

University of Groningen

Safety and Histological Effect of Liquid Nitrogen Metered Spray Cryotherapy in the Lung

Slebos, Dirk-Jan; Breen, David; Coad, James; Klooster, Karin; Hartman, Jorine; Browning, Robert; Shah, Pallav L.; McNulty, William H.; Mohsin, Mohammed al-Abdul; Irshad, Kashif

Published in:
American Journal of Respiratory and Critical Care Medicine

DOI:
[10.1164/rccm.201611-2220LE](https://doi.org/10.1164/rccm.201611-2220LE)

IMPORTANT NOTE: You are advised to consult the publisher's version (publisher's PDF) if you wish to cite from it. Please check the document version below.

Document Version
Final author's version (accepted by publisher, after peer review)

Publication date:
2017

[Link to publication in University of Groningen/UMCG research database](#)

Citation for published version (APA):

Slebos, D-J., Breen, D., Coad, J., Klooster, K., Hartman, J., Browning, R., ... Irshad, K. (2017). Safety and Histological Effect of Liquid Nitrogen Metered Spray Cryotherapy in the Lung. *American Journal of Respiratory and Critical Care Medicine*, 196(10), 1351-1352. <https://doi.org/10.1164/rccm.201611-2220LE>

Copyright

Other than for strictly personal use, it is not permitted to download or to forward/distribute the text or part of it without the consent of the author(s) and/or copyright holder(s), unless the work is under an open content license (like Creative Commons).

Take-down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

Downloaded from the University of Groningen/UMCG research database (Pure): <http://www.rug.nl/research/portal>. For technical reasons the number of authors shown on this cover page is limited to 10 maximum.

AJRCCM Letter to the Editor - Research letter

Safety and histological effect of liquid nitrogen metered spray cryotherapy in the lung.

Dirk-Jan Slebos MD¹, David Breen MD², James Coad MD³, Karin Klooster PhD¹, Jorine Hartman PhD¹, Robert Browning MD⁴, Pallav L Shah MD⁵, William H McNulty MD⁵, Mohammed al-Abdul Mohsin MD⁶, Kashif Irshad MD⁶

- 1) *Department of Pulmonary Diseases, University of Groningen, University Medical Center Groningen, The Netherlands.*
- 2) *Galway University Hospitals – Galway, Ireland*
- 3) *West Virginia University – Morgantown, West Virginia, United States*
- 4) *Walter Reed National Military Medical Center – Bethesda, Maryland, United States*
- 5) *The National Institute for Health Research Respiratory Biomedical Research Unit, Royal Brompton, London SW3 6NP, UK; Harefield NHS Foundation Trust, London, UK; Imperial College, London, UK*
- 6) *William Osler Health System – Brampton, Canada*

Correspondence: Dr. Dirk-Jan Slebos MD PhD, Dept. of Pulmonary Diseases, AA11, University Medical Center Groningen, PO Box 30001, 9700RB, The Netherlands. d.j.slebos@umcg.nl

Running Head: Safety study of Endobronchial Cryospray

Source of funding: This study was supported by CSA Medical, Inc., Lexington MA.

Keywords: Cryotherapy, liquid nitrogen, bronchoscopy, lung, chronic bronchitis, COPD

Wordcount: 1003

To the Editor,

Chronic bronchitis is characterized by inflammation, cough and increased mucus production. There is no cure, and current treatment options are limited to addressing symptoms [1]. The RejuvenAir® System (CSA Medical, Inc., Lexington MA, USA) is a device designed to bronchoscopically address chronic bronchitis by delivering liquid nitrogen (LN₂) as a Metered Cryospray™ (MCS). MCS is controlled by a thermocouple on the delivery catheter that feeds back real time information regarding the amount of LN₂ being delivered. That amount is tailored such that each bronchial airway receives a standardized amount of LN₂ based on airway size leading to a 10mm circular cryoablation with a depth between 0.1mm to 0.5mm.

