Microorganisms produce extracellular compounds that affect the final product quality in fermentation processes. Selection of overproducing mutants requires coupling of the extracellular product to the producer genotype, which can be achieved by single-cell compartmentalization. Emulsions contain up to billions of microdroplets/mL which significantly increases the screening throughput compared to microtiter-plate-based selections. Factors affecting the success of screening in microdroplets include the nature of the producing organism (robust, no invasive growth), the product (not soluble in oil) and the product assay (preferably fluorescence based). Together these factors determine the required microdroplet production technique and sorting set-up. Because microdroplets allow relatively inexpensive ultrahigh-throughput screening, they are likely to become a standard tool in the strain selection toolbox of the fermentation industry.

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Production of microdroplets
Microdroplets usually have volumes of 65 fl to 65 nL (diameter of 5–500 μm) [5,8,9,10] and they generally consist of a water-phase that contains the content of the microdroplet (e.g. nutrients, cells, assay reagents, hydrogel bead polymers), an oil-phase that can prevent cross-talk between compartments, and a surfactant that localizes at the water/oil interphase and stabilizes the microdroplet by reducing surface tension (Figure 1b). There are different types of microdroplets: water-in-oil (w/o) emulsions, water-in-oil-in-water (w/o/w) double emulsions and hydrogel beads, and all of them can be either monodisperse or polydisperse (Figure 1c).

Water-in-oil emulsions are generated by mixing an oil-phase containing oil-soluble surfactant and a water-phase. Mixing with a vortex shaker yields polydisperse microdroplets, while the use of microfluidic devices allows to produce monodisperse microdroplets (Figure 1c) [8]. Water-in-oil emulsions can be transformed into w/o/w double emulsions by re-emulsification of the primary w/o emulsion in a water-phase with water-soluble surfactant [6]. In both w/o and w/o/w double emulsions the microdroplets are surrounded by oil, which prevents cross-talk between compartments for compounds not soluble in the oil-phase. In this way every
Figure 1

(a) Micro-compartmentalization concept

No compartmentalization
- different mutants share a single compartment
- one batch culture
- mutants affect each other

Compartmentalization in microdroplets
- single mutants separated in micro-compartments surrounded by oil
- each compartment is a single batch culture
- mutants do not affect each other

(b) Key elements of microdroplet production

<table>
<thead>
<tr>
<th>water-phase supplemented with e.g.:</th>
<th>oil</th>
</tr>
</thead>
<tbody>
<tr>
<td>• nutrients</td>
<td>surfactant layer</td>
</tr>
<tr>
<td>• cells</td>
<td>gelation bath</td>
</tr>
<tr>
<td>• polymers for hydrogel beads (e.g. agarose, alginate)</td>
<td>cell</td>
</tr>
<tr>
<td>• assay reagents</td>
<td>microdroplet</td>
</tr>
<tr>
<td></td>
<td>surfactant</td>
</tr>
<tr>
<td></td>
<td>microfluidic device</td>
</tr>
</tbody>
</table>

5 – 500 µm
(65 fL – 65 nL)

(c) Microdroplet production techniques

<table>
<thead>
<tr>
<th>polydisperse</th>
<th>monodisperse</th>
</tr>
</thead>
<tbody>
<tr>
<td>Types</td>
<td>Types</td>
</tr>
<tr>
<td>• water in oil</td>
<td>• hydrogel beads in medium*</td>
</tr>
<tr>
<td>• hydrogel beads in oil*</td>
<td></td>
</tr>
<tr>
<td>Production rate: 10^6–10^10/h</td>
<td>Production rate: 10^9/h</td>
</tr>
</tbody>
</table>

* hydrogel beads can be incubated surrounded by oil (mutants do not affect each other) or surrounded by medium lacking oil (mutants affect each other).

Microdroplet types and their production techniques.
inoculated microdroplet acts as a single, monoclonal batch culture (Figure 1a).

