>157 (49%)</td>
<td>0</td>
<td></td>
</tr>
<tr>
<td><strong>Time until surgery</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>
Disease duration in years from diagnosis until first surgery, mean (SD) | 4.1 (6.5)

This table provides the clinical characteristics of patients with recurrent fibrostenotic and purely inflammatory CD in the combined cohort. The disease location and behaviour is based on the Montreal Classification for CD. Chi-squared and Mann-Whitney U-test (only in Age of onset) are used to calculate the p-value. N; number, CD; Crohn’s disease
<table>
<thead>
<tr>
<th>Chromosome</th>
<th>SNP</th>
<th>Position (Hg19)</th>
<th>Candidate Gene</th>
<th>Risk Allele</th>
<th>Risk allele Frequency in cases</th>
<th>Risk allele Frequency in controls</th>
<th>P-value</th>
<th>Odds Ratio</th>
<th>Risk allele Frequency in cases</th>
<th>Risk allele Frequency in controls</th>
<th>P-value</th>
<th>Odds Ratio</th>
<th>Odds Ratio</th>
</tr>
</thead>
<tbody>
<tr>
<td>16</td>
<td>rs11861007</td>
<td>79238685</td>
<td>WWOX</td>
<td>A</td>
<td>0.19</td>
<td>0.05</td>
<td>1.26 x 10^{-11}</td>
<td>4.3</td>
<td>0.13</td>
<td>0.07</td>
<td>0.01</td>
<td>2.1</td>
<td>6.09 x 10^{-11}</td>
</tr>
<tr>
<td>22</td>
<td>rs371513</td>
<td>21988599</td>
<td>CCDC116</td>
<td>A</td>
<td>0.22</td>
<td>0.13</td>
<td>1.02 x 10^{-04}</td>
<td>1.9</td>
<td>0.20</td>
<td>0.15</td>
<td>1.47 x 10^{-02}</td>
<td>1.4</td>
<td>7.97 x 10^{-05}</td>
</tr>
<tr>
<td>5</td>
<td>rs55965691</td>
<td>593812</td>
<td>CEP72</td>
<td>A</td>
<td>0.008</td>
<td>0.06</td>
<td>1.60 x 10^{-05}</td>
<td>0.14</td>
<td>0.025</td>
<td>0.06</td>
<td>5.97 x 10^{-02}</td>
<td>0.38</td>
<td>9.37 x 10^{-05}</td>
</tr>
<tr>
<td>5</td>
<td>rs6883704</td>
<td>599390</td>
<td>CEP72</td>
<td>C</td>
<td>0.01</td>
<td>0.06</td>
<td>5.04 x 10^{-05}</td>
<td>0.18</td>
<td>0.025</td>
<td>0.06</td>
<td>5.97 x 10^{-02}</td>
<td>0.38</td>
<td>1.01 x 10^{-04}</td>
</tr>
<tr>
<td>3</td>
<td>rs17033143</td>
<td>10630258</td>
<td>ATP2B2</td>
<td>A</td>
<td>0.14</td>
<td>0.07</td>
<td>1.84 x 10^{-04}</td>
<td>2.2</td>
<td>0.09</td>
<td>0.07</td>
<td>3.24 x 10^{-01}</td>
<td>1.36</td>
<td>3.51 x 10^{-04}</td>
</tr>
</tbody>
</table>

The five most significant allelic association analysis results (χ² test) for the 166,251 SNPs in the discovery cohort (242 patients with recurrent fibrostenotic CD vs 279 patients with purely inflammatory CD). Replication was performed for 39 SNPs in a replication cohort (80 patients with recurrent fibrostenotic CD vs 340 patients with purely inflammatory CD). Meta-analysis was performed in the combined cohort.

CD: Crohn’s disease, SNP: Single Nucleotide Polymorphism
Figure 1.