Based on older generations LN₂ spray interventions [2,3], it is hypothesized that LN₂ can induce an airway tissue healing effect by destroying the hyperplastic goblet cells and excess submucous glands. The extracellular matrix facilitates rapid regrowth of normal epithelium without scarring, a hallmark of cryoablation [4,5]. The healing resulting from LN₂ MCS is the basis of this proposed treatment.

To start this project, we evaluated the safety of delivering LN₂ in two small pilot studies. Two sprays were delivered into the lobar and first segmental bronchi, which would be the most distal extent of the anticipated airway treatment in chronic bronchitis, based on the significant goblet cell pathology present in these areas [6]. The primary endpoint for each study was the evaluation of device related serious adverse events (SAEs). The initial study (NCT02106143, previously reported as ATS-2016 conference abstract [7]) was conducted in subjects who were

scheduled to undergo lobectomy or pneumonectomy immediately following MCS. The second study (NCT02483052) evaluated the same initial, plus post procedure safety as these subjects had a planned lobectomy after MCS. Both studies had evaluation of potential histologic effects of MCS as a secondary endpoint and the specimens were reviewed by an independent pathologist skilled in cryothermic tissue injuries (JC). Procedures were performed by flexible bronchoscopy under general anesthesia using an endotracheal tube with positive pressure ventilation. During the LN₂ spray, the endotracheal tube was disconnected from the ventilator with the cuff deflated, both to allow N₂ gas egress. Major inclusion criteria included FEV₁ ≥50% of predicted, no bullae >3cm, and no previous lung surgery of any sort. Both studies were approved by the local ethics committees and all patients provided informed consent.

Overall, 16 subjects were enrolled and treated at 3 sites in Europe and Canada. The initial study enrolled 11 (3F/8M) subjects, with a mean age of 65.8±8.9years [range 45.7-74.8years], a mean FEV₁ %predicted of 86.1±28.1% [range 46-132%], and a smoking history of 28.3±20.5 pack-years [range 0.02-54.0]. Mean time to the scheduled surgical resection after MCS was 37.7±21.8 minutes [range 15-80minutes]. In the delayed study, 5 subjects (4F/1M) were enrolled, with a mean age of 66.8±6.2years [range 59.2-74.0years], a mean FEV₁ %predicted of 72.8±16.5% [range 51-91%], and a smoking history of 29.8±22.4 pack-years [range 4.5-47.0]. Mean time to resection after MCS was 14±1.4 days [range 12-16 days]. Final histopathological diagnosis was non-small cell lung cancer in 15, and small cell lung cancer in one patient.

All intended thirty-two LN₂ MCS were given. The delivery of MCS was both feasible and safe with no device related (S)AEs. There were no intra-operative complications with stable vital signs and no technical difficulties preventing application of MCS. Three SAE's were reported in

each study, all determined to be unrelated to the study device, the MCS treatment or bronchoscopy as adjudicated by an independent medical monitor. The SAE's all occurred post completion of MCS and VATS: atrial fibrillation (8 days post), mucus plug (3 days post), death >30 days post LN₂ spray due to hospital acquired pneumonia after pneumonectomy in the non-treated lung, and one subject with an intra-operative bleeding during video assisted thoroscopic surgery (VATS, 14 days post MCS), an immediate post-operative bleeding, and again 18 days after VATS.

Eight immediate resection subjects treated bronchial segments were available, with in five of them revealing presence of the intended cryothermic histologic changes. In the regions with greater epithelial changes, minimal to mild acute inflammation was present. In two specimens there was extension into the submucosa with cryothermic changes involving the submucosal glands with preservation of the underlying submucosal structure including no effect on underlying cartilage (figure 1).