Hydrogel beads are generated by adding polymers such as agarose or alginate to the water-phase of polydisperse or monodisperse w/o emulsions following gelation [11,12], or by dripping a solution containing polymers in a gelation bath [9,13]. Next to incubation surrounded by oil, hydrogel beads can be incubated in medium. In that case hydrogel beads (or more complex beads featuring a surrounding shell architecture) resemble ‘semi-open’ vessels and can be engineered to specific molecular-weight cutoffs ranging from the low kD-range to pores almost of micrometer size [14].

The advantage of polydisperse microdroplets is that the required set-up is inexpensive, easy to use and the microdroplet production rate is high (10⁹–10¹⁰/h). Their disadvantage is that compartment volumes differ. Depending on whether cells do or do not grow in microdroplets, this leads to differences in either the absolute product amount or the product concentration in microdroplets respectively. When correction for microdroplet-volume is not possible, it is therefore hard to catch variants with only moderate levels of improvement. In those cases the use of monodisperse microdroplets is a powerful alternative, because they typically have less than 3% volume variation. However, compared to polydisperse microdroplets their production rate is lower (10⁵–10⁶/h) and a more advanced set-up is required [8].

Detailed protocols for the production of monodisperse and polydisperse microdroplets can be found in [8,15], and [5,16,17] contain information about advanced microfluidic emulsion processing.

Encapsulation of cells in microdroplets
To couple the product concentration to its producer genotype, encapsulation of a single cell-variant/genotype per microdroplet is required (Figure 1a). This encapsulation follows a Poisson distribution [8]. When the number of added cells is for example ten times lower than the number of generated microdroplets, 9% of the generated microdroplets contain a single cell, and less than 0.5% contain multiple cell-variants (Poisson distribution, λ = 0.1). When several rounds of enrichment are possible and variant libraries are huge, higher encapsulation ratios are often feasible. The λ parameter can also be adjusted to allow more than one cell-variant per microdroplet, for instance to co-localize producer and sensor cells [10,18,19*]. Special microfluidic devices can increase the percentage of microdroplets with single cells to 70–100% by aligning cells before microdroplet formation or post-encapsulation sorting [5,20], but good mixing of the inlet streams is required, especially for hydrogel beads [11]. A recent review that summarizes and discusses different encapsulation and cell-alignment techniques can be found in [20].

Strain selections using microdroplets
Organisms
Factors affecting the success of screening in microdrolpets include the producing organism, the product of interest, the product assay and the sorting set-up (Figure 2).

Micro-organisms can be cultured aerobically or anaerobically in microdroplets [32,33]. Successful strain selections in microdroplets include Escherichia coli [19*,23,28], Saccharomyces cerevisiae [12,23,26], Bacillus subtilis [13], Bacillus coagulans [29], Lactococcus lactis [1,9*,31], Yarrowia lipolytica [3*], cyanobacteria [21,24,25] and algae [21,27]. As microdroplets are typically inoculated with a single cell there is no competition between genotypes within a droplet. The platform is therefore also well-suited for slow growing micro-organisms. When cells clump, encapsulation of a single cell per microdroplet is not possible. If clumps consist of multiple genotypes selection is in theory possible, even though the selection efficiency will be reduced because the final product-concentration depends on a combination of genotypes. However, we are not aware of examples of successful selections in such a system. Mammalian cells and filamentous organisms can also be encapsulated and sorted, but they require microdroplets in the nL volume range which reduces the throughput [34,35]. For filamentous organisms apical growth of hyphae should be prevented, to avoid uncontrolled microdroplet coalescence [35]. When this is not possible, mutants can be selected using alternative approaches such as conventional microtiter-plate set-ups.

Detection of relevant phenotypes often requires growth in microdroplets. Best et al. for instance grew cyanobacteria and algae in microdroplets to select mutants that reached increased cell densities [21]. The minimal required microdroplet volume is determined by the desired number of generations per microdroplet and the final cell concentration in the medium (Figure 2a). Microalgae could for instance grow for 11 generations in 2 nL microdroplets, reaching a final cell concentration of 10⁹/mL [27].

Hydrogel beads can also be taken up in growth medium before they are incubated surrounded by oil. This allows uncoupling of growth, product formation and product detection, making it a versatile selection system. Schmitt et al. for example grew single L. lactis cells in hydrogel beads taken up in growth medium. After micro-colony formation they induced the expression of lantibiotic production genes, re-emulsified the hydrogel beads in oil to prevent cross-talk between microdroplets and assessed the lantibiotic effectivity using co-localized sensor cells [9*].
Microdroplet screening for extracellular compounds

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Figure 2

Decision tree for strain selection in microdroplets.

(a) Which organism(s)?

- **Fit for microdroplet cultivation**
  - E. coli, B. subtilis, B. coagulans, L. lactis, S. cerevisiae, Y. lipolytica, cyanobacteria, algae, filamentous fungi
- **Unfit for microdroplet cultivation**
  - Large, multicellular organisms

Growth required?

- **no**
  - Minimal droplet volume depends on cell-size
- **yes**
  - Minimal droplet volume \( V_d \) depends on number of generations \( \text{gen} \) and final cell density \( [\text{cells}] \): \( V_d = 2^{\text{gen}} / [\text{cells}] \)

(b) Which extracellular compound?

- **Compound of interest is not soluble in oil**
  - E.g. charged molecules, sugars, enzymes
- **Compound of interest is partly oil-soluble**
  - E.g. hydrophobic compound, small and uncharged molecules, flavor volatiles

(c) Which assay?

- **Signal: fluorescence**
  - Is the compound of interest highly fluorescent?
    - **yes**
      - Chemical conversion
      - Enzymatic conversion
      - Biosensors
    - **no**
      - Cross-feeding
- **Signal: cell concentration**
  - Is the compound of interest biomass?
    - **yes**
      - Challenge
    - **no**

(d) Which compartmentalization method and sorting set-up?

- **Fluorescence based sorting**
  - On chip
    - w/o emulsion
  - Particle sorter*
    - w/o/w emulsion
    - hydrogel beads (*only when the signal is captured during sorting*)
  - FACS when droplet volume <35 pL, large particle sorter when droplet volume >35 pL
- **Cell number based sorting**
  - Serial propagation
    - w/o emulsion
- **Weight based sorting**
  - On chip
    - w/o emulsion
- **Scatter based sorting**
  - On chip
    - w/o emulsion
    - Particle sorter*
    - w/o/w emulsion
    - hydrogel beads

*FACS when droplet volume <35 pL, large particle sorter when droplet volume >35 pL
### Table 1
Overview recent literature on strain selections using microdroplets