In the delayed resection study, a bronchoscopy was performed just before lung resection took place. The video of these bronchoscopies were independently reviewed by an experienced bronchoscopist (RB), with three subjects showing a slight mucosal whitening, one of which seemed to represent subtle white rings consistent with cryoablation. Airway caliber was normal and there were no signs of infection. All five subjects had bronchial segments submitted for histopathological evaluation. Due to the lack of visual mucosal changes post fixation, identification of the treatment sites represented a major limitation in the subsequent histologic evaluation. Histology findings were consistent with complete re-epithelialization at the treatment site. No persistent treatment-related epithelial erosions, acute inflammation,

granulomatous inflammation, mucosal necrosis, fibroproliferative healing or collagenous luminal narrowing were identified (see figure 2 for a representative example). The bronchial cartilage rings primarily appeared unremarkable with one specimen showing focal cartilage ring necrosis, consistent with possible cryothermic injury or non-specific degenerative changes.

In summary, the data obtained in both clinical trials met the primary and secondary endpoints with safe delivery of MCS to the segmental and/or lobar bronchi without device related serious adverse events. Histology from immediate resection specimens documented MCS effect, and non-scarring healed tissue from the delayed resection group.

The information learned from this study is important for the future development of a bronchoscopic treatment for patients who suffer from chronic bronchitis. While the future intended indication was not specifically studied in these patients, they share some chronic bronchitis characteristics such as exposure to cigarettes, and most having COPD. Using MCS in a similar population and determining the immediate and delayed healing response has provided important information in the stepwise clinical development of the system. These results warrant further investigation in a larger clinical trial targeting chronic bronchitis patients, which is currently underway (NCT02483637).

References

1. Kim V, Criner GJ. Chronic bronchitis and chronic obstructive pulmonary disease. *Am J Respir Crit Care Med*. 2013;187:228-37.
2. Au JT, Carson J, Monette S, Finley DJ. Spray cryotherapy is effective for bronchoscopic, endoscopic and open ablation of thoracic tissues. *Interact Cardiovasc Thorac Surg*. 2012;15:580-4.
3. Krinsky WS, Broussard JN, Sarkar SA, Harley DP. Bronchoscopic spray cryotherapy: assessment of safety and depth of airway injury. *J Thorac Cardiovasc Surg*. 2010;139:781-2.
4. Coad JE, Bischof JC. Histologic differences between cryothermic and hyperthermic therapies. *Proc. of SPIE* 2003;4954:27-36.
5. Godwin BL, Coad JE. Healing Responses Following Cryothermic and Hyperthermic Tissue Ablation. *Proc. of SPIE* 2009;7181:1-9.
6. Mullen JB, Wright JL, Wiggs BR, Pare PD, Hogg JC. Reassessment of inflammation of airways in chronic bronchitis. *Br Med J*. 1985;291:1235-9.
7. Breen D, Coad J, Slebos DJ. A Prospective Study Of Rejuvenair System Radial Spray Cryotherapy To Determine Safety And Histological Effect In The Lung. *Am J Respir Crit Care Med* 193;2016:A6884.

Legend to the figures

Figure 1

Representative 2 hour post liquid nitrogen metered cryospray treated bronchial micrograph demonstrating surface and superficial ductal epithelial removal to a depth up to 500 microns (Hematoxylin and eosin stained, 400x magnification). The submucosal extracellular matrix remains intact (non-denatured) with mild capillary congestion and minimal luminal-oriented acute inflammation and edema.

Figure 2

Representative 14 days post liquid nitrogen metered cryospray treated bronchial micrograph demonstrating complete rejuvenative healing characterized by pseudostratified respiratory epithelium with occasional goblet cells, preservation of the submucosa and cartilage, and absence of inflammation and fibrosis (Hematoxylin and eosin stained, 400x magnification).