<table>
<thead>
<tr>
<th>Producer organism</th>
<th>Droplet characteristics</th>
<th>Product</th>
<th>Assay</th>
<th>Selection</th>
<th>Ref</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>On-chip sorting</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>L. lactis</em></td>
<td>w/o emulsion</td>
<td>Monodisperse 50</td>
<td>Riboflavin</td>
<td>Riboflavin auto-fluorescence</td>
<td>On-chip sorting 2.10^5/h</td>
</tr>
<tr>
<td>Cyanobacteria</td>
<td>w/o emulsion</td>
<td>Monodisperse 65</td>
<td>Biomass (chlorophyll fluorescence)</td>
<td>Chlorophyll auto-fluorescence</td>
<td>On-chip sorting 1.10^6/h</td>
</tr>
<tr>
<td>Algae</td>
<td>w/o emulsion</td>
<td>Monodisperse 34</td>
<td>Biomass (chlorophyll fluorescence)</td>
<td>Chlorophyll auto-fluorescence</td>
<td>On-chip sorting 1.10^6/h</td>
</tr>
<tr>
<td><em>S. cerevisiae</em></td>
<td>w/o emulsion</td>
<td>Monodisperse 34</td>
<td>Tyrosine</td>
<td>Fluorophore binding aptamer</td>
<td>Enzyme assay pyranose oxidase coupled to a H_2O_2-dependent fluorogenic conversion</td>
</tr>
<tr>
<td><em>S. cerevisiae</em></td>
<td>w/o emulsion</td>
<td>Monodisperse 221</td>
<td>Consumption of xylose</td>
<td>On-chip sorting 1.10^4/h</td>
<td>[23]</td>
</tr>
<tr>
<td><em>E. coli</em></td>
<td>w/o emulsion</td>
<td>Monodisperse 221</td>
<td>Lactate</td>
<td>Enzyme assay lactate oxidase coupled to a H_2O_2-dependent fluorogenic conversion</td>
<td>On-chip sorting 1.10^5/h</td>
</tr>
<tr>
<td>Cyanobacteria</td>
<td>w/o emulsion</td>
<td>Monodisperse 524</td>
<td>Ethanol</td>
<td>Enzyme assay ethanol oxidase coupled to a H_2O_2-dependent fluorogenic conversion</td>
<td>On-chip sorting 4.10^5/h</td>
</tr>
<tr>
<td>Cyanobacteria</td>
<td>w/o emulsion</td>
<td>Monodisperse 10</td>
<td>Lactate</td>
<td>Enzyme assay lactate dehydrogenase coupled to a NADH-dependent fluorogenic conversion</td>
<td>On-chip sorting 4.10^5/h</td>
</tr>
<tr>
<td><em>E. coli</em></td>
<td>w/o emulsion</td>
<td>Monodisperse 87/1,023</td>
<td>2-ketoisovalerate</td>
<td>Biosensor</td>
<td>On-chip sorting 1.10^6/h</td>
</tr>
<tr>
<td><em>S. cerevisiae</em></td>
<td>w/o emulsion</td>
<td>Monodisperse 22</td>
<td>p-Coumaric acid</td>
<td>Biosensor</td>
<td>On-chip sorting 1.10^6/h</td>
</tr>
<tr>
<td>Algae</td>
<td>w/o emulsion</td>
<td>Monodisperse 2,145</td>
<td>Biomass (droplet weight)</td>
<td>Magnetic separation based on microdroplet weight</td>
<td>On-chip sorting –</td>
</tr>
<tr>
<td><strong>Particle sorters</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>E. coli</em></td>
<td>agarose beads in oil</td>
<td>Monodisperse 65</td>
<td>pBAD promoters with desired characteristics</td>
<td>Promotor-dependent fluorescence</td>
<td>FACS –</td>
</tr>
<tr>
<td><em>Y. lipolytica</em></td>
<td>w/o/w emulsion</td>
<td>Monodisperse 34</td>
<td>Riboflavin</td>
<td>Riboflavin auto-fluorescence</td>
<td>FACS</td>
</tr>
<tr>
<td><em>B. coagulans</em></td>
<td>w/o/w emulsion</td>
<td>Monodisperse 12</td>
<td>Lactate</td>
<td>Fluorescent pH indicator</td>
<td>FACS</td>
</tr>
<tr>
<td>Siberian bear oral microbial community</td>
<td>w/o/w emulsion</td>
<td>Monodisperse 4</td>
<td>Antimicrobials against <em>S. aureus</em></td>
<td>Biosensor</td>
<td>FACS</td>
</tr>
<tr>
<td><em>S. cerevisiae</em></td>
<td>agarose beads in medium</td>
<td>Polydisperse 22</td>
<td>Antimicrobials against <em>S. aureus</em></td>
<td>Biosensor</td>
<td>FACS</td>
</tr>
</tbody>
</table>
### Microdroplet screening for extracellular compounds

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### Compounds

In microdroplets surrounded by oil, the oil-phase prevents diffusion of hydrophilic compounds (e.g. charged molecules, sugars, (poly-)peptides) between compartments, and therefore ensures coupling of the product concentration to its producer (Figure 1a). However, hydrophobic compounds leak into the oil-phase and therefore are not compartmentalized. One might circumvent this limitation by rapid enzymatic conversion of the hydrophobic product into a hydrophilic compound [24]. When assay reagents are oil-soluble, chemical modification can reduce their hydrophobicity to allow compartmentalization [36].

### Product assays and microdroplet sorting

Assays for the produced extracellular compounds aim to couple the product-concentration to a measurable signal (Figure 2c). Developing these assays is often the most demanding task in a screening campaign. Assays that require direct analysis on, for example, an HPLC, MS, NMR or GC setup are less suited for the microdroplet format. Although compound libraries have been screened in microdroplets using MALDI-TOF MS [37,38], the method has a low-throughput. However, microdroplet-based screening campaigns are well suited for a wide range of spectroscopy methods (Table 1).

For w/o and w/o/w emulsions it is recommended to use assays with reagents that are not oil-soluble, and that can be added to the water-phase during microdroplet production. The addition of reagents after microdroplet generation and incubation is possible [23–25], but droplet fusion or liquid injection requires specialized equipment and know-how. Hydrogel beads are more flexible in this regard, because they can also be surrounded by a water-phase. This allows for instance viability staining of biosensors after incubation [12]. Once a measurable signal is obtained, microdroplets can be sorted either on-chip or with particle sorters (Figure 2d). For on-chip sorting generally w/o emulsions are used (Table 1). Particle sorters use water as carrier phase and therefore they can sort w/o/w double emulsions or hydrogel beads taken up in a water-phase (Table 1).

The required product assay, sorting set-up and compartmentalization method are highly intertwined (Figure 2c and d). Assay read-outs that are regularly used to select microdroplets are fluorescence emission and cell-concentrations.

#### Fluorescence

Direct selection for fluorescence is only possible when the product itself is highly fluorescent. Auto-fluorescence of riboflavin was for instance used to select *L. lactis* mutants producing four times more riboflavin [1]. When the compound of interest is not highly fluorescent, (bio)chemicals, enzymes or biosensors can couple the presence of the

---

### Table 1 (Continued)

<table>
<thead>
<tr>
<th>Producer organism</th>
<th>Droplet characteristics</th>
<th>Volume (pL)</th>
<th>Throughput</th>
<th>Assay</th>
<th>Selection</th>
<th>Ref</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>E. coli</em></td>
<td>agarose beads in oil</td>
<td>22</td>
<td>5 · 10⁷/h</td>
<td>Biosensor</td>
<td>S. aureus viability staining</td>
<td>[13]</td>
</tr>
<tr>
<td><em>B. subtilis</em></td>
<td>alginate beads</td>
<td>65,000</td>
<td>6 · 10⁷/h</td>
<td>Riboflavin</td>
<td>Biosensor</td>
<td>[14]</td>
</tr>
<tr>
<td><em>M. flavus</em></td>
<td>alginate beads</td>
<td>50,000</td>
<td>6 · 10⁷/h</td>
<td>Effective lantibiotics</td>
<td>Biosensor</td>
<td>[15]</td>
</tr>
<tr>
<td><em>L. lactis</em></td>
<td>alginate beads</td>
<td>42</td>
<td>NA</td>
<td>Biosensor</td>
<td>L. lactis growth based fluorescence using DNA staining or L. lactis, growth based fluorescence using GFP expression</td>
<td>[26]</td>
</tr>
</tbody>
</table>

---

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product to a fluorescence signal. Zhu et al. for instance used a chemical fluorescent pH indicator to select lactate producing B. coagulans strains [29]. Biochemicals which can be used are for instance fluorophore-binding aptamers, which yield a fluorescence signal when both a dye and the target molecule are bound [39]. Abatemarco et al. used these aptamers to select S. cerevisiae mutants that secreted 28-fold more tyrosine compared to their wildtype ancestor [22*].

Enzymes can also couple the presence of a compound to a fluorescence signal [23,24,36]. Hammar et al. connected lactate production of cyanobacteria to a fluorescence signal by coupling NADH production via lactate dehydrogenase combined to an NADH-dependent conversion of a fluorogenic substrate [25]. When the compound of interest is an enzyme itself, the addition of fluorogenic substrates can couple the enzyme activity to fluorescence [15,39–41].

Lastly whole-cell biosensors can also couple the presence of an extracellular product to a fluorescence signal. In general genetically accessible strains are used (Table 1), as biosensor development often requires genetic engineering. Biosensors are relatively cheap, since a high number of cells can be obtained in low-cost media, whereas other conversion systems (e.g. chemical or enzymatic) require tedious purification processes and analytical instruments such as chromatography and spectrophotometry [42,43].