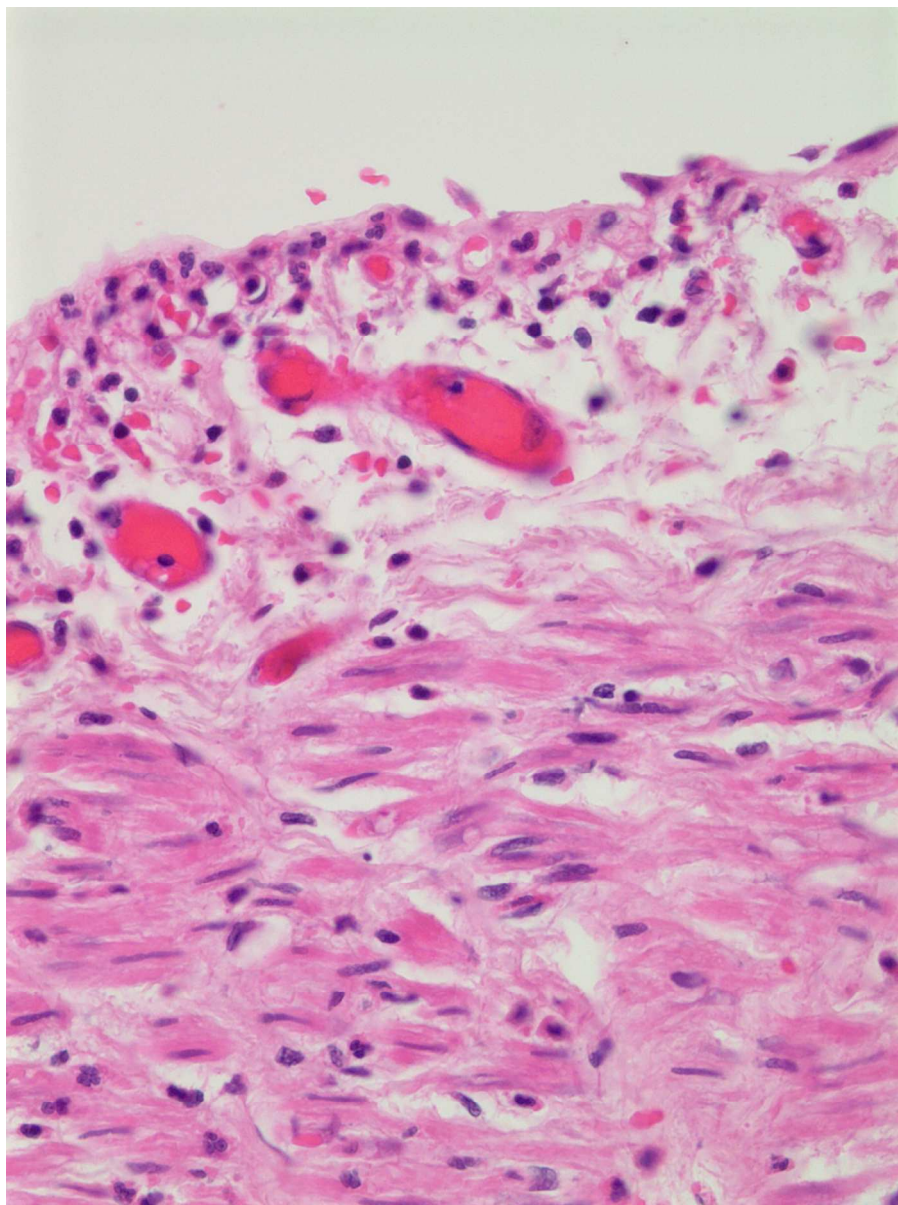


Figure 1

Representative 2 hour post liquid nitrogen metered cryospray treated bronchial micrograph demonstrating surface and superficial ductal epithelial removal to a depth up to 500 microns (Hematoxylin and eosin stained, 400x magnification). The submucosal extracellular matrix remains intact (non-denatured) with mild capillary congestion and minimal luminal-oriented acute inflammation and edema.