We here classify biosensors in three types (Figure 3). Type 1 biosensors harbor reporter genes that respond to a

---

**Figure 3**

<table>
<thead>
<tr>
<th>Type 1. Biosensor contains sensing element for product (e.g. responsive promotor) coupled to a reporter gene (e.g. gfp)</th>
</tr>
</thead>
<tbody>
<tr>
<td>compound of interest</td>
</tr>
<tr>
<td>responsive promotor</td>
</tr>
<tr>
<td>DNA</td>
</tr>
<tr>
<td>gfp</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Type 2. Biosensor is constitutively fluorescent and only grows in presence of product (auxotrophy)</th>
</tr>
</thead>
<tbody>
<tr>
<td>compound of interest</td>
</tr>
<tr>
<td>→ required by biosensor to grow</td>
</tr>
<tr>
<td>constitutive promotor</td>
</tr>
<tr>
<td>DNA</td>
</tr>
<tr>
<td>gfp</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Type 3. Biosensor is killed by product and its lysis causes loss of fluorescence or dead cells are stained</th>
</tr>
</thead>
<tbody>
<tr>
<td>compound of interest</td>
</tr>
<tr>
<td>→ kills the biosensor</td>
</tr>
<tr>
<td>cell death/lysis</td>
</tr>
</tbody>
</table>

Biosensor types.
Mutants with a high cell concentration can be enriched by serial propagation in microdroplets. Bachmann et al. for instance used this method to enrich *L. lactis* mutants that produced 71% more cells which coincided with a 26% higher biomass yield [31]. Next to serial propagation, microdroplets or hydrogel beads with increased cell concentrations can also be selected by sorting based on increased scattering of light [46], or weight-based magnetic sorting on a microfluidic device [27].

**Conclusions**

Screening in microdroplets allows selection for increased microbial production of extracellular compounds relevant to the (food) fermentation industry (Table 1). Multiple microdroplet production techniques are available (Figure 1c). They can be combined with different assays to generate fluorescence and cell concentration-based signals, which can be sorted in various ways (Figure 2). The optimal combination of production technique, assay and sorting system differs per screening question (Figure 2).

The applicability of microdroplets for screening is limited by the availability of suitable assays for the compound of interest. Current assays often require substantial tuning before they can be used in microdroplets, which is time-consuming [19**,36,39]. The most generic set-ups that are currently available use metabolite oxidases or dehydrogenases coupled to the generation of fluorescence [23–25], or viability staining of biosensors [9**,12,30]. Future research could focus on high-throughput screening for targeted aptamers [47] and (automated) biosensor development with a focus on enhanced sensitivity and specificity, increased dynamic ranges and improved transfer of sensing elements between organisms [39,48]. For filamentous fungi, future studies need to focus on increasing droplet stability, to prevent coalescence caused by apical growth of hyphae.

Screening in microdroplets is furthermore limited by the lack of a set-up to select for oil-soluble compounds. This is especially relevant for the food industry, because flavor volatiles are often oil-soluble. Possible solutions could focus on capturing and measuring oil-soluble compounds in microdroplets, for instance by measuring concentration gradients rather than absolute concentrations [49], or by capturing oil-soluble compounds in microdroplets by chemical or enzymatic conversion/modification [36].

Overall the recent technological progress on microdroplet production and sorting brings this technology within reach for strain selection in the (food) fermentation industry. While there are still a number of challenges, it has a high potential for (non GMO) strain improvement and due to its high throughput it should allow to select for phenotypes that are not accessible with conventional screening methods.
Conflict of interest statement
H.B. is employed by NIZO Food Research, a contract research organization. NIZO Food Research had no role in the decision to publish or the preparation of the manuscript.


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References and recommended reading
 Papers of particular interest, published within the period of review, have been highlighted as:

- of special interest
- of outstanding interest


This study highlights the difference in outcome when selecting for intra- or extracellular product concentration.


The authors selected producers of effective antibiotics by uncoupling producer growth from product formation. Their work demonstrates the versatility of hydrogel beads as selection system [22].


The authors use cross-feeding between micro-organisms within one microdroplet to create a positive feedback loop, in which overproducers reach increased cell concentrations.


This study is an example of the use of fluorophore-binding aptamers to couple the product concentration to a fluorescence signal. The authors used this system to select tyrosine overproducing S. cerevisiae strains.


This study demonstrates the use of multicolor fluorescence detection on microfluidic devices, which allows sorting based on multiple fluorescence signals.