133x177mm (300 x 300 DPI)

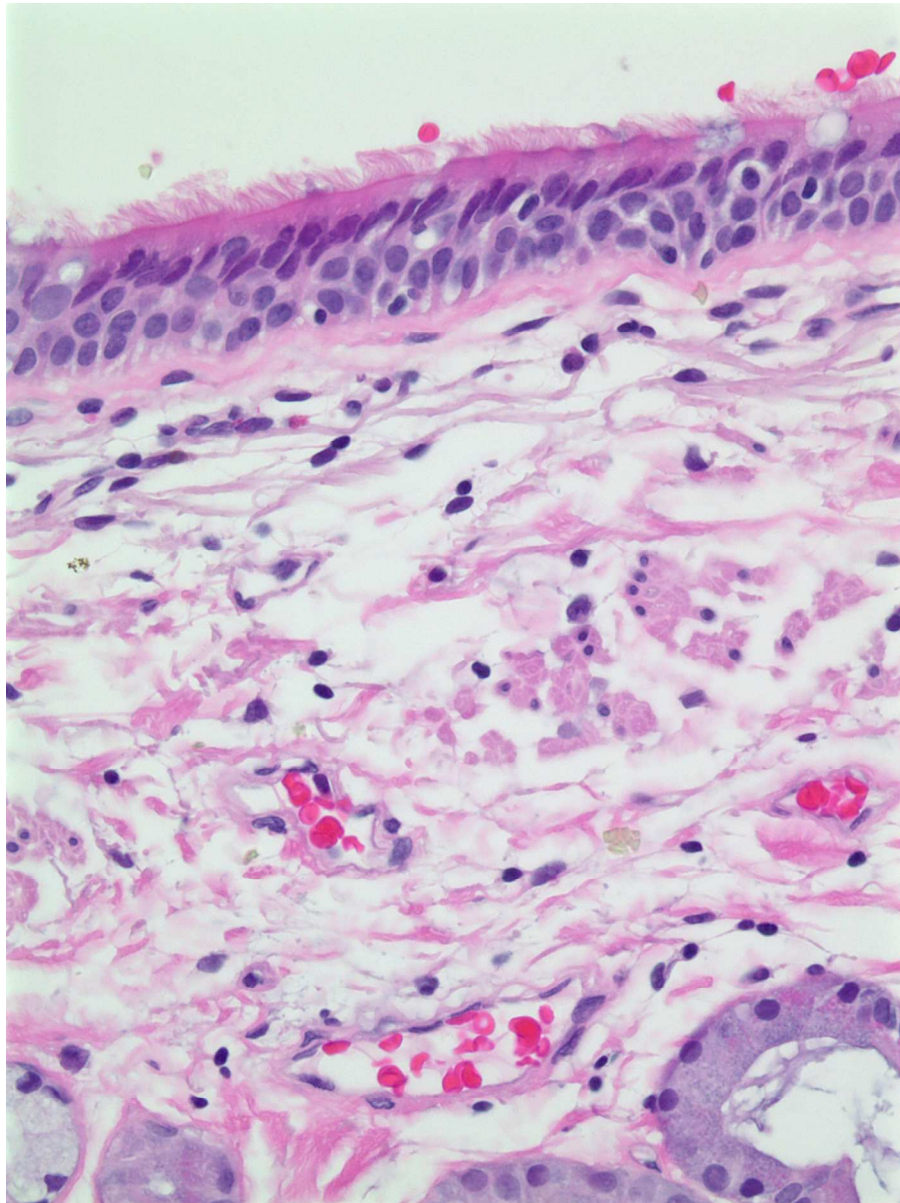


Figure 2
Representative 14 days post liquid nitrogen metered cryospray treated bronchial micrograph demonstrating complete rejuvenative healing characterized by pseudostratified respiratory epithelium with occasional goblet cells, preservation of the submucosa and cartilage, and absence of inflammation and fibrosis (Hematoxylin and eosin stained, 400x magnification).

133x177mm (300 x 300 DPI)